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Editorial Board Member of World Journal of Hepatology, Igor Skrypnyk, MD, MDS, PhD, Professor, Internal Medicine #1, Poltava State Medical University, Poltava 36011, Ukraine. inskrypnyk@gmail.com

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EDITORIAL

## Roles of transforming growth factor-β signaling in liver disease

Xiao-Ling Wang, Meng Yang, Ying Wang

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Xiao-Ling Wang, Meng Yang, Ying Wang, Clinical Laboratory, Shanxi Academy of Traditional Chinese Medicine, Taiyuan 030012, Shanxi Province, China

Corresponding author: Xiao-Ling Wang, PhD, Chief Technician, Clinical Laboratory, Shanxi Academy of Traditional Chinese Medicine, No. 46 Bingzhou West Street, Yingze District, Taiyuan 030012, Shanxi Province, China. wangxiaoling24@163.com

### Abstract

In this editorial we expand the discussion on the article by Zhang et al published in the recent issue of the World Journal of Hepatology. We focus on the diagnostic and therapeutic targets identified on the basis of the current understanding of the molecular mechanisms of liver disease. Transforming growth factor- $\beta$  (TGF- $\beta$ ) belongs to a structurally related cytokine super family. The family members display different time- and tissue-specific expression patterns associated with autoimmunity, inflammation, fibrosis, and tumorigenesis; and, they participate in the pathogenesis of many diseases. TGF- $\beta$  and its related signaling pathways have been shown to participate in the progression of liver diseases, such as injury, inflammation, fibrosis, cirrhosis, and cancer. The often studied TGF- $\beta$ /Smad signaling pathway has been shown to promote or inhibit liver fibrosis under different circumstances. Similarly, the early immature TGF- $\beta$  molecule functions as a tumor suppressor, inducing apoptosis; but, its interaction with the mitogenic molecule epidermal growth factor alters this effect, activating anti-apoptotic signals that promote liver cancer development. Overall, TGF-β signaling displays contradictory effects in different liver disease stages. Therefore, the use of TGF- $\beta$ and related signaling pathway molecules for diagnosis and treatment of liver diseases remains a challenge and needs further study. In this editorial, we aim to review the evidence for the use of TGF- $\beta$  signaling pathway molecules as diagnostic or therapeutic targets for different liver disease stages.

Key Words: Transforming growth factor-β signaling; Liver disease; Molecular mechanism; Targets; Diagnosis

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**Core Tip:** Transforming growth factor- $\beta$  (TGF- $\beta$ ) participates in diverse signaling pathways and exhibits contradictory roles in liver immunity, inflammation, repair, fibrosis, and other biological processes. In this editorial, we review the evidence for potential TGF- $\beta$  signaling pathway molecules as diag-nostic or therapeutic targets for different liver disease stages.

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### INTRODUCTION

Transforming growth factor- $\beta$  (TGF- $\beta$ ) is a superfamily of cytokines with 33 member proteins, including TGF- $\beta$  isoforms (TGF- $\beta$ 1/2/3), bone morphogenetic protein (BMP), growth and differentiation factor (GDF), activin, nodal, inhibin, and AMH (anti-Mullerian hormone). These proteins exhibit time- and tissue-specific expression patterns (Figure 1), and they participate in the development of pathological processes such as autoimmunity, inflammation, fibrosis, and tumori-genesis. In different liver diseases, TGF- $\beta$  and its related signaling pathways regulate the progression from inflammation to fibrosis, to cirrhosis, and to liver cancer[1]. TGF- $\beta$ 1, TGF- $\beta$ 2, and TGF- $\beta$ 3-mediated signals are essential for liver homeostasis and repair, and they are key immune system regulators[2]. TGF- $\beta$ /SMAD is a classical signaling pathway with effects that can both promote and inhibit liver fibrosis[3]. Immature TGF- $\beta$  signaling affects different stages of liver disease progression, and the study of TGF- $\beta$  and its signaling pathways to improve liver disease diagnoses and treatments remains an active field of study. In this paper, we review the evidence for diagnostic or therapeutic targets of TGF- $\beta$  signaling pathways at different stages of liver disease.

### **TGF-β AND HEPATITIS**

When the liver is affected by factors such as viruses, alcohol, drugs and autoimmune, liver cells are damaged, triggering an immune response that leads to liver inflammation. When liver disease is in a state of injury, white blood cells rapidly infiltrate liver parenchyma and promote liver inflammation by producing soluble mediators that activate other immune cells and non-parenchymal cell populations[4].

GDF-15 is a GDF subfamily member of the TGF- $\beta$  superfamily, and although the exact mechanism of GDF-15 in liver disease has not been elucidated, elevated expression of GDF-15 has been detected in the liver of animal models of diet-induced non-alcoholic steatohepatitis (NASH). The study found that mice with GDF-15 knocked out exhibited a more severe phenotype, with more steatosis, fibrosis, inflammation, metabolic disorders, or liver damage, but the mechanism of action is unclear[5]. Endogenous GDF-15 expression directly inhibits fibrosis-related genes and osteopontin in hepatic stellate cells to prevent further progression of NASH and related metabolic disorders. Given the findings of this animal study, it is feasible to consider GDF-15 as a potential innovative therapeutic target for patients with NASH.

BMP contains more than 15 ligands in the TGF- $\beta$  superfamily. BMP8B is not clearly characterized in the members of this superfamily. Although the liver pathophysiology of BMP8B is still in its infancy, BMP8B has been found to signal the SMAD2/3 and SMAD1/5/9 branches of the TGF- $\beta$ -BMP pathway in hepatic astrocytes (HSC) to promote the expression of inflammatory phenotypes[6]. At the same time, in vivo, the deficiency of BMP8B prevents HSC activation, reduces inflammation and affects the wound healing response, thus limiting the progression of NASH[6]. Since BMP8B is virtually absent in healthy livers, it may provide a promising new therapeutic target for the treatment of NASH.

TGF- $\beta$ 1 is one of the three subtypes of the TGF- $\beta$  family, and numerous studies have demonstrated that TGF- $\beta$ 1 inhibits hepatitis C virus (HCV) reproduction throughout the viral hepatitis life cycle through the TGF- $\beta$ /SMAD signaling pathway. Although the antiviral mechanism of TGF- $\beta$ 1 remains to be elucidated, a new discovery has been made that when  $\alpha$ -helix3 of TGF- $\beta$ 1 is deleted, it remains active against HCV transmission[7]. Therefore, TGF- $\beta$ 1 may inhibit HCV transmission in a manner independent of the TGF- $\beta$ /SMAD signaling pathway. Based on the findings of this study, exploring the mechanism of action of TGF- $\beta$ 1 outside the TGF- $\beta$ /SMAD signaling pathway provides another possibility for the TGF- $\beta$  subtype to treat viral hepatitis.

### TGF-B AND HEPATIC FIBROSIS

Liver fibrosis is the result of excessive extracellular matrix (ECM) deposition after liver injury, and it can lead to cirrhosis and hepatocellular carcinoma[8]. In essence, liver fibrosis is a response to repeated wound repair after continuous liver tissue damage. Continuous liver inflammation can lead to activation of stationary hepatic stellate cells and differentiation into myofibroblasts, which are the main source of ECM when hepatic astrocyte cells are injured[9].

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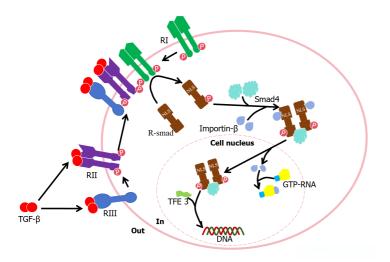


Figure 1 Transforming growth factor- $\beta$  signaling pathway on hepatocytes. During transforming growth factor- $\beta$  (TGF- $\beta$ ) signaling, TGF- $\beta$  ligands first bind to the RIII receptor. RIII receptors recruit ligands near the plasma membrane, increase the local concentration of TGF-ß ligands on the cell surface, and then transmit TGF-β to RII, activating the RII kinase domain. After RII kinase activation, the intracellular kinase domain is also phosphorylated, leading to RI kinase activation and signal transduction to the cytoplasm. Phosphorylated Smads proteins change their structure, exposing nuclear localization signals to be sent to the nucleus and activate DNA transcription. TF: Transcription factor; P: Phosphorylation; NLS: Nuclear localization signals; TGF- $\beta$ : Transforming growth factor- $\beta$ .

Activin A stimulates fibroblasts and hepatic stellate cells to produce TGF- $\beta$ 1, a central regulator of the fibrotic response [10]. Activin A can also induce the expression of TGF- $\beta$ 1 and other fibrotic regulators and intermediates, such as tissue inhibitor of metalloproteinase-1, plasminogen activator inhibitor 1, endothelin and collagen type 1, and a range of cell types [11]. In addition, TGF- $\beta$  itself stimulates the production of activin A by fibroblasts. Other fibrotic regulators such as TNF- $\alpha$ , IL13, endothelin, angiotensin, and thrombin also drive activin A production in various cell types and tissues [12]. These data suggest that activin A is an important fibrosis mediator and it may be useful as a diagnostic tool and therapeutic target against fibrotic diseases.

BMP-2 is a negative regulator of hepatocyte proliferation that is down-regulated in regenerated livers. BMP-2 significantly inhibits the expression of TGF- $\beta$  and its homologous type I and type II receptor peptides, and it induces Smad3 phosphorylation in hepatic astrocytes. Moreover, the expression levels of α-SMA and fibronectin (stimulated by TGF-\u03b31) are significantly reduced, and so are the expression levels of TGF-\u03b31-regulated epithelial-mesenchymal transformation (EMT) markers in hepatic astrocytes[13]. The interaction between BMP-2 and the TGF-β1 signaling axis may underlie the anti-fibrosis mechanism of BMP-2 during liver fibrosis, and BMP-2 may be a new therapeutic target for the treatment of that disease.

The TGF- $\beta$  subtype may also participate in liver fibrosis according to the evidence: Increased levels of TGF- $\beta$ 1 have been found in animal models of liver fibrosis caused by carbon tetrachloride or by poisoning from schistosomiasis[14]; and, increased mRNA levels of TGF-B1 have also been observed in patients with liver fibrosis[14]. Therefore, these studies suggest that TGFβ1 may provide effective drug targets for liver fibrosis treatment[15].

SMAD proteins mediate intracellular TGF-β signaling. Growing evidence suggests that miRNAs are involved in liver fibrosis processes and hepatic astrocyte activation by targeting SMAD proteins[16]. MiR-199a is important for EMT and promotes liver fibrosis by enhancing the expression of fibrosis genes encoding α1 procollagen, collagenase 3, and metalloproteinase inhibitor 1[17]. In addition, overexpression of miR-200 inhibits SMAD3 activity and attenuates TGF-β1induced fibrosis. This suggests that miR-200a is regulated by TGF-β1/SMAD3 and that it interacts with SMAD3 to demonstrate its functional activity[18]. Thus, inhibition of SMAD3 signaling may be a potential fibrotic disease intervention strategy.

GDF-15 has been associated with hepatitis and liver fibrosis. Blocking GDF-15 with neutralizing antibodies results in delayed activation of primary hepatic stellate cells and remission of carbon tetrachloride-induced hepatic fibrosis. In primary hepatic astrocytes, TGF-β pathway activation is partially inhibited by GDF-15 neutralizing antibodies[19]; and, GDF-15 can activate AMPK in mouse hepatocytes and inhibit gluconeogenesis by preventing TGF-β1/SMAD3 signaling pathway activation. Liver fibrosis is suppressed by SMAD3 phosphorylation inhibition in this pathway [20]. The positive effects of GDF-15 on hepatic astrocyte activation and liver fibrosis make it a potentially therapeutic target to alleviate liver fibrosis.

### **TGF-B AND LIVER CIRRHOSIS**

Chronic hepatitis and persistent liver fibrosis lead to cirrhosis with pseudo-lobular formation, an end-stage disease that kills more than one million people globally each year. Cirrhosis blocks essential liver functions such as albumin production, bilirubin metabolism, and clotting factor synthesis, resulting in edema, ascites, and spontaneous bleeding episodes[21]. The cirrhosis diagnosis is based on physical findings, histological examination, or imaging evidence.



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However, cirrhosis is often asymptomatic during its initial stages and is difficult to detect.

TGF- $\beta$ 1 seems to increase in proportion to the severity of fibrosis rather than the presence of myofibroblasts (MFBs) during early and middle fibrosis development stages. With cirrhosis progression, the number of macrophages expressing TGF- $\beta$ 1 decreases, but the levels of TGF- $\beta$ 1 keep increasing. And, in advanced fibrotic and cirrhotic liver disease, the hepatocytes in several pseudo-lobules continue to express TGF- $\beta$ 1[22]. Therefore, in hepatic tissues presenting advanced fibrosis and cirrhosis, TGF- $\beta$ 1 production is associated with hepatocytes and offers a therapeutic strategy targeting TGF- $\beta$  signaling. In the presence of cirrhosis, many hepatocytes in peri-lobular regions express higher levels of TGF- $\beta$  than the hepatocytes in central regions[23]. TGF- $\beta$  expression gets increased in both early and advanced primary biliary cirrhosis [24]. Given the importance of TGF- $\beta$  in the pathogenesis of cirrhosis, TGF- $\beta$  signaling pathways may prove effective targets for cirrhosis treatment.

Activins are composed of homologous or heterodimer subunits ( $\beta A$ ,  $\beta B$ ,  $\beta C$ ,  $\beta E$ ), each encoded by a single gene. During liver regeneration, the activin  $\beta A$  gene expression is decreased during hepatocyte replication and increased after the completion of liver regeneration[25]. Activin  $\beta A$  is thought to act as an autocrine inhibitor of hepatocyte growth that can induce hepatocyte apoptosis and inhibit liver regeneration in vivo, leading to increased liver damage during advanced cirrhosis stages[26].

BMP-7, a member of the BMP ligand family, can inhibit the downstream signals to TGF-β. BMP-7 actively induces type I collagen deposition by increasing matrix metalloproteinases-13 and decreasing tissue inhibitor of matrix metalloproteinases-2 expression, thereby suppressing fibrosis in a mouse cirrhosis model[27]. Thus, BMP-7 may be a useful therapeutic target in the TGF-β signaling pathway.

### **TGF-B AND LIVER CANCER**

The liver is a regenerative tissue prone to cancer[28]. Hepatocellular carcinoma (HCC) is a major cause of cancer deaths worldwide[29]. The disease usually occurs in the context of inflammation caused by a chronic viral infection or metabolic liver disease[30]; therefore, clarifying the pathogenesis of liver cancer can help identify high-risk patients and guide the development of new tumor intervention and treatment strategies. TGF-β signaling is a key pathway involved in the pathogenesis of HCC and its role needs further elucidation.

TGF- $\beta$ 1 has been shown to play a dual role in carcinogenesis: On the one hand, it acts as a tumor suppressor during the early liver cancer stages by inducing apoptosis and eliminating precursor lesions. On the other hand, liver tumor cells produce large amounts of TGF- $\beta$ , but become resistant to its pro-apoptotic effects during advanced stages[31].

Activins (members of the TGF- $\beta$  superfamily subgroup) may also have an important role in the development of liver cancer[31]. Activin E, may have growth-limiting functions similar to those of activin A[32]. Studies have shown that activin E has a negative regulatory effect on the growth of liver cancer cells and inhibits regenerative DNA synthesis[33]. The mRNA levels of activin E have been shown to increase rapidly and then get decreased to near the basal levels 48 h after a partial hepatectomy[34]. Therefore, activin E has been associated with growth inhibition and apoptosis induction, suggesting that its cell growth regulatory function may serve as a new therapeutic target against liver cancer.

The signal transduction signals between TGF- $\beta$ 1 and BMP-7 are key for differentiation and development of many cell types, including mucosal Langerhans cells[35], myoblasts[35], and embryonic stem cells[36]. Therefore, we hypothesized that the dynamic transformation from TGF- $\beta$ 1 signals to BMP-7 signals is necessary for the development of HCC. The results of a study seem to corroborate this idea: when TGFBR2 and overexpressed ACVR1 are knocked down simultaneously to interfere with the TGF- $\beta$ 1/BMP-7 pathway balance, the TGF- $\beta$ 1 is inhibited while the BMP-7 pathway is promoted and strongly activated in HCC cells, significantly exacerbating the aggressiveness and stemness of the cells [37]. Thus, TGF- $\beta$ 1/BMP-7 pathway biomarkers may be useful to predict the clinical outcomes of patients with HCC.

BMP-9 is present in large quantities in healthy livers and helps to stabilize existing blood vessels and the epithelial phenotype of liver cells. However, under chronic injury, hepatocytes undergo malignant transformation and produce large amounts of BMP-9, which down-regulate the production of molecules necessary for maintaining the interstitial contact of cells around the tumor, inducing EMT of tumor cells and promoting the development of metastasis[38]. Thus, BMP-9 enhances tumor cell growth and metastases in HCC. But, the effects of BMP-9 on the surrounding stromal cells (fibroblasts) or other cell types around the tumor remain to be studied.

### CONCLUSION

Liver diseases remain a major health threat, and exploring TGF- $\beta$  and its signaling pathways is important to improve diagnosis and therapeutic strategies. Herein, we summarized the research progress on TGF- $\beta$  and its signaling pathways in the context of liver diseases (Table 1). On the basis of the evidence, we believe that the TGF- $\beta$ /Smad signaling pathway, the TGF- $\beta$ /BMP pathway, and TGF- $\beta$ /GDF proteins may be useful targets for diagnosis and treatment of these diseases. Although some signals involved in different liver disease stages have shown contradictory effects, the discovery and study of these potential targets is important. For example, metformin, a commonly used drug in the clinical practice, has been shown to inhibit the expression of TGF- $\beta$ 1 and liver fibrosis *via* its effects on the TGF- $\beta$ /Smad signaling pathway [39]. The available diagnostic tools and treatment options against liver diseases remain inadequate. Exploring possible diagnostic and treatment strategies based on TGF- $\beta$  and its signaling pathways is a promising source of improved tools against different liver diseases.

#### Table 1 Potential targets of transforming growth factor-β signaling in liver disease

Potential targets	Members of the family	Disease	Research progress	Ref.
GDF	GDF-15	Hepatitis	GDF-15 directly inhibits fibrosis-related genes and osteopontin in hepatic stellate cells	[5]
		Hepatic fibrosis	GDF 15 attenuates chemically induced hepatic fibrosis and delays hepatic stellate cell activation	[5, 19]
		Hepatic fibrosis	Through TGF- $\beta$ 1/SMAD3 signaling pathway, SMAD3 phosphorylation is blocked and liver fibrosis is inhibited	[20]
BMP	BMP8B	Hepatitis	BMP 8B signals through the SMAD 2/3 and SMAD1/5/9 branches of the TGFβ-BMP pathway in hepatic stellate cells to promote their pro-inflammatory phenotype	[ <mark>6</mark> ]
	BMP-2	Hepatic fibrosis	BMP-2 significantly inhibits the expression of TGF- $\beta$ and its homologous type I and Type II receptor peptides, and induces Smad3 phosphorylation	[13]
	BMP-7	Liver cirrhosis	Adenovirus BMP-7 transduction induces a decrease in type I collagen deposition by increasing MMP-13 and decreasing TIMP-2 expression	[27]
	BMP-9	Liver cancer	Down-regulating the interstitial contact of cells surrounding the tumor induces epithelial-mesenchymal transformation of tumor cells, thereby promoting tumor metastasis and growth	[38]
TGF-β	TGF-β1	Hepatitis	TGF- $\beta$ 1 with its $\alpha$ -helix3 deleted is still effective against HCV, and TGF- $\beta$ 1 may inhibit HCV transmission in a manner independent of the TGF- $\beta$ /SMAD signaling pathway	[7]
	TGF-β1	Liver cirrhosis	Few macrophages express TGF-β1 when cirrhosis is present; however, the protein is expressed in hepatocytes within several pseudolobules of the liver with advanced fibrosis and cirrhosis	[22- 24]
	TGF-β	Liver cancer	TGF- $\beta$ is a tumor suppressor in the early stages of liver cancer by inducing apoptosis and eliminating precursor lesions. During later stages, liver tumor cells often become resistant to apoptosis and produce large amounts of TGF- $\beta$	[31]
Activin	TGF-β1/BMP-7	Liver cancer	Simultaneous knockdown of TGFBR2 and overexpressed ACVR1 induces complete activation of the BMP-7 pathway in HCC cells, significantly aggravating the invasiveness and stemness of HCC cells	[35- 37]
	Activin A	Hepatic fibrosis	Activin A can induce the expression of $\mbox{TGF}\beta1$ and other fibrotic regulators and intermediates	[11- 13]
	Activin βA	Liver cirrhosis	Activin $\beta$ A induces hepatocyte apoptosis and inhibits liver regeneration <i>in vivo</i> , resulting in increased damage to liver regeneration during the later cirrhosis stages	[25, 26]
	Activin βE	Liver cancer	Activin can negatively regulate the growth of hepatocellular carcinoma cells and inhibit the synthesis of raw DNA	[31- 34]
miRNA	miR-199a	Hepatic fibrosis	miR-199a enhances the expression of fibrotic genes to promote liver fibrosis, such as genes encoding $\alpha 1$ procollagen, collagenase 3, and metalloproteinase inhibitor 1	[16, 17]
	miR-200		miR-200 inhibits SMAD3 activity and attenuates TGF-β1-induced fibrosis	[ <mark>18</mark> ]

GDF: Growth and differentiation factor; TGF-β: Transforming growth factor-β; BMP: Bone morphogenetic protein; MMP: Matrix metalloproteinases; TIMP: Tissue inhibitor of matrix metalloproteinases; HCC: Hepatocellular carcinoma; HCV: Hepatitis C virus.

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ORCID number: Xiao-Ling Wang 0000-0001-7737-1078; Meng Yang 0009-0003-4488-969X; Ying Wang 0009-0007-4152-5355.

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### REFERENCES

- 1 Batlle E, Massagué J. Transforming Growth Factor-β Signaling in Immunity and Cancer. Immunity 2019; 50: 924-940 [PMID: 30995507 DOI: 10.1016/j.immuni.2019.03.024]
- Fabregat I, Moreno-Càceres J, Sánchez A, Dooley S, Dewidar B, Giannelli G, Ten Dijke P; IT-LIVER Consortium. TGF-β signalling and 2 liver disease. FEBS J 2016; 283: 2219-2232 [PMID: 26807763 DOI: 10.1111/febs.13665]
- Zhang CY, Liu S, Yang M. Treatment of liver fibrosis: Past, current, and future. World J Hepatol 2023; 15: 755-774 [PMID: 37397931 DOI: 3 10.4254/wjh.v15.i6.755]
- Hammerich L, Tacke F. Hepatic inflammatory responses in liver fibrosis. Nat Rev Gastroenterol Hepatol 2023; 20: 633-646 [PMID: 4 37400694 DOI: 10.1038/s41575-023-00807-x]
- Desmedt S, Desmedt V, De Vos L, Delanghe JR, Speeckaert R, Speeckaert MM. Growth differentiation factor 15: A novel biomarker with 5 high clinical potential. Crit Rev Clin Lab Sci 2019; 56: 333-350 [PMID: 31076013 DOI: 10.1080/10408363.2019.1615034]
- Vacca M, Leslie J, Virtue S, Lam BYH, Govaere O, Tiniakos D, Snow S, Davies S, Petkevicius K, Tong Z, Peirce V, Nielsen MJ, Ament Z, Li 6 W, Kostrzewski T, Leeming DJ, Ratziu V, Allison MED, Anstee QM, Griffin JL, Oakley F, Vidal-Puig A. Bone morphogenetic protein 8B promotes the progression of non-alcoholic steatohepatitis. Nat Metab 2020; 2: 514-531 [PMID: 32694734 DOI: 10.1038/s42255-020-0214-9]
- Zou LL, Li JR, Li H, Tan JL, Wang MX, Liu NN, Gao RM, Yan HY, Wang XK, Dong B, Li YH, Peng ZG. TGF-β isoforms inhibit hepatitis C 7 virus propagation in transforming growth factor beta/SMAD protein signalling pathway dependent and independent manners. J Cell Mol Med 2021; 25: 3498-3510 [PMID: 33682288 DOI: 10.1111/jcmm.16432]
- Zheng KX, Yuan SL, Dong M, Zhang HL, Jiang XX, Yan CL, Ye RC, Zhou HQ, Chen L, Jiang R, Cheng ZY, Zhang Z, Wang Q, Jin WZ, Xie 8 W. Dihydroergotamine ameliorates liver fibrosis by targeting transforming growth factor β type II receptor. World J Gastroenterol 2023; 29: 3103-3118 [PMID: 37346154 DOI: 10.3748/wjg.v29.i20.3103]
- Ye X, Li J, Liu Z, Sun X, Wei D, Song L, Wu C. Peptide mediated therapy in fibrosis: Mechanisms, advances and prospects. Biomed 9 Pharmacother 2023; 157: 113978 [PMID: 36423541 DOI: 10.1016/j.biopha.2022.113978]
- Hedger MP, de Kretser DM. The activins and their binding protein, follistatin-Diagnostic and therapeutic targets in inflammatory disease and 10 fibrosis. Cytokine Growth Factor Rev 2013; 24: 285-295 [PMID: 23541927 DOI: 10.1016/j.cytogfr.2013.03.003]
- Yndestad A, Larsen KO, Oie E, Ueland T, Smith C, Halvorsen B, Sjaastad I, Skjønsberg OH, Pedersen TM, Anfinsen OG, Damås JK, 11 Christensen G, Aukrust P, Andreassen AK. Elevated levels of activin A in clinical and experimental pulmonary hypertension. J Appl Physiol (1985) 2009; 106: 1356-1364 [PMID: 19196915 DOI: 10.1152/japplphysiol.90719.2008]
- Hardy CL, Lemasurier JS, Olsson F, Dang T, Yao J, Yang M, Plebanski M, Phillips DJ, Mollard R, Rolland JM, O'Hehir RE. Interleukin-13 12 regulates secretion of the tumor growth factor-{beta} superfamily cytokine activin A in allergic airway inflammation. Am J Respir Cell Mol Biol 2010; 42: 667-675 [PMID: 19635933 DOI: 10.1165/rcmb.2008-0429OC]
- Chung YH, Huang YH, Chu TH, Chen CL, Lin PR, Huang SC, Wu DC, Huang CC, Hu TH, Kao YH, Tai MH. BMP-2 restoration aids in 13 recovery from liver fibrosis by attenuating TGF-β1 signaling. Lab Invest 2018; 98: 999-1013 [PMID: 29789683 DOI: 10.1038/s41374-018-0069-9]
- Braunersreuther V, Viviani GL, Mach F, Montecucco F. Role of cytokines and chemokines in non-alcoholic fatty liver disease. World J 14 Gastroenterol 2012; 18: 727-735 [PMID: 22371632 DOI: 10.3748/wjg.v18.i8.727]
- Ahmed H, Umar MI, Imran S, Javaid F, Syed SK, Riaz R, Hassan W. TGF-B1 signaling can worsen NAFLD with liver fibrosis backdrop. Exp 15 Mol Pathol 2022; 124: 104733 [PMID: 34914973 DOI: 10.1016/j.yexmp.2021.104733]
- Xu F, Liu C, Zhou D, Zhang L. TGF-B/SMAD Pathway and Its Regulation in Hepatic Fibrosis. J Histochem Cytochem 2016; 64: 157-167 16 [PMID: 26747705 DOI: 10.1369/0022155415627681]
- Murakami Y, Toyoda H, Tanaka M, Kuroda M, Harada Y, Matsuda F, Tajima A, Kosaka N, Ochiya T, Shimotohno K. The progression of 17 liver fibrosis is related with overexpression of the miR-199 and 200 families. PLoS One 2011; 6: e16081 [PMID: 21283674 DOI: 10.1371/journal.pone.0016081]
- Wang B, Koh P, Winbanks C, Coughlan MT, McClelland A, Watson A, Jandeleit-Dahm K, Burns WC, Thomas MC, Cooper ME, Kantharidis 18 P. miR-200a Prevents renal fibrogenesis through repression of TGF-β2 expression. Diabetes 2011; 60: 280-287 [PMID: 20952520 DOI: 10.2337/db10-0892]
- 19 Qi P, Ma MZ, Kuai JH. Identification of growth differentiation factor 15 as a pro-fibrotic factor in mouse liver fibrosis progression. Int J Exp Pathol 2021; 102: 148-156 [PMID: 33983642 DOI: 10.1111/iep.12398]
- 20 Jurado-Aguilar J, Barroso E, Bernard M, Zhang M, Peyman M, Rada P, Valverde ÁM, Wahli W, Palomer X, Vázquez-Carrera M. GDF15 activates AMPK and inhibits gluconeogenesis and fibrosis in the liver by attenuating the TGF- $\beta$ 1/SMAD3 pathway. *Metabolism* 2024; **152**: 155772 [PMID: 38176644 DOI: 10.1016/j.metabol.2023.155772]
- Tsochatzis EA, Bosch J, Burroughs AK. Liver cirrhosis. Lancet 2014; 383: 1749-1761 [PMID: 24480518 DOI: 21 10.1016/S0140-6736(14)60121-5]
- Jeong WI, Do SH, Yun HS, Song BJ, Kim SJ, Kwak WJ, Yoo SE, Park HY, Jeong KS. Hypoxia potentiates transforming growth factor-beta 22 expression of hepatocyte during the cirrhotic condition in rat liver. Liver Int 2004; 24: 658-668 [PMID: 15566519 DOI: 10.1111/j.1478-3231.2004.0961.x]
- Liu X, Hu H, Yin JQ. Therapeutic strategies against TGF-beta signaling pathway in hepatic fibrosis. Liver Int 2006; 26: 8-22 [PMID: 23 16420505 DOI: 10.1111/j.1478-3231.2005.01192.x]



- Voumvouraki A, Koulentaki M, Tzardi M, Sfakianaki O, Manousou P, Notas G, Kouroumalis E. Increased TGF-β3 in primary biliary 24 cirrhosis: an abnormality related to pathogenesis? World J Gastroenterol 2010; 16: 5057-5064 [PMID: 20976842 DOI: 10.3748/wjg.v16.i40.5057
- 25 Gold EJ, Zhang X, Wheatley AM, Mellor SL, Cranfield M, Risbridger GP, Groome NP, Fleming JS. betaA- and betaC-activin, follistatin, activin receptor mRNA and betaC-activin peptide expression during rat liver regeneration. J Mol Endocrinol 2005; 34: 505-515 [PMID: 15821113 DOI: 10.1677/jme.1.01657]
- Date M, Matsuzaki K, Matsushita M, Tahashi Y, Sakitani K, Inoue K. Differential regulation of activin A for hepatocyte growth and 26 fibronectin synthesis in rat liver injury. J Hepatol 2000; 32: 251-260 [PMID: 10707865 DOI: 10.1016/s0168-8278(00)80070-7]
- 27 Cervantes-Garcia D, Cuellar-Juarez AG, Borrego-Soto G, Rojas-Martinez A, Aldaba-Muruato LR, Salinas E, Ventura-Juarez J, Muñoz-Ortega MH. Adenoviralbone morphogenetic protein7 and/or doxazosin therapies promote the reversion of fibrosis/cirrhosis in a cirrhotic hamster model. Mol Med Rep 2017; 16: 9431-9440 [PMID: 29039539 DOI: 10.3892/mmr.2017.7785]
- 28 Tomasetti C, Li L, Vogelstein B. Stem cell divisions, somatic mutations, cancer etiology, and cancer prevention. Science 2017; 355: 1330-1334 [PMID: 28336671 DOI: 10.1126/science.aaf9011]
- Yang JD, Hainaut P, Gores GJ, Amadou A, Plymoth A, Roberts LR. A global view of hepatocellular carcinoma: trends, risk, prevention and 29 management. Nat Rev Gastroenterol Hepatol 2019; 16: 589-604 [PMID: 31439937 DOI: 10.1038/s41575-019-0186-y]
- Zaidi S, Gough NR, Mishra L. Mechanisms and clinical significance of TGF-β in hepatocellular cancer progression. Adv Cancer Res 2022; 30 156: 227-248 [PMID: 35961701 DOI: 10.1016/bs.acr.2022.02.002]
- 31 Deli A, Kreidl E, Santifaller S, Trotter B, Seir K, Berger W, Schulte-Hermann R, Rodgarkia-Dara C, Grusch M. Activins and activin antagonists in hepatocellular carcinoma. World J Gastroenterol 2008; 14: 1699-1709 [PMID: 18350601 DOI: 10.3748/wjg.14.1699]
- Rodgarkia-Dara C, Vejda S, Erlach N, Losert A, Bursch W, Berger W, Schulte-Hermann R, Grusch M. The activin axis in liver biology and 32 disease. Mutat Res 2006; 613: 123-137 [PMID: 16997617 DOI: 10.1016/j.mrrev.2006.07.002]
- 33 Chabicovsky M, Herkner K, Rossmanith W. Overexpression of activin beta(C) or activin beta(E) in the mouse liver inhibits regenerative deoxyribonucleic acid synthesis of hepatic cells. *Endocrinology* 2003; 144: 3497-3504 [PMID: 12865331 DOI: 10.1210/en.2003-0388]
- 34 Lau AL, Kumar TR, Nishimori K, Bonadio J, Matzuk MM. Activin betaC and betaE genes are not essential for mouse liver growth, differentiation, and regeneration. Mol Cell Biol 2000; 20: 6127-6137 [PMID: 10913194 DOI: 10.1128/MCB.20.16.6127-6137.2000]
- Capucha T, Koren N, Nassar M, Heyman O, Nir T, Levy M, Zilberman-Schapira G, Zelentova K, Eli-Berchoer L, Zenke M, Hieronymus T, 35 Wilensky A, Bercovier H, Elinav E, Clausen BE, Hovav AH. Sequential BMP7/TGF-B1 signaling and microbiota instruct mucosal Langerhans cell differentiation. J Exp Med 2018; 215: 481-500 [PMID: 29343501 DOI: 10.1084/jem.20171508]
- Lee PT, Li WJ. Chondrogenesis of Embryonic Stem Cell-Derived Mesenchymal Stem Cells Induced by TGF\$1 and BMP7 Through Increased 36 TGFβ Receptor Expression and Endogenous TGFβ1 Production. J Cell Biochem 2017; 118: 172-181 [PMID: 27292615 DOI: 10.1002/jcb.25623]
- Ning J, Ye Y, Bu D, Zhao G, Song T, Liu P, Yu W, Wang H, Li H, Ren X, Ying G, Zhao Y, Yu J. Imbalance of TGF-β1/BMP-7 pathways 37 induced by M2-polarized macrophages promotes hepatocellular carcinoma aggressiveness. Mol Ther 2021; 29: 2067-2087 [PMID: 33601054 DOI: 10.1016/j.ymthe.2021.02.016]
- 38 Herrera B, Dooley S, Breitkopf-Heinlein K. Potential roles of bone morphogenetic protein (BMP)-9 in human liver diseases. Int J Mol Sci 2014; 15: 5199-5220 [PMID: 24670474 DOI: 10.3390/ijms15045199]
- 39 Kong L, Ma J, Dong L, Zhu C, Zhang J, Li J. Metformin exerts anti-liver fibrosis effect based on the regulation of gut microbiota homeostasis and multi-target synergy. Heliyon 2024; 10: e24610 [PMID: 38288020 DOI: 10.1016/j.heliyon.2024.e24610]



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