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Contents

Monthly Volume 16 Number 10 October 15, 2024

EDITORIAL

4037	Improving clinical outcomes of patients with hepatocellular carcinoma: Role of antiviral therapy, conversion therapy, and palliative therapy <i>Shelat VG</i>
4042	Unresectable hepatocellular carcinoma: Transarterial chemoembolization combined with lenvatinib in combination with programmed death-1 inhibition is a possible approach <i>Zhao FY, Wang DY, Qian NS</i>
4045	Advances in endoscopic diagnosis and management of colorectal cancer Li SW, Liu X, Sun SY
4052	Multidisciplinary approaches in the management of advanced hepatocellular carcinoma: Exploring future directions
	Liu XJ, Lin YX, Chen LX, Yang WJ, Hu B
4055	Clinical implications of the latest advances in gastrointestinal tumor research
	Dai W, Li YQ, Zhou Y
4060	Targeting methyltransferase-like 5-mediated sphingomyelin metabolism: A novel therapeutic approach in gastric cancer
	Zhang JJ, Yuan C, Dang SC
	REVIEW
4064	Research progress of tumor-associated macrophages in immune checkpoint inhibitor tolerance in colorectal cancer
	Fan Q, Fu ZW, Xu M, Lv F, Shi JS, Zeng QQ, Xiong DH
	MINIREVIEWS
4080	Update understanding on diagnosis and histopathological examination of atrophic gastritis: A review

Ma XZ, Zhou N, Luo X, Guo SQ, Mai P

ORIGINAL ARTICLE

Retrospective Cohort Study

4092 Establishing prognostic models for intrahepatic cholangiocarcinoma based on immune cells Wang ZR, Zhang CZ, Ding Z, Li YZ, Yin JH, Li N



Contents

Monthly Volume 16 Number 10 October 15, 2024

Retrospective Study

Constructing a nomogram to predict overall survival of colon cancer based on computed tomography 4104 characteristics and clinicopathological factors

Hu ZX, Li Y, Yang X, Li YX, He YY, Niu XH, Nie TT, Guo XF, Yuan ZL

4115 Computed tomography-based radiomic model for the prediction of neoadjuvant immunochemotherapy response in patients with advanced gastric cancer

Zhang J, Wang Q, Guo TH, Gao W, Yu YM, Wang RF, Yu HL, Chen JJ, Sun LL, Zhang BY, Wang HJ

4129 Characteristics and risk factor analyses of high-grade intraepithelial neoplasia in older patients with colorectal polyps

Zhang X, Wang Y, Zhu T, Ge J, Yuan JH

- 4138 Clinicopathological analysis of small intestinal metastasis from extra-abdominal/extra-pelvic malignancy Zhang Z, Liu J, Yu PF, Yang HR, Li JY, Dong ZW, Shi W, Gu GL
- 4146 Uninvolved liver dose prediction in stereotactic body radiation therapy for liver cancer based on the neural network method

Zhang HW, Wang YH, Hu B, Pang HW

Observational Study

4157 Small particle drug-eluting beads-transarterial chemoembolization combined with targeted therapy in the clinical treatment of unresectable liver cancer

Qi JS, Zhao P, Zhao XB, Zhao YL, Guo YC

4166 Nationwide questionnaire survey on pediatric pancreatic tumors in Japan Makita S, Uchida H, Kano M, Kawakubo N, Miyake H, Yoneda A, Tajiri T, Fukumoto K

Clinical and Translational Research

4177 Burden landscape of hepatobiliary and pancreatic cancers in Chinese young adults: 30 years' overview and forecasted trends

Chen DS, Chen ZP, Zhu DZ, Guan LX, Zhu Q, Lou YC, He ZP, Chen HN, Sun HC

Basic Study

4194 Long noncoding RNA steroid receptor RNA activator 1 inhibits proliferation and glycolysis of esophageal squamous cell carcinoma

He M, Qi Y, Zheng ZM, Sha M, Zhao X, Chen YR, Chen ZH, Qian RY, Yao J, Yang ZD

4209 Jianpi-Huatan-Huoxue-Anshen formula ameliorates gastrointestinal inflammation and microecological imbalance in chemotherapy-treated mice transplanted with H22 hepatocellular carcinoma Wang YN, Zhai XY, Wang Z, Gao CL, Mi SC, Tang WL, Fu XM, Li HB, Yue LF, Li PF, Xi SY

4232 Intratumoural microorganism can affect the progression of hepatocellular carcinoma Liu BQ, Bai Y, Chen DP, Zhang YM, Wang TZ, Chen JR, Liu XY, Zheng B, Cui ZL



Conte	<i>World Journal of Gastrointestinal Oncology</i> Monthly Volume 16 Number 10 October 15, 2024									
	Wolting Volume 10 Number 10 October 15, 2024									
4244	Clinical significance of upregulated Rho GTPase activating protein 12 causing resistance to tyrosine kinase inhibitors in hepatocellular carcinoma									
	Wang XW, Tang YX, Li FX, Wang JL, Yao GP, Zeng DT, Tang YL, Chi BT, Su QY, Huang LQ, Qin DY, Chen G, Feng ZB, He RQ									
	CASE REPORT									
4264	Rare and lacking typical clinical symptoms of liver tumors: Four case reports									
	Zhao Y, Bie YK, Zhang GY, Feng YB, Wang F									
4274	Conversion therapy in advanced perihilar cholangiocarcinoma based on patient-derived organoids: A case report									
	He YG, Zhang LY, Li J, Wang Z, Zhao CY, Zheng L, Huang XB									
4281	Transformed gastric mucosa-associated lymphoid tissue lymphoma originating in the colon and developing metachronously after <i>Helicobacter pylori</i> eradication: A case report									
	Saito M, Tanei ZI, Tsuda M, Suzuki T, Yokoyama E, Kanaya M, Izumiyama K, Mori A, Morioka M, Kondo T									
	LETTER TO THE EDITOR									
4289	Conversion therapy for unresectable hepatocellular carcinoma: Advances and challenges									
420)	He YF									

CORRECTION

Correction to "Research progress of ferroptosis regulating lipid peroxidation and metabolism in 4298 occurrence of primary liver cancer"

Shu YJ, Lao B, Qiu YY



Contents

Monthly Volume 16 Number 10 October 15, 2024

ABOUT COVER

Editorial Board of World Journal of Gastrointestinal Oncology, Gaetano Piccolo, MD, PhD, Doctor, Department of Health Sciences, University of Milan, San Paolo Hospital, Via Antonio di Rudini 8, Milan 20142, Lombardy, Italy. gpiccolo1983@gmail.com

AIMS AND SCOPE

The primary aim of World Journal of Gastrointestinal Oncology (WJGO, World J Gastrointest Oncol) is to provide scholars and readers from various fields of gastrointestinal oncology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJGO mainly publishes articles reporting research results and findings obtained in the field of gastrointestinal oncology and covering a wide range of topics including liver cell adenoma, gastric neoplasms, appendiceal neoplasms, biliary tract neoplasms, hepatocellular carcinoma, pancreatic carcinoma, cecal neoplasms, colonic neoplasms, colorectal neoplasms, duodenal neoplasms, esophageal neoplasms, gallbladder neoplasms, etc.

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ORIGINAL ARTICLE

Basic Study Clinical significance of upregulated Rho GTPase activating protein 12 causing resistance to tyrosine kinase inhibitors in hepatocellular carcinoma

Xiao-Wei Wang, Yu-Xing Tang, Fu-Xi Li, Jia-Le Wang, Gao-Peng Yao, Da-Tong Zeng, Yu-Lu Tang, Bang-Teng Chi, Qin-Yan Su, Lin-Qing Huang, Di-Yuan Qin, Gang Chen, Zhen-Bo Feng, Rong-Quan He

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Abstract

BACKGROUND

Hepatocellular carcinoma (HCC) is a major health challenge with high incidence and poor survival rates in China. Systemic therapies, particularly tyrosine kinase inhibitors (TKIs), are the first-line treatment for advanced HCC, but resistance is common. The Rho GTPase family member Rho GTPase activating protein 12 (ARHGAP12), which regulates cell adhesion and invasion, is a potential therapeutic target for overcoming TKI resistance in HCC. However, no studies on the expression of ARHGAP12 in HCC and its role in resistance to TKIs have been reported.

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AIM

To unveil the expression of ARHGAP12 in HCC, its role in TKI resistance and its potential associated pathways.

METHODS

This study used single-cell RNA sequencing (scRNA-seq) to evaluate *ARHGAP12* mRNA levels and explored its mechanisms through enrichment analysis. CellChat was used to investigate focal adhesion (FA) pathway regulation. We integrated bulk RNA data (RNA-seq and microarray), immunohistochemistry and proteomics to analyze *ARHGAP12* mRNA and protein levels, correlating with clinical outcomes. We assessed ARHGAP12 expression in TKI-resistant HCC, integrated conventional HCC to explore its mechanism, identified intersecting FA pathway genes with scRNA-seq data and evaluated its response to TKI and immunotherapy.

RESULTS

ARHGAP12 mRNA was found to be highly expressed in malignant hepatocytes and to regulate FA. In malignant hepatocytes in high-score FA groups, MDK-[integrin alpha 6 (ITGA6) + integrin β -1 (ITGB1)] showed specificity in ligand-receptor interactions. ARHGAP12 mRNA and protein were upregulated in bulk RNA, immunohistochemistry and proteomics, and higher expression was associated with a worse prognosis. ARHGAP12 was also found to be a TKI resistance gene that regulated the FA pathway. ITGB1 was identified as a crossover gene in the FA pathway in both scRNA-seq and bulk RNA. High expression of ARHGAP12 was associated with adverse reactions to sorafenib, cabozantinib and regorafenib, but not to immunotherapy.

CONCLUSION

ARHGAP12 expression is elevated in HCC and TKI-resistant HCC, and its regulatory role in FA may underlie the TKI-resistant phenotype.

Key Words: Hepatocellular carcinoma; Focal adhesion; Tyrosine kinase inhibitor; Rho GTPase activating protein 12; Drug resistance; Molecular mechanism; Biomarker

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Core Tip: Molecular therapy has the potential to reverse tyrosine kinase inhibitors (TKIs) resistance in hepatocellular carcinoma (HCC). We combined 22861 single-cell RNA sequencing malignant cells, 5113 bulk RNA data (bulk RNA-seq and microarrays), and 562 immunohistochemistry samples to explore Rho GTPase activating protein 12 (ARHGAP12) high expression from mRNA and protein. The higher its expression, the worse the prognosis of HCC patients. HCC TKIs showed ARHGAP12 resistance. Mechanistically, we confirmed ARHGAP12 can regulate focal adhesion by combining single-cell RNA sequencing and reconfirmed it from bulk RNA of HCC and HCC TKIs. Drug sensitivity analysis showed that high expression of ARHGAP12 was associated with TKIs resistance, but not with immunotherapy.

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INTRODUCTION

Hepatocellular carcinoma (HCC) has been a major challenge in the field of public health. According to statistics from 2022, liver cancer ranked the fifth in the incidence of cancer in China, with 367700 cases[1]. Among primary liver cancers, HCC is the predominant type, accounting for approximately 75%-85%[2,3]. Unfortunately, over half of HCC patients are diagnosed at the advanced stage of the disease. The five-year overall survival rate for HCC remains a concern, with only 14.1% survival rate[4-6]. Treatments for advanced HCC mostly focus on systemic therapies, with tyrosine kinase inhibitors (TKIs) being the first-line medication[7-9]. TKIs primarily inhibit tyrosine kinases involved in the vascular endothelial growth factor receptors signalling pathway in HCC, thereby reducing angiogenesis and delaying tumor recurrence[10,11]. However, in most clinical scenarios, approximately 70% of advanced HCC patients showed poor response to TKI therapy and developed primary resistance. Numerous comprehensive analyses of clinical trials indicate that the inability to identify key driver genes responsible for TKI response has been a major obstacle, which prevents HCC patients from enjoying survival benefits[12-15]. Discovery of TKI resistance genes and strategies to reverse the resistant state under TKI treatment are crucial for improving treatment outcomes in HCC patients.

As a member of the Rho GTPase family that regulates actin and cell skeleton, Rho GTPase activating protein 12 (ARHGAP12) encodes Rho GTPase-activating protein and is widely expressed in human tissues and tumor cells,

potentially modulating cell invasion and adhesion [16-18]. However, aberrant expression of ARHGAP12 can drive disease progression. Reduced expression of ARHGAP12 can impact the tightness of urothelial cells in the ureteral cell line, potentially causing bladder pain syndrome[19]. Overexpression of circular ARHGAP12 can promote invasion of nasopharyngeal carcinoma cells[18]. Homozygous variants of ARHGAP12 have been identified in an early-onset colorectal cancer patient[20]. Functionally, ARHGAP12 is closely associated with adhesion. The target of ARHGAP12, Rac1, is localized in podocytes and contributes to regulating podocyte morphology, especially cell adhesion, suggesting a potential role of ARHGAP12 in cell adhesion regulation^[21]. In melanoma cells, the RPEL mutant of ARHGAP12 with actin binding defects exhibits significantly increased activity in various biological assays, such as pseudopod formation, compared to the wild-type protein, and shows heightened presence at adhesion sites, facilitating intercellular adhesion [22]. TKIs are crucial in the treatment of HCC, and their resistance is associated with the formation of focal adhesions (FAs). Modulating the generation of FAs might serve as a breakthrough in reversing resistance^[23]. ARHGAP12, with its adhesive function, represents a potential therapeutic target, yet there are currently no reports on the expression and role of ARHGAP12 in HCC.

This study investigates the clinical significance and potential mechanisms of ARHGAP12 in HCC. Combining bulk RNA (including RNA sequencing and microarray), immunohistochemistry (IHC), and single-cell RNA sequencing (scRNA-seq), we researchers comprehensively explore the expression of ARHGAP12 in HCC and its resistance to TKIs from both mRNA and protein perspectives. Through cellular communication and pathway enrichment analysis, we aim to elucidate the potential regulatory mechanisms of ARHGAP12 in HCC and under the action of TKIs, as well as investigate the impact of ARHGAP12 on immune therapy response and drug sensitivity to TKIs.

MATERIALS AND METHODS

Assessing the expression levels of ARHGAP12 in various cell types of HCC using scRNA-seq

To explore the value of ARHGAP12 at the scRNA-seq level, we included HCC and non-HCC in the evaluation dataset, excluded metastasis and portal vein tumor thrombus, and finally collected the dataset GSE149614 from Gene Expression Omnibus. To remove low-quality cells, cells with gene expression counts below 200 or above 8000 were excluded, unique molecular identifiers less than 200, and dead or dying cells with mitochondrial counts greater than 10% were filtered. After dimensionality reduction using the uniform manifold approximation and projection algorithm with a resolution of 0.4, we performed clustering and cell type annotation [24]. Using the inferCNV package, we identified malignant hepatocytes by comparing endothelial cells and fibroblasts as reference cells with hepatocytes[25]. Finally, we obtained endothelial cells, fibroblasts, B cells, T/natural killer cells, myeloid cells, and malignant hepatocytes to investigate the expression of ARHGAP12 in these cell types. We further analyzed the expression levels of ARHGAP12 in immune cells, stromal cells, and malignant hepatocytes using the TISCH2 database.

Potential mechanism analysis of ARHGAP12 at the scRNA-seq level

To further analyze the potential mechanisms of ARHGAP12, we identified genes positively correlated with ARHGAP12 in malignant hepatocytes from scRNA-seq data using Pearson correlation ($r \ge 0.1$, P < 0.05). These genes were then subjected to enrichment analysis using the David database. Next, we scored the main FA pathway using AddModuleScore and further divided the malignant hepatocytes into high-score and low-score groups based on the median score. The "CellChat" package was also used to analyze ligand-receptor interactions within this pathway[26].

Selection and collection of gene expression data from bulk RNA resources

In order to explore whether there is homogeneity of ARHGAP12 among different ethnic groups around the world, that is, whether ARHGAP12 is at a uniform expression level in different populations. We searched and downloaded RNA-seq and gene chip datasets containing samples from HCC and non-cancerous liver tissues to identify differential expression of ARHGAP12. The inclusion criteria were as follows: (1) Samples were primarily derived from human HCC tissues; (2) HCC tissues were collected as the experimental group and non-cancerous liver tissues as the control group; and (3) There were at least 3 samples in each group. The exclusion criteria were: (1) Inadequate sample size; (2) Unextractable ARHGAP12 expression data; and (3) Samples obtained from patients with metastatic recurrent HCC. The datasets were merged according to the platform information, and abnormal data samples were removed. Inter-group correction was performed first, and then batch correction was performed using the ComBat function of sva and limma-voom (Supplementary Figure 1).

IHC staining of samples from our institution and external proteomic analysis

To explore the homogeneity of different populations and investigate the common expression level of ARHGAP12, a total of 816 tissue samples (404 HCC samples and 412 non-cancer control samples) were collected from the First Affiliated Hospital of Guangxi Medical University and Red Cross Hospital of Yulin City. IHC staining was performed according to a strict protocol. Tissues were first fixed with formalin and embedded in paraffin, and then sectioned to prepare tissue sections. Subsequently, the sections were dewaxed and hydrated with xylene, and antigen retrieval was performed in boiling ethylenediaminetetraacetic acid buffer (pH = 9.0) to restore antigenicity. After incubation with hydrogen peroxide to effectively inhibit endogenous enzyme activity, the sections were stained with a polyclonal antibody against ARHGAP12 (PA551384, Invitrogen™, dilution ratio 1:1000) and incubated at 37 °C for 30 minutes. At the end of the staining process, the slides were stained by diaminobenzidine colorimetric reaction and rinsed with phosphate buffered



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saline buffer between steps to remove unbound antibodies. After hematoxylin staining, the slides were dehydrated with a series of gradient ethanol solutions and finally mounted with neutral resin. The stained sections were scored under a microscope, where the brown area indicated a positive reaction and the blue area indicated a negative control. The staining results on the slides were scored as follows: 0 (no staining), 1 (pale yellow), 2 (light brown), and 3 (dark brown). The percentage of positive cells on the slides was defined using a five-point scoring system: $0 \leq 5\%$ of cells), 1 (6%-25% of cells), 2 (26%-50% of cells), 3 (51%-75% of cells), and 4 (\geq 76% of cells). The IHC total score was evaluated by two pathologists independently, multiplying the percentage of positive cells and staining score[27-29]. All patients provided informed consent, and this study was approved by the Ethics Committee of the First Affiliated Hospital of Guangxi Medical University (No. 2020-KY-Guoji-058). Additionally, we collected proteomic data from Proteomic Data Commons for subsequent integrated analysis.

Relationship between ARHGAP12 expression and clinical parameters

Using the inclusion and exclusion criteria mentioned above, along with the availability of prognostic information, we obtained three prognostic datasets [E-TABM-36, GSE76427, and The Cancer Genome Atlas (TCGA)]. We performed Kaplan-Meier analysis to explore the relationship between ARHGAP12 expression and prognosis (Supplementary Figure 2). We further analyzed the differential expression of ARHGAP12 among various clinical and pathological parameter groups using the clinical data collected from the abovementioned IHC analysis.

Expression levels of ARHGAP12 in TKI-resistant HCC

Using the inclusion and exclusion criteria mentioned above, along with information on TKI treatment, we collected and downloaded microarray and high-throughput sequencing datasets of TKI-resistant HCC. The TKI-sensitive HCC samples served as the control group, while the TKI-resistant HCC samples were the experimental group (Supplementary Figure 3).

Potential mechanisms of ARHGAP12 in TKI resistance

First, we explored the protein-protein interaction (PPI) network of ARHGAP12 using GeneMANIA. We then integrated the resistance genes calculated by standardized mean difference (SMD) under TKI treatment with the upregulated differentially expressed genes from bulk RNA data analysis (SMD > 0, P < 0.05). We intersected these genes with the ARHGAP12 positively correlated genes obtained from Pearson correlation analysis in bulk RNA data (correlation coefficient ≥ 0.3 in at least 10 datasets, P < 0.05). The intersected genes were subjected to pathway enrichment analysis using the David database. Finally, we intersected the FA pathway genes obtained previously with genes identified in the same pathway from the scRNA-seq data.

Relationship between ARHGAP12 and potential drug treatment

TCGA data were used to analyze the effect of ARHGAP12 expression on immune escape and immunotherapy. Cellminer database and GDSC2 data were integrated to predict the relationship between ARHGAP12 expression and TKI-drug sensitivity using oncoPredict package.

Statistical analysis

To evaluate the differences in ARHGAP12 mRNA and protein expression levels in HCC and TKI-resistant HCC samples, we first used the more robust, reliable, and applicable Wilcoxon rank-sum test, and used the "pROC" package to draw receiver operating characteristic (ROC) curves to jointly analyze the ARHGAP12 expression levels of individual data. In the process of using SMD to comprehensively evaluate the ARHGAP12 expression level, when the heterogeneity was low $(l^2 < 50\%)$, we used a fixed effect model to calculate the SMD; otherwise, a random effect model was used. The summary ROC (sROC) curve was drawn using STATA 16.0, and the larger the area under the curve (AUC), the higher the ARHGAP12 expression level. The Egger's and Begg's tests were used to assess publication bias, and P > 0.05 indicated no publication bias. Except for the Egger's and Begg's tests, P < 0.05 was considered statistically significant. Figure 1 showed the overall framework design of this research.

RESULTS

The high expression of ARHGAP12 in malignant hepatocytes

With the help of the inferCNV package, malignant hepatocytes were identified from hepatocytes based on epithelial cells and fibroblasts. The results showed that ARHGAP12 was significantly upregulated in 15990 malignant hepatocytes (Figure 2). Data from the TISCH2 database revealed high expression of ARHGAP12 in 4500 malignant cells from LIHC_GSE166635 and 2371 malignant cells from LIHC_GSE146115 (Figure 3).

The potential molecular mechanisms of ARHGAP12 at the scRNA-seq level

To explore the potential mechanisms of ARHGAP12, we performed enrichment analysis on ARHGAP12-correlated genes $(r \ge 0.1, P \le 0.05)$ in malignant hepatocytes. The results revealed that the FA pathway ranked first in terms of enrichment (Figure 4A). Within the FA pathway score, fibroblast cells, endothelial cells, and malignant hepatocytes ranked as the top three cell types, with higher scores indicating higher pathway gene expression within the gene set (Figure 4B). Cellular communication analysis revealed a greater number of interaction networks between malignant hepatocytes with high FA



Wang XW et al. ARHGAP12 in HCC

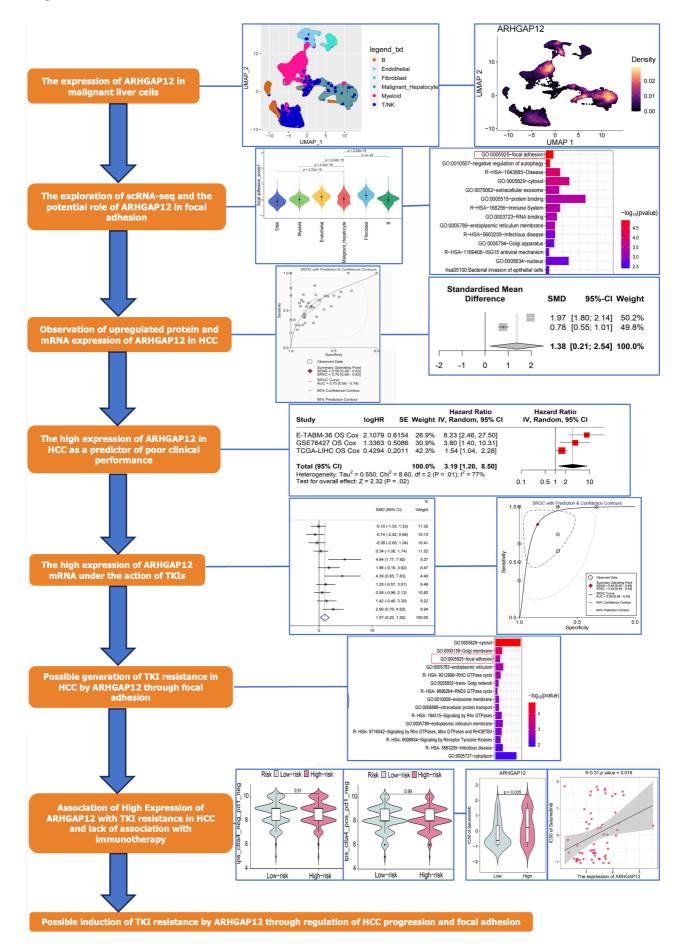
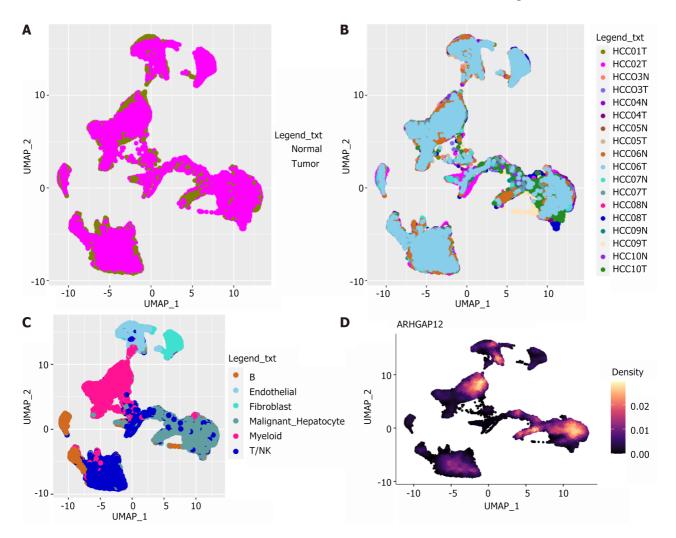


Figure 1 Research overflow of this study. ARHGAP12: Rho GTPase activating protein 12; HCC: Hepatocellular carcinoma; TKI: Tyrosine kinase inhibitor.

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October 15, 2024 Volume 16 Issue 10



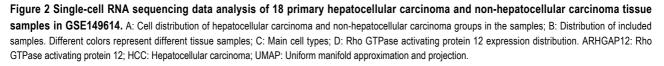




Figure 3 Expression levels of Rho GTPase activating protein 12 in various cell types in the TISCH2 database. ARHGAP12: Rho GTPase activating protein 12.

pathway scores and fibroblast cells, while malignant hepatocytes with low FA pathway scores exhibited stronger interactions with myeloid cells (Figure 4C and D).

Additionally, to further investigate the potential mechanisms of the FA pathway, we identified the interaction between ligands and receptors within the pathway. In the high FA pathway score group of malignant hepatocytes, the strongest incoming and outgoing actions were observed in the MK signaling transduction (Figure 5A). In the ligand-receptor analysis targeting MK, the signal transmission from MDK-(ITGA6 + ITGB1) to endothelial cells was found to be stronger in the high FA pathway score group of malignant hepatocytes (Figure 5B). MDK exhibited the highest expression in the

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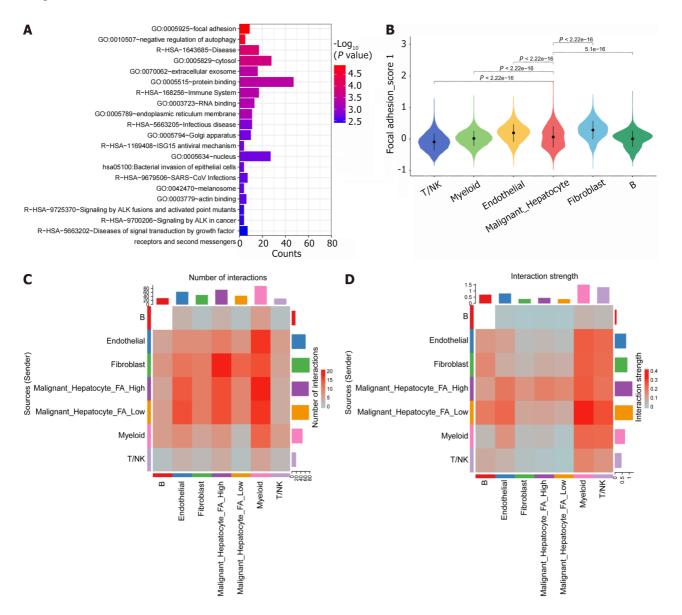


Figure 4 Investigation of the potential mechanisms of Rho GTPase activating protein 12 in the focal adhesion pathway. A: Enrichment analysis; B: Focal adhesion pathway scores in various cell types; C: Number of interactions in various cell types; D: Strength of interactions in various cell types. FA: Focal adhesion.

high FA pathway score group of malignant hepatocytes in heatmap (Figure 5C).

Upregulation of ARHGAP12 mRNA in bulk RNA data

According to the Wilcoxon rank-sum test, ARHGAP12 mRNA showed significant overexpression across 10 platforms (P < P0.05, AUC \ge 0.7, Supplementary Figures 4, 5, 6 and 7). In the random-effects model, comprehensive overexpression of ARHGAP12 mRNA was observed in 5113 HCC samples [SMD = 0.44, 95% confidence interval (CI): 0.30-0.58, Figure 6A]. Furthermore, the sROC analysis confirmed the high expression of ARHGAP12 mRNA (AUC = 0.70, Figure 6B). Egger's and Begg's tests indicated the absence of publication bias (P = 0.412 and 0.838) (Figure 6C and D).

Upregulation of ARHGAP12 protein in HCC tissues

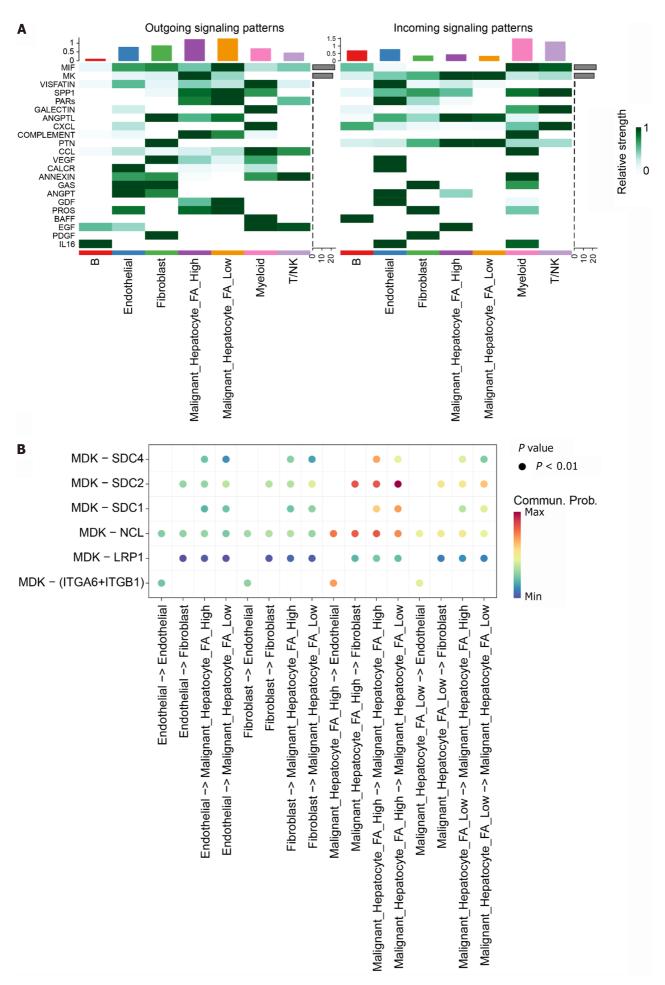
Immunohistochemical data indicated intense staining of ARHGAP12 antibody in HCC tissues (Figure 7A). Violin plots and ROC curves demonstrated the high expression of ARHGAP12 protein (Figure 7B). Combining in-house immunohistochemical data with proteomic data further confirmed the upregulation of ARHGAP12 protein in 562 HCC samples (SMD = 1.38, 95%CI: 0.21-2.54, Figure 7C).

Relationship of high ARHGAP12 expression in HCC with clinical pathological progression

To further investigate the relationship between ARHGAP12 expression and clinical significance, integrated prognosis analysis indicated that higher ARHGAP12 expression correlated with poorer patient prognosis (Figure 8A). The scatter plots showed that ARHGAP12 expression was higher in stage M2 compared to stage M0 in vascular invasion, higher in stage T2 compared to stage T1, and higher in stage N1 compared to stage N0 in lymph node staging (Figure 8B, C and D).



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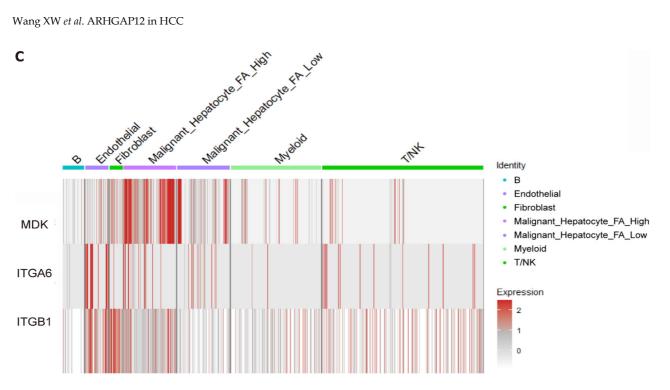


Figure 5 Analysis of the MK communication network. A: Heatmap of outgoing and incoming signal patterns; B: Ligand-receptor network of MK signaling; C: Heatmap plot of MDK-(integrin alpha 6 + integrin β-1) expression. ITGA6: Integrin alpha 6; ITGB1: Integrin β-1; FA: Focal adhesion.

Table 1 presented the statistical data regarding ARHGAP12 expression in clinical samples in relation to age, gender, pathological grading, tumor necrosis, satellite nodules, vascular invasion, and tumor-node-metastasis stages, with significant differences observed in gender, vascular invasion, and TN staging (P < 0.05).

Upregulation of ARHGAP12 mRNA under the TKIs

Under the treatment of TKIs, two datasets demonstrated high expression of ARHGAP12 mRNA (P < 0.05, AUC ≥ 0.7 , Supplementary Figure 8). Integrated analysis revealed a comprehensive upregulation of ARHGAP12 mRNA in 41 TKIresistant HCC samples (SMD = 1.07, 95% CI: 0.22-1.92, Figure 9A). Likewise, the sROC curve demonstrated the high expression of ARHGAP12 mRNA in TKI resistance (AUC = 0.90, Figure 9B).

Potential regulatory mechanisms of ARHGAP12 in TKI-resistant HCC

To further explore the potential functions of ARHGAP12 and its regulatory mechanisms in TKI treatment, we employed PPI analysis to investigate the potential interacting factors of ARHGAP12. We found that ARHGAP12 was involved in endocytosis and regulates cellular morphological changes (Supplementary Figure 9). Subsequently, we intersected the TKI-resistant gene set, upregulated differentially expressed genes from bulk RNA data, and genes that were positively correlated with ARHGAP12. We found that the intersection genes were mainly enriched in FA. To validate this finding, we intersected the FA pathway genes obtained with those in the scRNA-seq data, and identified ITGB1 as the unique common gene (Figure 10).

The relationship between ARHGAP12 expression and the response to TKIs as well as immunotherapy in HCC

To explore the relationship between ARHGAP12 expression and drug treatment in HCC, we first investigated TCGA data and observed that higher ARHGAP12 expression was associated with increased immune escape. However, no significant correlation was found with immunotherapy response (Figure 11A). Assessment of TKI drug sensitivity revealed that higher ARHGAP12 expression was associated with poorer response to sorafenib, cabozantinib, and regorafenib treatments, with increased dosage requirement correlated with elevated ARHGAP12 expression (P < 0.05, Figure 11B).

DISCUSSION

Aberrant expression of ARHGAP12 is a risk factor affecting the prognosis of certain diseases, potentially promoting cell invasion and adhesion[16-18]. In a novel study, we integrated multiple scRNA-seq datasets, confirming that ARHGAP12 mRNA was significantly overexpressed in malignant hepatocytes and uncovering its potential molecular mechanisms through the regulation of the FA pathway. In malignant hepatocytes, the high FA pathway score group predominantly regulated the MK pathway. Notably, within the MK pathway, the MDK-(ITGA6 + ITGB1) ligand- receptor interaction exhibited distinct specificity. Furthermore, by combining bulk RNA, IHC, and proteomics data, we determined that ARHGAP12 mRNA and protein were highly expressed in HCC tissues, with higher expression levels correlating with poorer patient outcomes. This study was the first to report elevated ARHGAP12 expression in TKI-resistant samples,



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Table 1 Expression of Rho GTP	Pase activating pro	tein 12 in different g	roups based on c	linical pathological features	
Clinicanothological factures	ARHGAP12 pr	otein expression	t(t toot) or E(ANOVA)	<i>P</i> value	
Clinicopathological features	Number	mean	SD	<i>t</i> (<i>t</i> -test) or <i>F</i> (ANOVA)	P value
Age				1.6400	0.1017
> 60	101	5.743	3.504		
≤ 60	303	6.342	3.065		
Gender				2.1130	0.0352
Male	339	6.046	3.172		
Female	65	6.954	3.179		
Pathology grade				1.4470	0.2366
Grade I-II	203	6.043	3.347		
Grade II-III	85	5.995	3.099		
Grade III-IV	113	6.630	2.960		
Tumor necrosis				1.5980	0.1112
Yes	178	6.279	2.931		
No	80	5.570	3.989		
Satellite nodule				0.7686	0.2944
Yes	81	6.079	3.145		
No	287	6.199	3.275		
Vascular invasion				4.9580	0.0075
M0	199	5.733	3.048		
M1	142	6.501	3.217		
M2	58	7.062	3.438		
TNM stages					
Т				5.1990	0.0072
T1	45	4.631	3.734		
T2	44	7.063	3.982		
Т3	9	4.000	4.000		
Ν				2.1830	0.0315
N0	96	5.563	4.016		
N1	3	10.670	2.309		
М				1.5650	0.1208
M0	98	5.653	4.035		
M1	1	12.000	0.000		
HBV				0.3460	0.7296
HBV-positive	182	6.332	3.626		
HBV-negative	62	6.155	3.002		

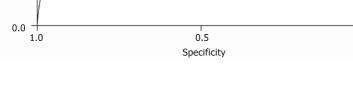
Microvascular invasion grading: M0 level: Microvascular invasion not found; M1 grade: ≤ 5 microvascular invasions and occur in the area of proximal adjacent liver tissue (≤ 1 cm); M2 grade: > 5 microvascular invasions, or microvascular invasions that occur in the area of liver tissue distally adjacent to the cancer (> 1 cm). ARHGAP12: Rho GTPase activating protein 12; TNM: Tumor-node-metastasis; HBV: Hepatitis B virus.

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Α	Experimental	Control	Standardised mean	
Study	-		SD Difference	SMD 95%-CI Weight
E_MTAB_4171 E_MTAB_8887 GPL10553 GPL10553 GPL10558 GPL10687 GPL10999 GPL11154 GPL13667 GPL14550 GPL14951 GPL15433 GPL15433 GPL16043 GPL16099 GPL16099 GPL16791 GPL17077 GPL17586 GPL17077 GPL17586 GPL18451 GPL20301 GPL20795 GPL20115 GPL201047 GPL20795 GPL21047 GPL20795 GPL21047 GPL23126 GPL24676 GPL3921 GPL4133 GPL5175 GPL570 GPL571 GPL6244 GPL6480 GPL6793 GPL6947 GPL9052 GPL9115 GPL96 ICGC_LIRI_JP TCGA_GTEX_HCC Random effects model Heterogeneity: $I^2 = 83\%$, τ	15 12.76 0.5919 23 4.28 0.8008 24 2.82 0.1636 523 5.61 0.2093 268 2.95 0.1818 3 3.95 0.7460 245 3.10 1.1921 88 6.63 0.9799 338 6.12 0.5210 67 3.56 0.0669 93 9.06 1.0650 21 3.40 0.5733 10 7.60 0.8187 25 1.21 0.799 310 7.60 0.8187 25 1.21 0.799 305 0.261 1.2859 310 7.60 0.8187 25 2.261 1.2859 310 7.21 0.1606 215 2.952 2.5300 12 1.26 0.4067 48 4.052 2.1587 30 3.71 0.9566 10 3.29	15 12.68 0.3 17 4.08 0.5 24 2.73 0.1 403 5.53 0.12 188 2.93 0.11 3 2.65 0.22 168 2.84 1.00 342 5.82 0.33 10 3.50 0.00 18 8.25 0.92 21 3.37 0.00 26 7.62 0.92 25 1.19 0.73 5 -0.04 0.0 114 4.73 0.83 6 1.13 0.11 8 9.88 0.33 34 7.07 0.11 13 3.07 2.00 12 1.28 0.33 30 3.61 2.33 30 3.61 2.33 30 3.61 2.33 30 3.61 2.33 200 2.89 0.11 18 10.39 0.60 48	983	0.15 [-0.57; 0.86] 1.8% 0.27 [-0.36; 0.90] 2.1% 0.66 [0.07; 1.24] 2.2% 0.46 [0.33; 0.59] 3.6% 0.14 [-0.03; 0.31] 3.5% 1.83 [-0.48; 4.14] 0.3% 0.22 [0.03; 0.42] 3.4% -0.07 [-0.43; 0.28] 3.0% 0.70 [0.55; 0.86] 3.5% 0.91 [0.23; 1.59] 1.9% 0.77 [0.25; 1.29] 2.4% 0.48 [-0.13; 1.09] 2.1% -0.02 [-0.75; 0.71] 1.8% 0.02 [-0.75; 0.71] 1.8% 0.02 [-0.53; 0.58] 2.3% 0.86 [-0.47; 2.19] 0.8% 0.46 [0.23; 0.69] 3.3% -0.18 [-1.57; 1.21] 0.8% 1.32 [1.00; 1.63] 3.1% 0.17 [-0.81; 1.15] 1.3% 0.97 [0.47; 1.47] 2.5% -0.05 [-0.61; 0.51] 2.3% -0.07 [-0.87; 0.73] 1.6% 0.20 [-0.26; 0.66] 2.6% 0.80 [0.33; 1.27] 2.6% 0.68 [-0.22; 1.59] 1.4% -0.04 [-1.64; 1.56] 0.6% 0.49 [0.30; 0.69] 3.4% 1.05 [0.85; 1.25] 3.4% 1.05 [0.85; 1.25] 3.4% 1.05 [0.75; 2.25] 1.8% 0.68 [0.26; 1.09] 2.8% 0.58 [0.47; 0.70] 3.6% 0.57 [0.31; 0.84] 3.2% 0.51 [0.18; 0.85] 3.0% 0.31 [-0.00; 0.61] 3.1% 0.39 [0.11; 0.67] 3.2% -0.67 [-0.96; -0.39] 3.2% 0.04 [-0.32; 0.40] 3.0% 0.31 [-0.00; 0.61] 3.1% 0.39 [0.11; 0.67] 3.2% 0.04 [-0.32; 0.40] 3.0% 0.18 [0.22; 1.57] 1.9% 0.044 [0.30; 0.58] 100.0%
B SROC	C with Prediction & Co	nfidence Contours	Ø	
Sensitivity		22 S 13		Observed Data Summary Operating Point SENS = 0.56 [0.48 - 0.63] SPEC = 0.76 [0.68 - 0.83] SROC Curve AUC = 0.70 [0.66 - 0.74] 95% Confidence Contour 95% Prediction Contour

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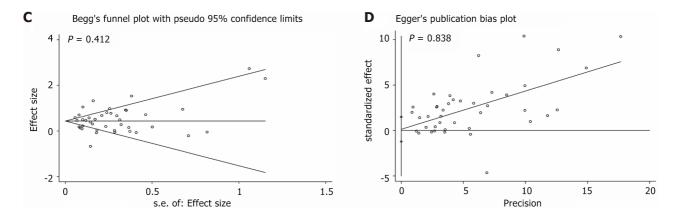


Figure 6 Integrated analysis on Rho GTPase activating protein 12 expression in hepatocellular carcinoma. A: Forest plot; B: Summary receiver operating characteristic curve; C: Egger's; D: Begg's. SMD: Standardized mean difference; CI: Confidence interval; sROC: Summary receiver operating characteristic; AUC: Area under the receiver operating characteristic curve.

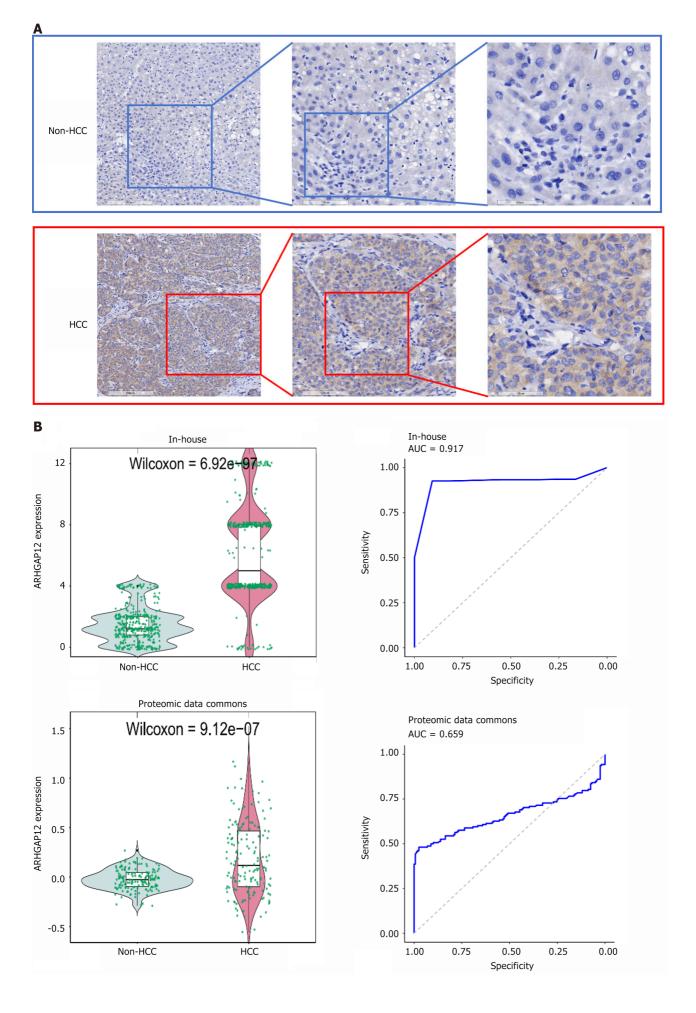
suggesting its potential role as a resistance gene in HCC. The integration of bulk RNA data further revealed that ARHGAP12 might also regulate the FA pathway, with scRNA-seq data identifying ITGB1 as a potential intersecting gene. Our results also showed that higher ARHGAP12 expression was associated with reduced efficacy of lapatinib, selumetinib and trametinib, while it has no significant impact on the efficacy of immunotherapy. Overall, ARHGAP12 appeared to contribute to TKI resistance in HCC by influencing the FA pathway, emerging as a potential key therapeutic target in the treatment of HCC.

Aberrant expression of ARHGAP12 may contribute to disease progression and lead to poor prognosis. In previous studies on ARHGAP12, upregulated miR-199a-5p has been shown to suppress ARHGAP12 expression, leading to decreased urothelial tightness in human ureteral cells and potentially causing bladder pain syndrome[19]. Additionally, in noncoding RNA, overexpression of circular ARHGAP12 promoted the migration and invasion of nasopharyngeal carcinoma cells, accelerating malignant progression through cytoskeletal remodeling[18]. Furthermore, under doxorubicin induction, upregulated circular ARHGAP12 enhanced apoptosis in mouse cardiac tissues[30]. In the genomic analysis of a patient with early-onset colorectal cancer, researchers identified the homozygous missense mutation in the ARHGAP12 gene^[20], suggesting that variant expression of ARHGAP12 could be a significant factor promoting colorectal cancer progression from the mutation perspective. Moreover, hepatocyte growth factor played a crucial role in inhibiting the transcription and protein expression of ARHGAP12 during a hepatocyte growth factor-induced invasive growth program^[31]. In melanoma, the binding of G-actin to ARHGAP12 could downregulate GAP activity, thereby controlling Rac-dependent signalling pathways and promoting the formation of invasive pseudopodia. Concurrently, high expression of ARHGAP12 has been observed to enhance cell adhesion strength, revealing its dual role in the regulation of cell behavior[22]. While no previous studies have focused on ARHGAP12 in HCC, previous studies on the ARHGAP gene family found that overexpression of ARHGAP18 stimulated growth, migration and invasion of HCC cells[32], while knocking down ARHGAP11A significantly inhibited HCC cell proliferation, and the upregulation of ARHGAP11A promoted mesenchymal-epithelial transition[33]. Interestingly, in the current study, no relationship was found between immunotherapy and ARHGAP12 expression, but high expression of ARHGAP12 resulted in TKI resistance. This suggests the potential significance of ARHGAP12 as a target for TKI treatment. The multi-level and multi-platform exploration conducted in this study strongly suggests that high expression of ARHGAP12 promotes adverse outcomes in HCC and contributes to TKI resistance. Thus, ARHGAP12 may serve as a potential therapeutic target for HCC.

ARHGAP12 may trigger TKI resistance in HCC through FA. Studies have indicated that FAs are large supramolecular complexes closely linked to various cellular behaviors and pathological events, playing a crucial role in interactions between cells and the extracellular matrix as well as the cytoskeleton and regulating cell migration, polarisation and the development of metastatic cancers[34-36]. As a component of FAs, focal adhesion kinase (FAK) plays a central role, functioning as a non-receptor tyrosine kinase and an adaptor protein that plays important roles in processes such as cell adhesion, migration, and survival. Aberrant activation or expression of FAK has been observed in many types of cancers, making it a potential target for cancer therapy and showing particular promise in TKI treatments[37-39]. Other research findings have demonstrated that Ras-related protein Rab-32 and platelet response protein 1 could activate the FAK signalling pathway in non-small cell lung cancer in vitro, conferring resistance to osimertinib and reducing the response to treatment[40]. In osimertinib-resistant lung cancer cell lines, activated FAK was present, and knocking down FAK expression effectively reversed osimertinib resistance mediated by interleukin-6[41]. Interestingly, ARHGAP12 has been shown to have a unique role in regulating cell adhesion. When the expression levels of ARHGAP12 increased, it localized mainly at cell adhesion focal points, significantly advancing the development of the cell adhesion process^[21,22]. In the current study, we have extensively explored the role of ARHGAP12 in the mechanism of resistance in HCC for the first time. ARHGAP12 may affect cell adhesion events by modulating the FA pathway, leading to TKI resistance. For example, with increasing ARHGAP12 expression, the efficacy of lapatinib, selumetinib and trametinib diminished. Furthermore, in PPI studies, we found that ARHGAP12 participated in membrane invagination, regulating changes in cell morphology, with alterations in endocytosis accompanying the generation of FAs[42]. This indirectly confirms the potential significant role of FAs in ARHGAP12's mechanism. This discovery offers a new perspective for addressing TKI resistance in the



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С			Experin	nental	I	Con	trol	9	Standa	rdise	ed mea	n			
	Study	Tota	l Mean	SD	Total	Mean	SD		Dif	feren	ce		SMD	95%-CI	Weight
	in-house Proteomic Data Commons	404 158	6.19 3 0.19 0				1.1505 0.1021				-			[1.80; 2.14] [0.55; 1.01]	
	Random effects model Heterogeneity: I^2 = 99%, τ^2 =	562 0.6950	, p < 0.0	1	571			-2	-1		1	2	1.38	[0.21; 2.54]	100.0%

Figure 7 Expression of Rho GTPase activating protein 12 protein in hepatocellular carcinoma tissues. A: Immunohistochemistry staining of control and hepatocellular carcinoma samples; B: Violin plots and receiver operating characteristic curves; C: Comprehensive expression levels of Rho GTPase activating protein 12 protein. ARHGAP12: Rho GTPase activating protein 12; HCC: Hepatocellular carcinoma; SMD: Standardized mean difference; CI: Confidence interval; AUC: Area under the receiver operating characteristic curve.

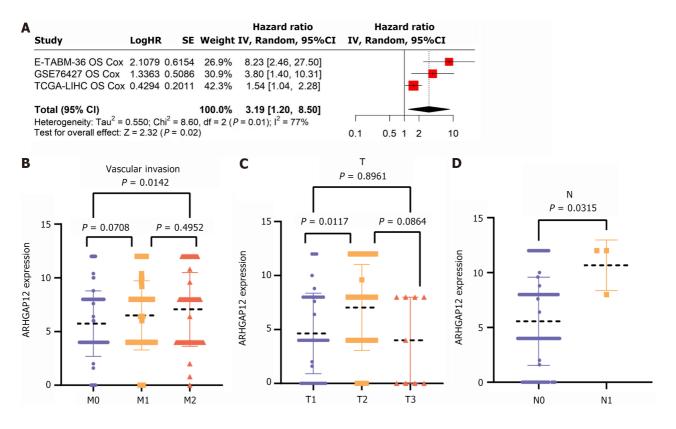
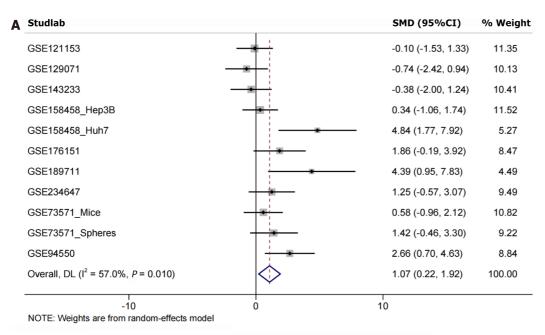


Figure 8 Relationship of Rho GTPase activating protein 12 with clinical pathological parameters in hepatocellular carcinoma. A: Comprehensive prognostic levels of Rho GTPase activating protein 12; B: Vascular invasion; C: T stage; D: N stage. ARHGAP12: Rho GTPase activating protein 12; C! Confidence interval; OS: Overall survival; HR: Hazard ratio.

treatment of HCC.

Targeted therapy of ARHGAP12 may be the key to treating HCC and reversing TKI resistance in HCC. In the complex pathogenesis of HCC, specific molecular targeted therapy provides a new and effective option for patients with advanced HCC[43,44]. As a targeted drug for the treatment of advanced HCC, TKI can inhibit HCC angiogenesis, invasion and metastasis by blocking signalling pathways, such as vascular endothelial growth factor and platelet-derived growth factor[45]. However, clinical data show that up to 70% of HCC patients have poor TKI treatment results, and both primary and acquired TKI resistance have led to limited survival benefits with TKI treatment[46,13]. The discovery of new molecular targets may provide a way to overcome TKI resistance in the treatment of HCC. Recent studies have found that as the first metabolic enzyme in the serine biosynthesis pathway, inactivation of PHGDH could increase reactive oxygen species levels and promote HCC apoptosis after sorafenib treatment. Thus, targeting PHGDH is a potential method to overcome TKI resistance in liver cancer^[47]. In addition, upregulated AXL protected resistant cells from oxidative stress, mitochondrial damage and immunogenic cell death by inhibiting tumor necrosis factor- α and STING-type I interferon pathways, which in turn led to poor responses of sorafenib- and lenvatinib-resistant HCC to TKI and immunotherapy. Targeting AXL combined with anti-programmed cell death protein 1 may provide a new option for HCC patients who have failed to respond to TKI treatment[48]. All signs indicate that molecular targeted therapy has great potential in reversing HCC resistance to TKI, and high expression of ARHGAP12 is not only an important driving force for HCC progression but may also be a core factor in the formation of TKI resistance. Therefore, ARHGAP12 is not only of great significance as a potential therapeutic target for HCC, but its targeted inhibition is also likely to become



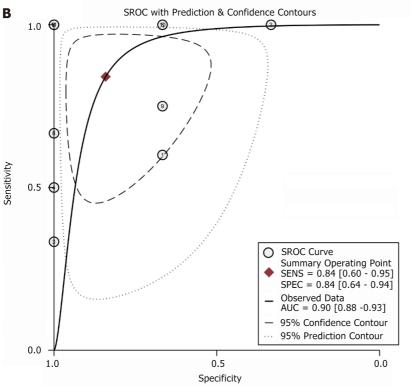


Figure 9 High expression of Rho GTPase activating protein 12 under tyrosine kinase inhibitors treatment. A: Forest plot showing elevated expression of Rho GTPase activating protein 12; B: Summary receiver operating characteristic curve plot. SMD: Standardized mean difference; CI: Confidence interval; sROC: Summary receiver operating characteristic; AUC: Area under the receiver operating characteristic curve.

provide a breakthrough in reversing drug resistance and restoring drug sensitivity. Precisely targeting ARHGAP12 is expected to bring new hope for survival to HCC patients whose treatment is limited due to drug resistance. This strategy not only heralds innovation in the treatment model, but is also likely to change the landscape of HCC treatment efficacy.

ARHGAP12 may regulate FA through ITGB1. Integrins, part of the cell adhesion molecule family, are crucial for key interactions between cells and the extracellular matrix, playing essential roles in cell adhesion, spreading, and migration. Integrins act as bridges between the extracellular matrix and the Rho GTPase family, transmitting extracellular information to the cell nucleus *via* the actin cytoskeleton. For example, ITGB1 regulated actin dynamics through Ras homolog A[17,49,50]. Integrins containing ITGB1 as a subunit played indispensable roles in various cell signalling pathways to maintain normal physiological functions. However, in the context of tumor cells, the roles of integrins underwent transformation as they not only contributed to the maintenance of stemness in tumor cells but also enhanced invasion, metastasis, and chemotherapy resistance. Notably, ITGB1, as a critical subunit of integrin proteins, could bind to multiple ligands, exhibiting diverse biological functions[51]. As the core of the integrin family, ITGB1's adhesive function

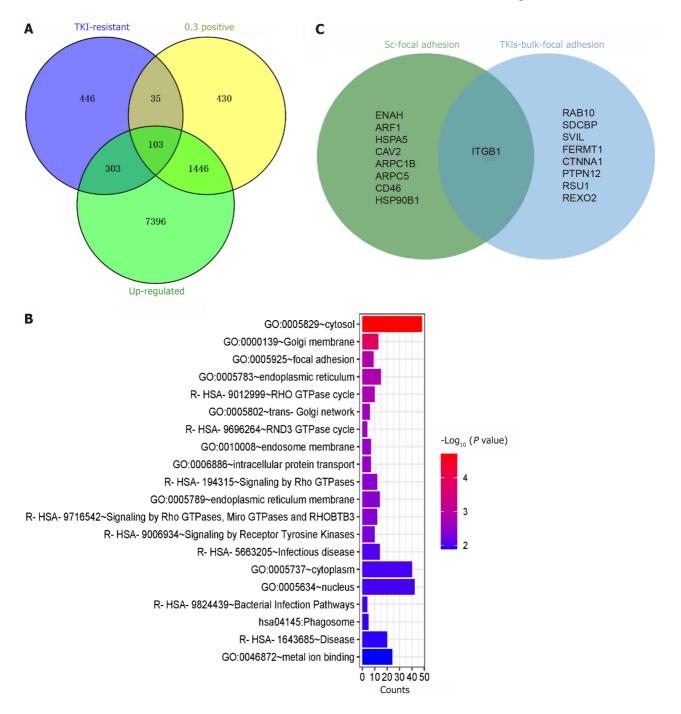


Figure 10 Potential regulatory mechanisms of Rho GTPase activating protein 12. A: Intersection of tyrosine kinase inhibitor-resistant genes, upregulated differentially expressed genes from bulk high-throughput data, and Rho GTPase activating protein 12 positively correlated genes; B: Enrichment analysis; C: Intersection of genes in the focal adhesion pathway from single-cell RNA sequencing and bulk RNA data. TKI: Tyrosine kinase inhibitor.

has been confirmed to be the key mechanism for disease progression[52]. Lysyl oxidase-like 2 has been shown to stabilize the protein structure of ITGB1 and activate the FAK/SRC signalling pathway mediated by ITGB1, significantly accelerating the formation of FAs. This process enhanced the pro-migratory function of ribosomal protein S7, promoting cell migration and invasion[53]. Elevated expression of ITGB1, a key gene in collagen-related cell adhesion molecules, could promote HCC progression[54]. Current research suggests that ITGB1 has a role in regulating FAs in HCC. The current study discovered a potential regulatory gene, ARHGAP12, in the FA pathway. Regarding FAs, ARHGAP12 may potentially upregulate ITGB1 to increase cell adhesion levels, promote HCC progression and, consequently, trigger TKI resistance, leading to poor prognosis. However, further investigation is required to determine the potential synergistic regulation of FAs by ARHGAP12 and ITGB1.

Our study has several limitations. First, clinical samples matter greatly to the research, and it is necessary to increase the sample size, which would be beneficial for this study. Secondly, additional adhesion-related research on this gene will enhance our understanding of ARHGAP12's clinical value, facilitating a better evaluation of its clinical significance. Moreover, the study is dependent on retrospective data and potential selection bias might exist. Increasing relevant clinical and animal experiments can offer better proof of our viewpoints.

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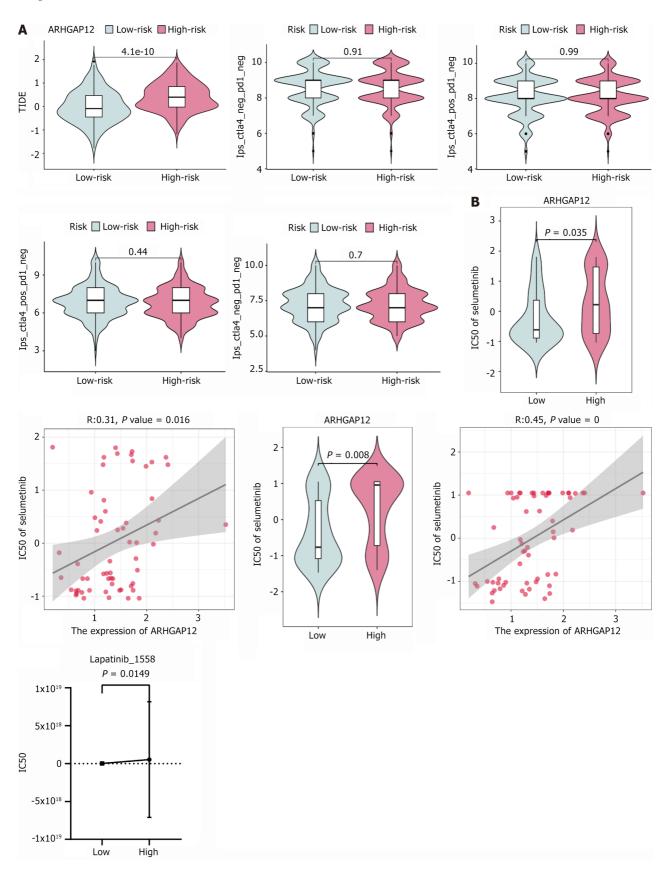


Figure 11 Sensitivity analysis of Rho GTPase activating protein 12 to tyrosine kinase inhibitors and immunotherapy. A: Relationship between Rho GTPase activating protein 12 expression and immune escape, immunotherapy; B: Relationship between Rho GTPase activating protein 12 expression and the intensity of tyrosine kinase inhibitors treatment. ARHGAP12: Rho GTPase activating protein 12.

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CONCLUSION

Our study reveals the high expression of ARHGAP12 in HCC, which may trigger TKI resistance, possibly through its regulatory role in FA. Overall, our research unveils molecular changes in HCC, linking ARHGAP12 with HCC and TKI resistance, providing new insights for the treatment of TKI-resistant HCC in the future.

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FOOTNOTES

Author contributions: Yao GP, Tang YL, Chen G, Feng ZB, and He RQ conceived and designed the study; Wang XW, Tang YX, Li FX, Wang JL, Zeng DT, Chi BT, Su QY, Huang LQ, and Qin DY performed the experiments, and analyzed the data; Wang XW, Tang YX, Li FX, Wang JL, Yao GP, Zeng DT, Chi BT, and Qin DY wrote the manuscript; Tang YL, Chen G, Feng ZB, and He RQ revised and corrected the draft; and all authors approved the final version of the article. Wang XW and Tang YX are co-first authors, contributing equally in study design, draft writing and data analysis. Wang XW also performed the immunohistochemical experiments and evaluated the corresponding results. Tang YX was responsible for computational pathology, including the calculation of standardized mean difference and summary receiver operating characteristic, as well as the analysis of related pathways. Feng ZB and He RQ are co-corresponding authors, contributing equally in designing the project, supervising various experiments and data mining, and correcting the draft.

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Conflict-of-interest statement: All the authors report no relevant conflicts of interest for this article.

Data sharing statement: Dataset available from the co-first author at tang_yx_patho@163.com. Participants gave informed consent for data sharing.

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