

Therapeutic potential of curcumin in digestive diseases

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Abstract

Curcumin is a low-molecular-weight hydrophobic polyphenol that is extracted from turmeric, which possesses a wide range of biological properties including anti-inflammatory, anti-oxidant, anti-proliferative and anti-microbial activities. Despite its diverse targets and substantial safety, clinical applications of this molecule for digestive disorders have been largely limited to case series or small clinical trials. The poor bioavailability of curcumin is likely the major hurdle for its more widespread use in humans. However, complexation of curcumin into phytosomes has recently helped to bypass this problem, as it has been demonstrated that this new lecithin formulation enables increased absorption to a level 29-fold higher than that of traditional curcuminoid products. This allows us to achieve much greater tissue substance delivery using significantly lower doses of curcumin than have been used in past clinical studies. As curcumin has already been shown to provide good therapeutic results in some small studies of both inflammatory and neoplastic bowel disorders, it is reasonable to anticipate an even greater efficacy with the advent of this new technology, which remarkably improves its bioavailability. These features are very promising and may represent a novel and effective therapeutic approach to both functional and organic digestive diseases.

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Key words: Curcumin; Curcumin-phytosome; Curcumin bioavailability; Digestive disorders

Core tip: Curcumin is a well-established molecule with multiple pharmacological activities, mainly anti-inflammatory and anti-proliferative. The major hurdle for a widespread clinical use has been represented by its poor bioavailability, which has been recently overcome by the development of a new formulation combining curcumin with phospholipids (curcumin-phytosome). This compound permits to improve markedly intestinal absorption of curcumin and guarantees a greater tissue delivery than the traditional curcuminoid mixtures. So, curcumin-phytosome has the potential to be exploited in many gastrointestinal diseases, both functional and organic.

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INTRODUCTION

In recent years, we have witnessed a shortage of certain types of drugs synthesized from chemical laboratories and a growing interest in therapeutic substances derived from natural plants. Curcumin represents one of these compounds, and this nutraceutical has already undergone many experimental and clinical studies to assess its use in the treatment of various human diseases.

This polyphenol has been shown to possess anti-inflammatory, anti-oxidant, immuno-modulatory, wound-healing, anti-proliferative and antimicrobial activities. These diverse properties, together with the fact that curcumin is innocuous, inexpensive and easily available,

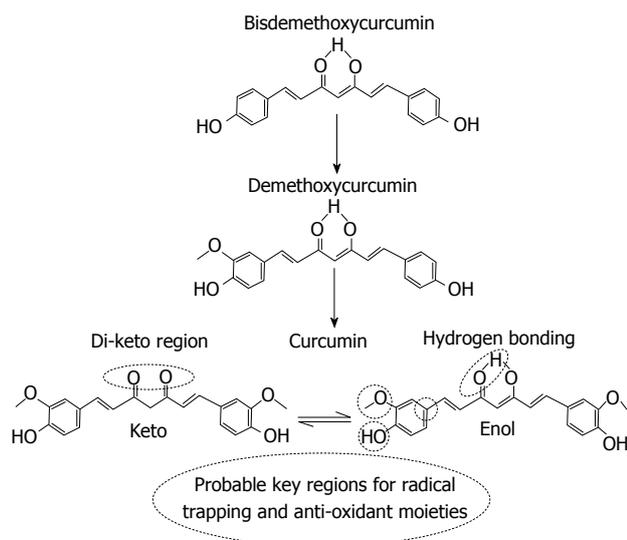


Figure 1 Proposed molecular pathway for the conversion of bisdemethoxycurcumin to demethoxycurcumin and finally to curcumin and the co-existence of keto and enol isomers of curcumin.

have sparked interest in its therapeutic application for several digestive disorders. Moreover, recent progress in the formulation of curcumin complexes with other substances, in particular with phospholipids, has remarkably increased the bioavailability of this compound, leading to greater absorption and a higher concentration in human tissues. This allows us to use lower dosages of curcumin than have been used in the past, which greatly reduces the number of tablets taken during the day while maintaining no adverse side effects.

Finally, distribution studies of curcumin in human tissues have shown that it preferentially accumulates in the intestine, colon and liver. This finding might be one major reason for the anticipation and observation of its most promising *in vivo* effects in gastrointestinal diseases when compared with other organ systems.

This review presents current knowledge of the physical and molecular properties of curcumin, its pharmacokinetics and metabolism, its mechanism of action and results of the few published clinical trials, as well as the potential therapeutic perspectives in patients with various digestive disorders.

Literature searches were performed in PubMed, Ovid, EMBASE and the Cochrane Library databases in accordance with published recommendations. We critically analyzed all full-text papers and reviews written in the English language and searched them using the terms curcumin, turmeric, colorectal cancer (CRC), inflammatory bowel diseases (IBD), functional digestive disorders, irritable bowel syndrome and liver diseases. Both animal and human studies were reviewed.

PHYSICAL AND MOLECULAR PROPERTIES OF CURCUMIN

Turmeric (the common name for *Curcuma longa*) is an In-

dian spice derived from the rhizomes of the plant and has a long history of use in Ayurvedic medicine as a treatment for inflammatory conditions^[1].

The primary active constituent of turmeric, which is responsible for its vibrant yellow color, is curcumin, which was first identified in 1910 by Lampe and Milobedzka^[2]. Curcumin exists as a bright yellow powder that provides the pigmentation of turmeric, which is used in the dye industry. Turmeric is composed of volatile oils (tumerone, atlantone, and zingiberone), sugars, proteins, resins and a group of the following three curcuminoids: about 75% curcumin (diferuloylmethane), about 16% demethoxycurcumin (DMC), about 8% bisdemethoxycurcumin (bDMC). DMC and bDMC possess similar molecular and biological properties. It is proposed that within natural pathways (Figure 1), bDMC is converted to DMC, which is then converted to curcumin^[3].

Curcumin (or diferuloylmethane) is a poly-phenolic molecule that exhibits keto-enol tautomerism and has a predominant keto form in acidic and neutral solutions and a stable enol form in alkaline medium^[4]. The molecule is lipophilic and consists of two aromatic rings connected by two unsaturated carbonyl groups; therefore, it has poor solubility in water. The molecule is stabilized by hydrogen-bonding associated with the central OH group. This may be one of the important functional sites that is responsible for the array of molecular biological activities^[5]. Curcumin is photosensitive, and precautions should be taken to avoid exposure and subsequent degradation.

PHARMACOKINETICS AND METABOLISM OF CURCUMIN

Absorption and systemic bioavailability

Over the past three decades, animal studies have shown that curcumin is hydrolytically unstable at intestinal pH, rapidly metabolized, conjugated in the liver, and excreted in the feces. Therefore, it has limited systemic bioavailability. The effects of reduced bioavailability of any agent within the body are low intrinsic activity, poor absorption, high rate of metabolism, inactivity of metabolic products and/or rapid elimination and clearance from the body. In this section, problems of limited curcumin bioavailability such as low serum levels, limited tissue distribution, apparent rapid metabolism and short half-life are described in detail.

Serum concentration

One of the major observations from curcumin studies is very low serum levels. The first reported study to examine the uptake, distribution, and excretion of curcumin was by Wahlstrom and Blennow^[6] in 1978 using Sprague-Dawley rats. Negligible amounts of curcumin in the blood plasma of rats after oral administration of 1 g/kg of curcumin showed that this molecule was poorly absorbed from the gut.

In 1980, Ravindranath *et al*^[7] showed that after oral administration of 400 mg of curcumin in rats, no curcumin

was found in the heart blood, whereas a trace amount (less than 5 µg/mL) was found in the portal blood from 15 min to 24 h after curcumin administration.

When curcumin was given orally at a dose of 2 g/kg in rats, a maximum serum concentration of 1.35 ± 0.23 µg/mL was observed after 0.83 h, whereas in humans, the same dose of curcumin resulted in either undetectable or extremely low (0.006 ± 0.005 µg/mL at 1 h) serum levels^[8].

A phase I clinical trial^[9] conducted among 25 patients with various precancerous lesions demonstrated that oral doses of 4, 6 and 8 g of curcumin administered daily for three months yielded serum curcumin concentrations of only 0.51 ± 0.11 , 0.63 ± 0.06 , and 1.77 ± 1.87 µm, respectively. This finding indicates that curcumin is poorly absorbed and may have limited systemic bioavailability. Serum levels peaked between one and two hours after administration and declined rapidly thereafter. This study did not identify curcumin metabolites, and urinary excretion of curcumin was undetectable.

Another phase I trial^[10] involving 15 patients with advanced colorectal cancer administered curcumin at doses between 0.45 and 3.6 g daily for four months. In three of six patients who were given the 3.6 g dose, the mean plasma curcumin measured after one hour on day 1 was 11.1 ± 0.6 nmol/L. This measurement remained relatively consistent at all-time points measured during the first month of curcumin therapy. The molecule was not detected in the plasma of patients taking lower doses.

A very recent study by Yang *et al.*^[11] showed that 10 mg/kg of curcumin given *iv* in rats yielded a maximum serum curcumin level of 0.36 ± 0.05 µg/mL, whereas a 50-fold higher curcumin dose administered orally yielded a maximum serum level of only 0.06 ± 0.01 µg/mL.

These studies clearly suggest that the route of administration affects achievable serum levels of curcumin, and they further indicate that the serum levels of this compound in rats and in humans are not directly comparable.

Tissue distribution

The uptake and distribution of curcumin in body tissues are obviously important factors determining its biological activity, yet a limited number of studies have addressed this issue.

Ravindranath *et al.*^[7] showed that after oral administration of 400 mg of curcumin in rats, only traces of the unchanged molecule were found in the liver and kidney. At 30 min, 90% of the curcumin was found in the stomach and small intestine, but only 1% was present at 24 h.

Another study of the same group evaluated the tissue distribution of curcumin using a tritium-labeled molecule^[12]. They found that radioactivity was detectable in the blood, liver, and kidney following doses of 40080, or 10 mg of (3H) curcumin. With 400 mg, considerable amounts of the radio-labeled products were present in tissues 12 d after dosing. The percentage of curcumin absorbed (60%-66% of the given dose) remained constant regardless of the dose, indicating that increased administration of the drug does not result in greater absorption.

Similarly, the concentrations of curcumin in normal and malignant colorectal tissue of patients receiving 3600 mg of the compound were 12.7 ± 5.7 and 7.7 ± 1.8 nmol/g, respectively, and these doses had pharmacological activity in the colorectum as measured by their effects on levels of M(1)G and cyclooxygenase-2 (COX-2) protein^[13]. Another study by the same authors showed no curcumin in the liver tissue of patients with hepatic metastases from colorectal cancer who received 450-3600 mg of curcumin daily for 1 wk prior to surgery^[14].

Metabolites

Various studies have evaluated the metabolism of curcumin in rodents and in humans. Once absorbed, curcumin is subjected to conjugations such as sulfation and glucuronidation at various tissue sites. The very first bio-distribution study reported the metabolism of the major part of curcumin orally administered in rats^[6]. The liver was indicated as the major organ responsible for metabolism of this drug^[15].

Holder *et al.*^[16] reported that the major biliary metabolites of curcumin in rats are glucuronides of tetrahydrocurcumin (THC) and hexahydrocurcumin. A minor biliary metabolite was dihydroferulic acid together with traces of ferulic acid. In addition to glucuronides, sulfate conjugates were found in the urine of curcumin-treated mice^[13].

Asai *et al.*^[17] evaluated the absorption and metabolism of orally administered curcumin in rats. The enzymatic hydrolysis of plasma samples showed that the predominant metabolites in plasma following oral administration were glucuronides/sulfates of curcumin. The plasma concentrations of conjugated curcuminoids reached a maximum at 1 h after administration. The presence of conjugative enzyme activities for glucuronidation and sulfation of curcumin in the liver, kidney and intestinal mucosa suggests that orally administered curcumin is absorbed from the alimentary tract and is present in the general blood circulation after largely being metabolized to form glucuronide/sulfate conjugates.

Whether curcumin metabolites are as active as curcumin itself is not clear^[18-20]. While most studies indicate that curcumin glucuronides and THC are less active than curcumin itself, other studies suggest that they may actually be more active than curcumin^[19-21].

Half-life

Systemic elimination or clearance of curcumin from the body is another important factor that determines its relative biological activity. Wahlstrom and Blennow^[6] reported that when 1 g/kg curcumin was given orally to rats, 75% of it was excreted in the feces, and negligible amounts were found in the urine. Intravenous (*iv*) and intraperitoneal (*ip*) administration of curcumin resulted in biliary excretion of the molecule from cannulated rats.

A clinical study of 15 patients receiving oral curcumin in doses between 36 and 180 mg daily for up to 4 mo found neither curcumin nor its metabolites in urine, but the drug was recovered from feces^[22]. The absorption and

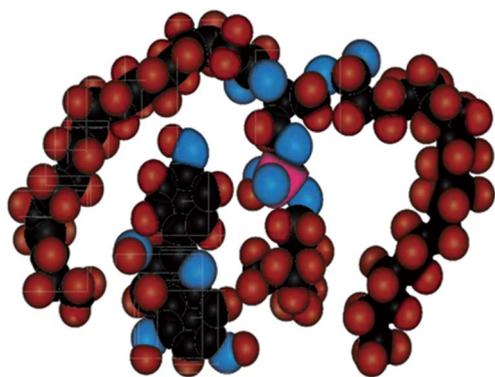


Figure 2 Phytosome molecular complex.

elimination half-lives of orally administered curcumin (2 g/kg) in rats were reported to be 0.31 ± 0.07 and 1.7 ± 0.5 h, respectively. However, in humans, the same dose of curcumin did not allow the calculation of these half-life values because the serum curcumin levels were below the detection limit at the majority of time points in most of the experimental subjects.

The existing evidence in the literature is not sufficient to make conclusions about the factors controlling the *in vivo* elimination half-life of curcumin, and future studies are warranted to address this issue.

METHODS TO OVERCOME THE TRADITIONAL LOW BIOAVAILABILITY OF CURCUMIN

Because of the above-mentioned poor bioavailability, which limits the therapeutic usefulness of curcumin, many attempts have been made to improve oral absorption of the compound^[23]. Among them, the complexation of curcumin with phospholipids using so-called phytosome technology has emerged as one of the most documented approaches from a preclinical and clinical standpoint.

Phytosome technology was developed in 1989 (Figure 2). Water-soluble phytosomes can be converted into a lipid-compatible molecular complex. Phytosomes are more available than uncomplexed products due to their enhanced capacity to cross the lipid biomembranes and to reach the systemic circulation^[24].

It is inferred that, at the intestinal level, the water-miscible phosphatidylcholine (PC) molecules enhance the dispersion of the poorly water-soluble polyphenol molecules into the water-soluble environment of the gastrointestinal lumen. PC further enhances transfer from the lumen into the lipid-soluble environment of the outer cell membrane of the epithelial absorptive cells (enterocytes). The enterocyte outer membrane has a lipid molecular bilayer that consists largely of PC. It is feasible that the PC in the phytosome merges into this PC domain of the enterocyte membrane, and by carrying the polyphenol with it, the PC “ushers” the polyphenol into the cell.

The bioavailability of the curcumin phytosome (CP)

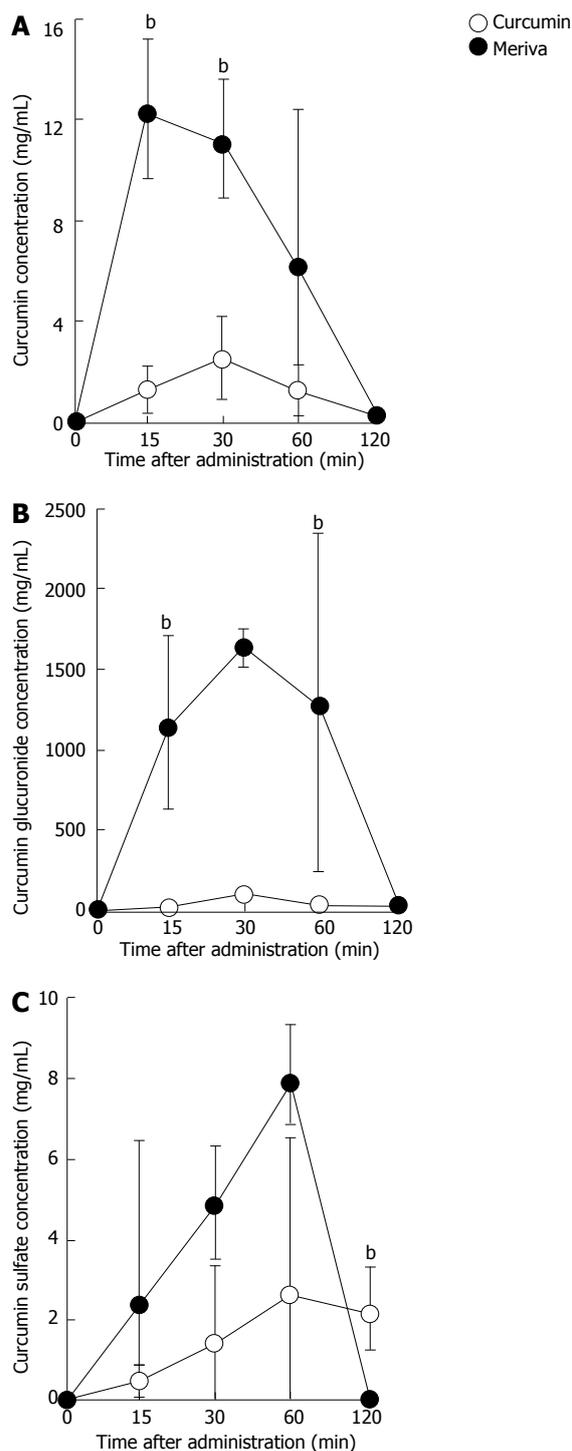


Figure 3 Plasma curcumin I in rats from curcumin phytosome or non-complexed curcumin. A: Curcumin concentration; B: Curcumin glucuronide concentration; C: Curcumin sulfate concentration. ^b*P* < 0.01 vs curcumin phytosome.

preparation (Meriva[®], Indena Spa, Milan, Italy) has been tested against an equivalent non-phytosome curcumin extract by Marczylo *et al*^[25]. These authors administered equivalent dosages (340 mg of curcumin) of curcumin or curcumin phytosome preparation to rats and reported a dramatic increase in the bioavailability among the animals that received the curcumin phytosome preparation (Figure 3). Peak plasma levels of curcumin were approximately

Table 1 Pharmacokinetics parameters in healthy volunteers after administration of curcuminphytosomes or unformulated curcumin

Curcuminoid	Formulation	AUC (ng/mL)	C _{max} (ng/mL)	t _{max} (h)	Relative absorption ²
Curcumin (1a)	Curcuminphytosome high	538.0 ± 130.7	50.3 ± 12.7	3.8 ± 0.6	19.2 ¹
	Curcuminphytosome low	272.6 ± 68.52	24.2 ± 5.9	4.2 ± 0.8	17.5 ³
	Reference	122.5 ± 29.3	9.0 ± 2.8	6.9 ± 2.2	1
Demethoxycurcumin (1b)	Curcuminphytosome high	655.0 ± 195.7	134.6 ± 40.6	2.4 ± 0.3	68.3 ⁴
	Curcuminphytosome low	297.4 ± 107.3	39.1 ± 11.4	3.1 ± 0.4	55.5 ⁴
	Reference	55.8 ± 15.5	4.2 ± 1.1	4.4 ± 1.0	1
Bisdemethoxycurcumin (1c)	Curcuminphytosome high	142.2 ± 58.2	24.9 ± 8.1	2.2 ± 0.4	56.8 ⁵
	Curcuminphytosome low	70.1 ± 34.3	8.8 ± 3.1	2.4 ± 0.6	53.1 ⁵
	Reference	24.6 ± 10.3	2.1 ± 0.8	3.4 ± 1.2	1
Total curcuminoids	Curcuminphytosome high	1336.0 ± 357.1	206.9 ± 54.9	2.7 ± 0.3	31.5 ⁶
	Curcuminphytosome low	640.2 ± 197.7	68.9 ± 16.9	3.3 ± 0.3	27.2 ⁶
	Reference	202.8 ± 53.8	14.4 ± 4.2	6.9 ± 2.2	1

¹Actual results not baseline subtracted, and errors are standard error of the mean ± SE; ²Area under the curve (AUC) normalized; ³Average: 18.3; ⁴Average: 61.9; ⁵Average: 54.1; ⁶Average: 29.14.

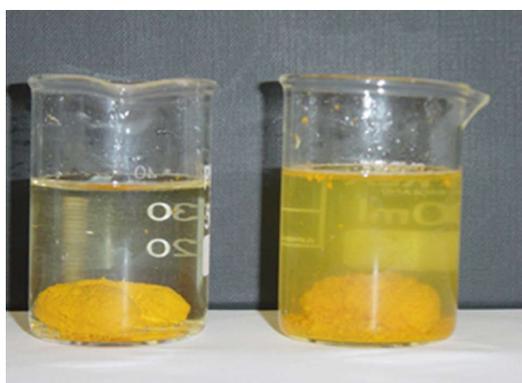


Figure 4 Image of Norflo[®] tablet in water after few seconds.

5-fold higher for CP than for traditional curcumin. Plasma levels of curcumin sulfate and curcumin glucuronide observed after the administration of CP were 3- to 20-fold higher, respectively, than those observed after the administration of uncomplexed curcumin. In the same study, significant amounts of curcumin were also measured at the tissue level and were found to have particular relevance for the liver and intestine.

More recently, Cuomo *et al*^[26] reported the results of a comparative pharmacokinetic study of healthy volunteers. In this randomized, double-blind, cross-over study, subjects received curcumin and the CP formulation at 2 dosage levels (209 and 376 total curcuminoids). The average dose-related absorption of curcumin following the 2 doses of CP was approximately 18-fold higher than the absorption of the reference curcumin. Moreover, the absorption of total curcuminoids was approximately 29-fold higher for CP in comparison with the unformulated reference, as the plasma concentration of demethoxycurcumin and bis-demethoxycurcumin from the former compound was approximately 50- to 60-fold higher than the concentration from the unformulated curcumin (Table 1).

CP is a powder that contains 20% curcumin, 40% microcrystalline cellulose and 40% phospholipids. It is utilized as an active ingredient in several food supplements in different markets. The product is available in various formulations including hard gel capsules and tablets. In

Italy, for example, the product has been developed as 500-mg tablets that combine CP with some dissolving substances (Curcsol) under the name Norflo[®] (Eyepharm, Genoa, Italy). These tablets dissolve very rapidly in the first part of the intestine, favoring the formation of an emulsion with bile acids (Figure 4), which permits almost complete absorption of phospholipids (unpublished data). The use of this formulation overcomes the risk that undissolved tablets may pass through the entire intestine and be eliminated in the feces either intact or only partially dissolved.

CP has been widely documented in several health settings, but few studies have focused on gastrointestinal disorders, which, nevertheless, seem to be a very promising therapeutic area. From this perspective, a colon-targeted delivery preparation could further optimize the clinical effects.

TOXICITY AND TOLERABILITY OF CURCUMIN

Curcumin has been reported to be safe in many human studies, and only minimal toxicity has been associated with this polyphenol^[27]. In a dose escalation study among 34 healthy volunteers, in whom the doses of curcumin ranged from 500 to 12000 mg, safety was assessed after 72 h. Only 7 subjects complained of disturbances, which were mild and included headache, skin rash, diarrhea and yellow stool^[9]. In another investigation lasting for 1-4 mo, escalating doses of curcumin from 0.45 to 3.6 g/d found rare instances of nausea and diarrhea, as well as an increase in alkaline phosphatase and LDH^[10]. Some patients treated with doses as high as 8 g/d for 2 wk reported abdominal pain and complained about the bulky volume of the tablets^[28]. As curcumin is particularly concentrated in the human liver, the risk of hepatotoxicity has been closely evaluated, but liver function tests have been shown to be unaffected with doses as high as 2-4 g/d^[29]. As one of the most documented bioavailable curcumin formulations, the CP formulation has been widely employed in the clinical setting with a daily dosage rang-

ing between 1 and 2 g, and this preparation has shown good tolerability and compliance, even in medium-term trials. However, we must stress that studies of more than 6 mo of treatment are lacking, and it is not possible to draw any firm conclusions regarding the long-term safety profile of this compound.

MECHANISMS OF ACTION AGAINST INFLAMMATORY AND NEOPLASTIC CONDITIONS

Anti-inflammatory mechanisms

Curcumin is a highly pleiotropic molecule capable of interacting with numerous molecular targets involved in inflammation. It has been proposed that this compound modulates the inflammatory response by the following mechanisms^[30,31]: (1) Down-regulation of COX-2, lipoxygenase, and inducible nitric oxide synthase (iNOS) enzymes; (2) Inhibition of the inflammatory cytokines, tumor necrosis factor-alpha (TNF- α), interleukin (IL)-1, -2, -6, -8, and -12, monocyte chemoattractant protein, and migration inhibitory protein; and (3) Down-regulation of mitogen-activated and Janus kinases.

COX-2 inhibition and iNOS inhibition are likely achieved *via* curcumin suppression of nuclear factor kappa B (NF- κ B) activation. NF- κ B is a ubiquitous eukaryotic transcription factor involved in the regulation of inflammation, cellular proliferation, transformation, and tumorigenesis^[32]. NF- κ B is not a single gene but rather a family of interrelated transcription factors that include the following five genes: NF- κ B1 (p50/p105), NF- κ B2 (p52/p100), RelA (p65), c-Rel, and RelB^[33]. The member proteins form homo- or heterodimers, of which the p50/p65 heterodimer is the most abundant and is responsible for the majority of NF- κ B canonical transcriptional activity. Generally, NF- κ B dimers associate with an inhibitory- κ B (I κ B- α) protein that keeps the dimer in the cytoplasm in an inactive state.

NF- κ B activation begins with the activation of an I κ B kinase (IKK) complex that consists of catalytic subunits IKK- α and IKK- β and the scaffolding subunit IKK- γ (the NF- κ B essential modifier)^[34]. Several mitogen-activated protein (MAP) kinases that also include NF- κ B-inducing kinase (NIK) activate IKK through the phosphorylation of IKK- α and IKK- β . IKK- β has higher activity than IKK- α for I κ B- α and is considered important in the canonical pathway.

In the canonical pathway, as shown in Figure 4, phosphorylation of I-kappa B kinase (I κ B) kinase α/β by mitogen-activated protein kinase (MAPK) is followed by phosphorylation of I κ B- α , which occurs in an inactive complex with p50/p65. Phosphorylated I κ B- α is released and degraded in the cytoplasm. The active heterodimer of p50/p65 enters the nucleus to regulate expression of multiple genes^[35].

Curcumin is thought to suppress NF- κ B activation and proinflammatory gene expression by blocking phos-

phorylation of inhibitory factor I κ B. Suppression of NF- κ B activation subsequently down-regulates COX-2 and iNOS expression, thus inhibiting the inflammatory process and tumorigenesis^[33]. In an animal model of inflammation, curcumin also inhibited arachidonic acid metabolism and inflammation in mouse skin epidermis *via* down-regulation of the cyclooxygenase and lipoxygenase pathways^[36].

In vitro studies indicate that curcumin inhibition of inflammatory cytokines is achieved through suppression of cytokine gene expression and down-regulation of intercellular signaling proteins, such as protein kinase C^[36].

Curcumin anticancer effects

There has been some promising research concerning curcumin as a safe therapeutic agent for many cancers, including colorectal cancer. This has been shown through various studies in cell cultures, animal models, and humans^[2,37].

Carcinogenesis is a complex process mainly consisting of the following three phases: initiation, promotion, and progression^[38]. There is suggestive evidence that inflammation may play a role in the three phases of carcinogenesis^[39]. Cancer initiation is produced by oxidative stress and chronic inflammation^[2]. Inflammation acts as a key regulator in the promotion of these initiated cells, possibly by providing them with proliferating signals and by preventing apoptosis^[40]. The role of inflammation in tumor induction and subsequent malignant progression has also been investigated^[41]. An inflammatory response produces cytokines, which act as growth and/or angiogenic factors, leading transformed cells to proliferate and undergo promotion. Leukocytes produce cytokines and angiogenic factors as well as matrix-degrading proteases that allow the tumor cells to proliferate, invade, and metastasize. Tumor-infiltrating lymphocytes secrete matrix-degrading proteinases such as matrix metallo-peptidase 9 (MMP-9) and thus promote neoplastic proliferation, angiogenesis, and invasion^[42].

These details demonstrate the role of inflammation in all three stages of carcinogenesis. Substantial evidence for the role of inflammation in cancer is provided by the frequent up-regulation of inflammatory mediators such as NF- κ B. The pathways activated by NF- κ B up-regulators are implicated not only in tumor growth and progression but also in the development of cancer cell resistance to anti-cancer drugs, radiation and death cytokines. NF- κ B is an excellent target for anti-cancer therapy^[43].

Effects on tumor initiation by curcumin

Curcumin has demonstrated a significant reduction in the levels of iNOS, which produces oxidative stress, which is itself one of the main causes of tumor initiation. Curcumin inhibits the induction of nitric oxide synthase and is a potent scavenger of free radicals such as nitric oxide^[44].

NF- κ B has been implicated in the induction of iNOS. Curcumin prevents phosphorylation and degradation

of inhibitor $\kappa\text{B}-\alpha$ and thereby blocks NF- κB activation, which down-regulates iNOS gene transcription^[45]. Curcumin was found to inhibit cell proliferation and cytokine production by inhibiting NF- κB target genes involved in this mitogen induction of T-cell proliferation, interleukin IL-2 production and nitric oxide generation. The over-expression of cytokines, such as IL-10, IL-6, and IL-18, is accompanied by NF- κB induction that is controlled and inhibited by curcumin^[46]. Curcumin has been shown to increase expression of conjugation enzymes (phase II), which suppress ROS-mediated NF- κB , activator protein 1 (AP-1) and MAPK activation^[47].

Tumor proliferation and progression suppression by curcumin

We have already mentioned that NF- κB has an important role in cancer initiation, promotion and progression. In addition to suppressing various cell survival and cell proliferative genes, including Bcl-2, cyclin D1, IL-6, COX-2, and MMP-9, curcumin induces apoptosis, as shown by caspase activation and poly (ADP-ribose) polymerase-cleavage^[48-50].

Curcumin is also able to block NF- κB signaling and inhibit IKK activation. The suppression of cell survival and cell proliferation genes, including Bcl-2, cyclin D1, IL-6, COX-2 and MMP, has also been noted^[48,49]. It has been suggested that COX-2 induction is mediated by the NF- κB intracellular signaling pathway, and overexpression of COX-2 leads to malignant cell proliferation and invasion^[51,52]. Curcumin inhibits COX-2 expression by repressing degradation of the inhibitory unit inhibitor $\kappa\text{B}-\alpha$ and hindering the nuclear translocation of the functionally active subunit of NF- κB , thereby blocking improper NF- κB activation^[34].

Curcumin has been found to reduce the invasion and subsequent metastasis of cancer cells. It suppresses MMP expression, which is believed to play a major role in mediating neovascularization and is increased during tumor progression^[53].

Curcumin down-regulates MMP-9 expression by inhibiting NF- κB and AP-1 binding to the DNA promoter region. MMP-9 is one of the two determinants of neovascularization that help to form new capillaries from preexisting blood vessels^[54].

Curcumin has been noted to cause significant inhibition of tumor necrosis factor α -induced VCAM-1 expression, which is related to the activation of the MAPK NF- κB pathway^[55,56]. Curcumin has been shown to reduce cell migration and invasion induced by osteopontin, an extracellular matrix protein, through the NF- κB pathway^[57].

Curcumin may inhibit cancer cell growth through down-regulation of IL-1- and IL-8-induced receptor internalization. It controls cancer progression by either blocking tumor growth or inhibiting its invasive and aggressive potential. In both cases, most of the effects are exerted by curcumin-induced NF- κB inhibition^[57].

However, curcumin has been found to arrest the cell

cycle and to induce apoptotic cell death through inhibition of the Janus family of kinase-signal transducer and activator of transcription (JAK-STAT) signaling pathway^[58].

The JAK and STAT comprise an important signaling pathway involved in dysregulation of cell growth, invasion, angiogenesis, metastasis and resistance to apoptosis^[59,60]. The JAK-STAT system consists of the following three main components: (1) A receptor; (2) JAK; and (3) STAT.

The receptor is activated by a signal from interferon, IL-6, growth factors, or other chemical messengers^[61]. This activates the kinase function of the JAKs (JAK1, JAK2, and JAK3), which autophosphorylation (phosphate groups act as “on” and “off” switches on proteins). The STAT protein then binds to the phosphorylated receptor, where STAT is phosphorylated by JAK. The phosphorylated STAT protein binds to another phosphorylated STAT protein (dimerizes) and translocates into the cell nucleus. In the nucleus, it binds to DNA and promotes transcription of genes responsive to STAT^[62-63]. Studies have evaluated the regulators of cytokine signaling including protein tyrosine phosphatases (PTPases) such as Src homology 2 (SH2) domain-containing PTPases (SHP)-1 and SHP-2. Potential roles for SHP-1 and SHP-2 have been investigated for their use in the control of cytokine signaling through the dephosphorylation of JAKs and their receptors^[64].

Of the seven STAT proteins identified thus far, only activated STAT3 and STAT5 have been implicated in multiple myeloma, lymphomas, leukemias and several solid tumors^[65]. Aberrant STAT3 signaling is an important process in the development and progression of cancer; thus, agents that block its activation have therapeutic potential. Rajasingh *et al.*^[66] have demonstrated that *in vitro*, treatment with curcumin induced a dose-dependent decrease in JAK and STAT phosphorylation, resulting in the induction of growth-arrest and apoptosis in T cell leukemia. Curcumin reversibly inhibits STAT3 activation in human multiple myeloma cells and, by this mechanism, suppresses IL-6-induced cell proliferation^[67-68]. It also inhibits STAT3 activation in five different human Hodgkin and Reed-Sternberg lymphoma cell lines^[69].

It has been shown that curcumin inhibits lysophosphatidic acid-induced IL-6 and IL-8 secretion and STAT3 phosphorylation in ovarian cancer cells^[70], and curcumin has also been shown to have a significant effect upon CRC by blocking STAT3-driven cancer cell growth^[69]. In summary, the anti-inflammatory and anticancer effects of curcumin are listed in Table 2.

CLINICAL TRIALS EXPLORING THE THERAPEUTIC POTENTIAL OF CURCUMIN IN GASTROINTESTINAL DISEASES

Because of its higher bioavailability in the gastrointestinal

Table 2 Curcumin's anti-inflammatory and anticancer effects

Anti-inflammatory effects	Anticancer effects
Downregulation of NF- κ B, Inhibition, <i>via</i> NF- κ B, of COX-2, lipoxygenase, and iNOS enzymes Inhibition of the inflammatory cytokines, such as TNF- α , interleukin (IL)-1, -2, -6, -8, and -12, MCP, and migration inhibitory protein Inhibition of PPAR-g	Inhibition of carcinogen activation Stimulation of carcinogen detoxification Suppression of pro-inflammatory signaling Inhibition of STAT Induction of cancer cell apoptosis cell cycle arrest Inhibition of angiogenesis and metastasis Modulation of oncogenes and tumor suppressor genes

TNF- α : Tumor necrosis factor- α ; MCP: Monocyte chemoattractant protein; PPAR-g: Peroxisome proliferator-activated receptor-g; iNOS: Inducible nitric oxide synthase; STAT: Signal transducer and activator of transcription; NF- κ B: Nuclear factor kappa B; COX-2: Cyclooxygenase-2.

tract than in other organs, the therapeutic potential of curcumin has been investigated in several studies of digestive diseases including IBD, CRC and hepatic fibrosis.

Inflammatory bowel disease

Idiopathic IBD comprises the following two types of chronic intestinal disorders: Crohn's disease (CD) and ulcerative colitis (UC)^[71-73]. Accumulating evidence suggests that IBD results from an inappropriate inflammatory response to intestinal microbes in a genetically susceptible host^[74]. Pathogen recognition by innate immune cells is coupled to the secretion of cytokines that inform the adaptive immune system about the nature of the pathogen and instruct naïve T cells to differentiate into the appropriate T cell subtypes required to clear the infection^[75]. Thus, naïve T cells are induced to differentiate into Th1, Th2, Th17 and/or regulatory T cells (Treg) depending on the pathogen eliciting the response^[76]. Recent studies reveal that IL-6/IL-12 family cytokines (IL-6, IL-12, IL-23, IL-27 and IL-35) play pivotal roles in these lymphocyte cell-fate decisions, and their influence on the T cell developmental program is mediated primarily through activation of an evolutionarily conserved family of latent cytoplasmic transcription factors called STATs^[77,78].

The progressive damage to the gut is characterized by an aberrant inflammatory response to components of the bacterial microflora, and Th17 cells are thought to contribute to the destruction of gut tissues by inducing secretion of the extracellular matrix-degrading enzymes MIP-3 α and IL-21. Autocrine secretion of IL-21, which perpetuates a cycle of elevated IL-21 secretion, and sustained STAT3 activation in the gut play important roles in exacerbating the disease^[79]. In addition, pSTAT3 enhances survival of the pathogenic Th17 cells by up-regulating *Bcl-2*, *Bcl-xL*, and *Mcl-1* genes^[80] and may thereby contribute to maintaining the chronic inflammatory process.

Very recently, the role of NF- κ B in IBD has been elucidated^[73]. Colon biopsies in IBD patients with active disease showed increased levels of NF- κ B p65 protein, a member of the NF- κ B family of proteins. The amount of NF- κ B p65 in the tissue samples correlated with the severity of intestinal inflammation. This increased expression of NF- κ B results in an increased ability to secrete inflammatory cytokines, such as TNF- α , IL-1, IL-6,

IL-12, and IL-23, the latter of which are directly responsible for mucosal damage in IBD. TNF- α is also able to up-regulate the production of NF- κ B, which results in a cyclical feedback loop of inflammation^[81]. Additionally, the findings that the degree of gut tissue inflammation correlates with the level of pSTAT3 in histological sections of IBD patients support a role of STAT3 and Th17 cells in IBD^[82].

Anti-inflammatory drugs, immunosuppressants, and TNF blockers are used to manage IBD. However, the high cost and adverse effects associated with these drugs encourage the use of alternative management options^[83].

Because curcumin plays a key role in the inhibition of both the activation of NF- κ B pro-inflammatory cytokines and the IL-6/STAT3 signaling pathway, it could be proposed as a novel therapeutic agent in several inflammatory diseases, such as IBD^[84]. However, to date, there have been only two human studies of curcumin in patients with IBD that have achieved encouraging results. Holt *et al.*^[85] conducted a small, open-label, pilot study of curcumin in five patients with ulcerative colitis/proctitis and five patients with Crohn's disease. Patients with ulcerative proctitis, who were currently using 5-aminosalicylic acid (5-ASA) compounds and corticosteroids (four of five patients were on corticosteroids + 5-ASA compounds), were given 550 mg curcumin twice daily for one month and then 550 mg three times daily for the second month. Patients with CD were treated with 360 mg curcumin three times daily for 1 mo followed by 360 mg four times daily for another 2 mo. All patients were assessed at baseline and after two months of curcumin administration *via* hematological, biochemical, and inflammatory analysis (C-reactive protein and erythrocyte sedimentation rate) and by sigmoidoscopy and biopsy. Subjective analysis was performed *via* a self-reported symptom diary. In the ulcerative proctitis group, all five patients had significant improvement with reductions in concomitant medications in 4 patients. Although only four of five CD patients completed the study, they also improved, as evidenced by a lowered Crohn's Disease Activity Index. There was a mean reduction of 55 points and a mean reduction in the sedimentation rate of 10 mm/h. Based on the symptom diary ($P < 0.02$), all patients improved from baseline after two months of ther-

apy, and the inflammatory markers decreased to normal limits.

Subsequently, Hanai *et al.*^[86] evaluated the use of curcumin in 89 patients with quiescent UC in a randomized, double-blind, multicenter trial. After a four-week washout period, subjects were randomly assigned to a six-month regimen of either placebo ($n = 44$) or curcumin. The treatments consisted of 1000 mg after breakfast and 1000 mg after dinner ($n = 45$) in combination with sulfasalazine (SZ) (1-3 g/d; median 2 g/d) or mesalamine (1.5-3 g/d; median 2.25 g/d).

Patients were followed during treatment and for six months after the treatment ended; they received only SZ or mesalamine during the six-month follow-up period. Of 43 patients (2 patients violated the protocol) who received curcumin, 2 relapsed during the 6 mo of therapy (4.65%), compared to 8 of 39 patients (20.51%) in the placebo group ($P = 0.040$).

Recurrence rates evaluated on the basis of intention to treat showed a significant difference between curcumin and placebo ($P = 0.049$). Furthermore, curcumin improved both the clinical activity index (CAI) ($P = 0.038$) and the endoscopic index (EI) ($P = 0.0001$), measures that are used to evaluate the morbidity associated with UC. The authors drew the following three major conclusions: (1) Curcumin had better clinical efficacy over placebo in the prevention of relapse; (2) Curcumin significantly improved the CAI and EI; and (3) Curcumin was well-tolerated.

Based on these two studies, curcumin seems to be a promising and safe therapy for maintaining remission in patients with quiescent UC as well as for improving symptoms in patients with proctitis and CD. It is evident that further rigorous randomized controlled trials in larger samples of IBD patients are needed to validate the results of the above clinical studies. Considering its effect on multiple inflammatory pathways, curcumin also has the potential to be used as a steroid-sparing induction agent in mild to moderate colitis or as an adjunct to maintain remission in patients who are losing response to immunomodulators.

Colorectal cancer

Currently, it appears that the anti-carcinogenic properties of curcumin are most likely due to its effects on multiple molecular targets, such as NF- κ B factor and AP-1. These are both major transcription factors that regulate inflammation and thus affect cell proliferation, differentiation and even apoptosis.

We have already mentioned that curcumin has been shown to affect a variety of other key players involved in carcinogenesis, such as cyclooxygenase-2, matrix metalloproteinases 2 and 9 and tumor necrosis factor α -induced vascular cell adhesion molecule.

Sharma *et al.*^[10] conducted two separate clinical trials exploring the effect of curcumin on malignancies and tumor marker levels. In the first pilot study, the pharmacokinetics and pharmacodynamics of a standardized

Curcuma extract in capsule form (Phytopharm, United Kingdom) at doses ranging from 440 to 2200 mg/d, corresponding to 36-180 mg of curcumin, were evaluated. Fifteen patients with advanced CRC refractory to standard chemotherapies received Curcuma extract daily for up to 4 mo. In one patient, measurement of a serum tumor marker revealed a decrease in carcinoembryonic antigen levels from 310 ± 15 to 175 ± 9 μ g/L after two months of treatment with 440 mg Curcuma extract. Stable disease *via* computed tomography scan was observed in five of 15 patients. Oral Curcuma extract was well-tolerated, and dose-limiting toxicity was not observed.

In the second dose-escalation study^[10], 15 patients with advanced CRC refractory to standard chemotherapies consumed capsules compatible with curcumin doses of between 0.45 and 3.6 g/d for up to 4 mo. Levels of curcumin and its metabolites in plasma, urine, and feces were analyzed. Blood and imaging tests were performed at baseline and at various points throughout the trial. A daily dose of 3.6 g of curcumin caused decreases of 62% and 57% in inducible prostaglandin E2 (PGE2) production in blood samples taken 1 h after the dose was administered on days 1 and 29, respectively. PGE2 is an end product of cyclooxygenase that has been shown to stimulate the growth of human colorectal cancer cells.

Garcea *et al.*^[14] studied curcumin levels in the colorectum and the pharmacodynamics of curcumin in 12 patients with confirmed CRC. The staging of patients was noted; 2 patients were Duke A, 3 patients were Duke B, and 7 patients were Duke C. Patients were assigned to 450, 1800 or 3600 mg of curcumin per day for 7 d prior to surgery. The recoveries of curcumin in normal and malignant colorectal tissues of patients receiving 3.6 g of curcumin were 12.7 ± 5.7 and 7.7 ± 1.8 nmol/g, respectively. Curcumin levels were highest in the normal tissue of the cecum and the ascending colon as opposed to the transverse colon, the splenic flexure and the descending colon, which suggests a local effect. The levels of M1G were also decreased by curcumin treatment in malignant colorectal tissue. COX-2 levels were undetectable in normal tissue but were detectable in malignant colorectal tissue. Curcumin was not found to modulate the expression of Cox-2 in malignant tissues. The study concluded that a daily dose of 3.6 g of curcumin is pharmacologically efficacious in CRC patients.

Curcumin has also demonstrated potential for the prevention and treatment of CRC in combination with other agents. Familial adenomatous polyposis (FAP) is an autosomal-dominant disorder characterized by hundreds of colorectal adenomas that eventually develop into CRC. One study^[87] evaluated whether the combination of curcumin and quercetin could suppress adenomas in patients with FAP. Five patients with FAP received combinations of curcumin (480 mg) and quercetin (20 mg) orally three times a day, and the number and size of polyps were assessed at baseline and after therapy. Four patients had a retained rectum, and one had an ileoanal anastomosis. After 6 mo of combination treatment, all five patients had a

decrease in the number and size of polyps from baseline. Polyp number decreased by a mean of 60.4% ($P < 0.05$), and polyp size decreased by a mean of 50.9% ($P < 0.05$). This is the first human demonstration of the reduction in size and number of ileal and rectal polyps in patients with FAP by a curcumin-containing agent. Although the combinations seemed to reduce the adenomas, randomized controlled trials are needed to further validate these findings.

In a non-randomized, open-label clinical trial, Carroll *et al.*^[28] assessed the effects of oral curcumin (2 or 4 g per day for 30 d) on PGE2 within abnormal crypt foci (ACF) as the primary endpoint using 5-hydroxyeicosatetraenoic acid (5-HETE), ACF number, and proliferation in 44 eligible smokers with eight or more ACF on screening colonoscopy. They assessed pre- and post-treatment concentrations of PGE2 and 5-HETE by liquid chromatography tandem mass spectroscopy in ACF and normal-tissue biopsies; ACF number *via* rectal endoscopy; proliferation by Ki-67 immunohistochemistry; and curcumin concentrations by high-performance liquid chromatography in serum and rectal mucosal samples. Forty-one subjects completed the study. A significant 40% reduction in ACF number occurred with the 4-g dose ($P < 0.005$), whereas ACF were not reduced in the 2-g group. The ACF reduction in the 4-g group was associated with a significant, five-fold increase in post-treatment plasma curcumin/conjugate levels (*vs* pretreatment, $P = 0.009$).

In summary, the above studies suggest that curcumin is safe and has bright prospects for the treatment of patients with CRC. In fact, curcumin has been shown to be beneficial in all 3 stages of carcinogenesis and in all multifactorial illnesses such as cancer. An agent that acts at a number of different cellular levels offers the potential for effective prophylaxis and treatment. It is hoped that larger and methodologically sound clinical trials in patients with CRC will lead to the consideration of curcumin as an anticancer agent.

Liver disease

We are still remote from having available and effective drug therapies in hepatic diseases, with the exception of those with viral etiology. Especially in emerging liver diseases, such as non-alcoholic fatty liver disease (NAFLD), the only currently available therapies that have proven to be effective are those with nutritional agents such as vitamin E or those that are associated with antidiabetic drugs^[88,89]. The only effective therapy for NAFLD/NASH remains non-pharmacological and involves a multidisciplinary treatment based not only on diet but also on frequent aerobic physical activity. In this scenario, curcumin appears to provide an opportunity to cure or improve liver pathologies. Curcumin has the following 4 basic effects on the hepatobiliary system^[90]: (1) Choleric-cholagogue; (2) Antifibrotic; (3) Hepatoprotective; and (4) Antioxidant.

Choleric-cholagogue effect

Experimental studies have shown large hepatoprotective

effects for curcumin against a variety of hepatotoxic endogenous (from cholestasis to fatty infiltration) and exogenous insults (alcohol to xenobiotics), a significant percentage of which may progress to cirrhosis or hepatocellular carcinoma^[91,92]. In pharmacological terms, curcumin is a complete choleric-cholagogue. The cleavage products of curcumin (feluric and hydrofeluric acids) have cholecistokinetic properties because they squeeze the gallbladder, while another principle product, paratolilmethylcarbinol, has strong choleric activity^[15].

The choleric effect of curcumin increases bile production by approximately 62%. Its effect is not limited to the stimulation of contraction and is also expressed in the bile composition. Indeed, it has been reported that sodium curcumin increases the excretion of bile salts, cholesterol, and conjugate bilirubin, which increases the solubility and prevents the formation of stones in the gallbladder^[93].

Antifibrotic effect

Curcumin may attenuate hepatic fibrosis induced experimentally by various pathogenetic mechanisms due to its protective effect on the inhibition of tissue growth factor TGF- β ^[94]. In the development of liver fibrosis, this profibrogenic cytokine plays a key role by promoting the activation of stellate cells to myofibroblasts and through the production of extracellular matrix.

TGF- β is one of the main targets of curcumin, which likely occurs through NF- κ B^[86]. A second target of the antifibrotic effect of curcumin is its effect on metalloproteinases, which are involved in remodeling the extracellular matrix. In an experimental model of cirrhosis, curcumin normalized some parameters (ALT, glutathione, glycogen), thus signifying a resumption of hepatic metabolism. Other parameters, such as fibrosis, were only attenuated, which is likely due to the activation of metalloproteinases by curcumin itself^[95,96].

Hepatoprotective effect

Table 3 shows some examples of the hepatoprotective effects of curcumin against many hepatotoxic insults such as paracetamol, *Aspergillus aflatoxins*, or nitrosamines. The hepatoprotective effect derives mainly from its antioxidant activities, as well as its ability to reduce the formation of pro-inflammatory cytokines^[97].

Given its hepatoprotective effects, curcumin can be used in cases of insult by exogenous toxins derived from both the environment and lifestyle. It should be recalled that curcumin is able to induce the synthesis of phase II enzymes that protect cells from oxidative stress, such as glutathione transferase, heme-oxygenase and the NAPH-quinone reductase, which results in detoxification and reduced stress^[98].

Antioxidant effect

Curcumin is characterized by its high antioxidant activity, which is comparable to, if not higher than, that of vitamin C and is more than ten times higher than the activity of the scavenger vitamin E^[99].

Table 3 Substances and hepatic intoxication mechanisms contrasted by curcumin

Intoxications	Pathogenetic mechanisms	Curcumin effects
Iron (alcoholic liver disease; steatosis, viral hepatitis; anemia)	Fibrosis induced by oxidation	Anti-oxidant enzymatic activity
Alcohol (chronic or acute intoxication)	Phospholipase A2 activation	Phospholipase A2 inhibition by NF-κB
High-fat diet (lipid storage)	Focal degeneration, micronecrosis	Acil-CoA, cholesterol biliary acids; LDL peroxidation
Xenobiotics induced acute damage	ROS, lipid peroxidation, inflammation	Scavenger activity on NF-κB; anti-oxidant enzymatic activity
Xenobiotics induced chronic damage	Inflammation and hepatocellular necrosis	Hepatic fibrosis inhibition by NF-κB
Poisons (carbon tetrachloride)	Inflammatory self-maintenance	Hepatic inflammation inhibition by NF-κB

ROS: Reactive oxygen species; LDL: Low density lipoprotein; NF-κB: Nuclear factor kappa B.

Overall, the antioxidant action, especially towards cells subjected to increased oxidative stress such as hepatocytes, results in an increase of cellular resistance to oxidative damage for at least 18 h^[99]. The antioxidant properties of curcumin reside in the same chemical structure. Numerous natural antioxidants can be classified into the following two types of compounds: phenolic (sesame extract) and β-diketonic (extracts of eucalyptus).

Curcumin is one of the few antioxidants that possess both a phenolic group and one diketonic in the same molecule. This explains why curcumin possesses the ability to interrupt the chain that transmits the oxidation of biological structures until the oxidant energy is sufficient^[99].

In summary, the multiple positive effects of curcumin on both the biliary system and on liver structure and function encourage its clinical use, which needs to be validated in future controlled clinical trials.

Functional digestive disorders

The mechanisms of symptom generation in patients with functional digestive disorders are poorly understood due to the lack of a mucosal injury that enables us to explain their troublesome disturbances^[100]. Recent studies have shown that transient receptor potential vanilloid type 1 (TRPV1) receptors play a critical role in somatic and visceral nociceptive neural detection and transmission^[101], and they have been implicated in the induction of symptoms in these diseases. TRPV1 is a polymodal sensory transducer that can be activated by multiple noxious stimuli such as heat, low pH, and endogenous lipid derivatives such as anandamide as well as by exogenous substances that possess a vanilloid moiety such as capsaicin^[102]. Of remarkable importance, the curcumin molecule has the same vanilloid ring moiety as capsaicin, making TRPV1 its likely target, and it has been shown in animals that curcumin blocks TRPV1 activation by capsaicin in a competitive manner^[103]. It has been suggested that up-regulation of TRPV1 signaling may contribute to visceral hypersensitivity in functional gastrointestinal diseases, including esophageal hypersensitivity^[104]. This condition can be found in more than 50% of patients with non-erosive reflux disease, which represents the most frequent form of gastro-esophageal reflux disease^[105]. Recent epidemiological studies have shown that the rate of reflux patients with negative endoscopy can be as high as 75%^[106]. This relevant population contains subgroups of

patients with hypersensitive esophagus to both acid and non-acid reflux or patients with functional heartburn, who are difficult to treat with antisecretory therapies and who therefore may benefit from drugs that are able to act on TRPV1 receptors. In fact, curcumin has been shown to antagonize the vanilloid receptors even at low dosages and thus has the potential to modulate the response of TRPV1 to various stimulants and to prevent the generation of symptoms in patients with hypersensitive esophagus and functional heartburn^[103].

Moreover, the TRPV1 receptors are widely expressed in the entire gastrointestinal tract and enteric nervous system, and there is evidence that curcumin can inhibit GI nociception and reverse gut hypersensitivity by acting on peripheral terminals. Taking into account this mechanism of action, it cannot be excluded that this molecule may be beneficial in treating patients with functional dyspepsia and irritable bowel syndrome, which are disorders that remain clinically challenging in the setting of current drugs and whose patients may benefit from the pharmacological properties of curcumin on TRPV1 as a novel pain modulator.

Finally, as it has been shown that low-grade inflammation of the intestinal mucosa is responsible for symptoms of irritable bowel syndrome^[107], we cannot exclude that the well-known anti-inflammatory effects of curcumin may also improve the quality of life of patients with this disease.

CONCLUSION

In summary, curcumin is a well-known molecule with multiple pharmacological activities that have the potential to be used to treat many gastrointestinal diseases, both functional and organic. It appears to be a very promising therapeutic compound on the basis of thousands of pre-clinical studies, but its poor bioavailability has greatly hampered more widespread clinical use. However, the new formulation of curcumin with phospholipids has allowed us to overcome this problem by markedly improving intestinal absorption compared with the traditional unformulated curcuminoid mixtures. If curcumin is truly beneficial, as has been suggested by prior clinical trials using curcumin with limited bioavailability, we can expect to see greater therapeutic effectiveness from phospholipid-complexed curcumin, which enables increased absorption

and appropriate tissue delivery. These improved pharmacokinetic and pharmacodynamic properties are also able to significantly reduce the required dosages of curcumin and to increase the compliance of the product. Overall, these features make curcumin a very promising new therapeutic option for the treatment of gastrointestinal and hepatic diseases for which present therapies are largely unsatisfactory.

REFERENCES

- 1 **Ammon HP**, Wahl MA. Pharmacology of *Curcuma longa*. *Planta Med* 1991; **57**: 1-7 [PMID: 2062949 DOI: 10.1055/s-2006-960004]
- 2 **Aggarwal BB**, Kumar A, Bharti AC. Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res* 2003; **23**: 363-398 [PMID: 12680238]
- 3 **Kita T**, Imai S, Sawada H, Kumagai H, Seto H. The biosynthetic pathway of curcuminoid in turmeric (*Curcuma longa*) as revealed by ¹³C-labeled precursors. *Biosci Biotechnol Biochem* 2008; **72**: 1789-1798 [PMID: 18603793 DOI: 10.1271/bbb.80075]
- 4 **Balasubramanian K**. Molecular orbital basis for yellow curry spice curcumin's prevention of Alzheimer's disease. *J Agric Food Chem* 2006; **54**: 3512-3520 [PMID: 19127718 DOI: 10.1021/jf0603533]
- 5 **Priyadarsini KI**, Maity DK, Naik GH, Kumar MS, Unnikrishnan MK, Satav JG, Mohan H. Role of phenolic O-H and methylene hydrogen on the free radical reactions and antioxidant activity of curcumin. *Free Radic Biol Med* 2003; **35**: 475-484 [PMID: 12927597 DOI: 10.1016/S0891-5849(03)00325-3]
- 6 **Wahlström B**, Blennow G. A study on the fate of curcumin in the rat. *Acta Pharmacol Toxicol (Copenh)* 1978; **43**: 86-92 [PMID: 696348 DOI: 10.1111/j.1600-0773.1978.tb02240.x]
- 7 **Ravindranath V**, Chandrasekhara N. Absorption and tissue distribution of curcumin in rats. *Toxicology* 1980; **16**: 259-265 [PMID: 7423534 DOI: 10.1016/0300-483X(80)90122-5]
- 8 **Shoba G**, Joy D, Joseph T, Majeed M, Rajendran R, Srinivas PS. Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Med* 1998; **64**: 353-356 [PMID: 9619120 DOI: 10.1055/s-2006-957450]
- 9 **Cheng AL**, Hsu CH, Lin JK, Hsu MM, Ho YF, Shen TS, Ko JY, Lin JT, Lin BR, Ming-Shiang W, Yu HS, Jee SH, Chen GS, Chen TM, Chen CA, Lai MK, Pu YS, Pan MH, Wang YJ, Tsai CC, Hsieh CY. Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or premalignant lesions. *Anticancer Res* 2001; **21**: 2895-2900 [PMID: 11712783]
- 10 **Sharma RA**, Euden SA, Platton SL, Cooke DN, Shafayat A, Hewitt HR, Marczylo TH, Morgan B, Hemingway D, Plummer SM, Pirmohamed M, Gescher AJ, Steward WP. Phase I clinical trial of oral curcumin: biomarkers of systemic activity and compliance. *Clin Cancer Res* 2004; **10**: 6847-6854 [PMID: 15501961 DOI: 10.1158/1078-0432.CCR-04-0744]
- 11 **Yang KY**, Lin LC, Tseng TY, Wang SC, Tsai TH. Oral bioavailability of curcumin in rat and the herbal analysis from *Curcuma longa* by LC-MS/MS. *J Chromatogr B Analyt Technol Biomed Life Sci* 2007; **853**: 183-189 [PMID: 17400527 DOI: 10.1016/j.jchromb.2007.03.010]
- 12 **Ravindranath V**, Chandrasekhara N. Metabolism of curcumin--studies with [³H]curcumin. *Toxicology* 1981; **22**: 337-344 [PMID: 7342372 DOI: 10.1016/0300-483X(81)90027-5]
- 13 **Garcea G**, Jones DJ, Singh R, Dennison AR, Farmer PB, Sharma RA, Steward WP, Gescher AJ, Berry DP. Detection of curcumin and its metabolites in hepatic tissue and portal blood of patients following oral administration. *Br J Cancer* 2004; **90**: 1011-1015 [PMID: 14997198 DOI: 10.1038/sj.bjc.6601623]
- 14 **Garcea G**, Berry DP, Jones DJ, Singh R, Dennison AR, Farmer PB, Sharma RA, Steward WP, Gescher AJ. Consumption of the putative chemopreventive agent curcumin by cancer patients: assessment of curcumin levels in the colorectum and their pharmacodynamic consequences. *Cancer Epidemiol Biomarkers Prev* 2005; **14**: 120-125 [PMID: 15668484]
- 15 **Hoehle SI**, Pfeiffer E, Sólyom AM, Metzler M. Metabolism of curcuminoids in tissue slices and subcellular fractions from rat liver. *J Agric Food Chem* 2006; **54**: 756-764 [PMID: 16448179 DOI: 10.1021/jf058146a]
- 16 **Holder GM**, Plummer JL, Ryan AJ. The metabolism and excretion of curcumin (1,7-bis-(4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione) in the rat. *Xenobiotica* 1978; **8**: 761-768 [PMID: 726520 DOI: 10.3109/00498257809069589]
- 17 **Asai A**, Miyazawa T. Occurrence of orally administered curcuminoid as glucuronide and glucuronide/sulfate conjugates in rat plasma. *Life Sci* 2000; **67**: 2785-2793 [PMID: 11105995 DOI: 10.1016/S0024-3205(00)00868-7]
- 18 **Sandur SK**, Pandey MK, Sung B, Ahn KS, Murakami A, Sethi G, Limtrakul P, Badmaev V, Aggarwal BB. Curcumin, demethoxycurcumin, bisdemethoxycurcumin, tetrahydrocurcumin and turmerones differentially regulate anti-inflammatory and anti-proliferative responses through a ROS-independent mechanism. *Carcinogenesis* 2007; **28**: 1765-1773 [PMID: 17522064 DOI: 10.1093/carcin/bgm123]
- 19 **Pfeiffer E**, Hoehle SI, Walch SG, Riess A, Sólyom AM, Metzler M. Curcuminoids form reactive glucuronides in vitro. *J Agric Food Chem* 2007; **55**: 538-544 [PMID: 17227090 DOI: 10.1021/jf0623283]
- 20 **Kim JM**, Araki S, Kim DJ, Park CB, Takasuka N, Baba-Toriyama H, Ota T, Nir Z, Khachik F, Shimidzu N, Tanaka Y, Osawa T, Uraji T, Murakoshi M, Nishino H, Tsuda H. Chemopreventive effects of carotenoids and curcumins on mouse colon carcinogenesis after 1,2-dimethylhydrazine initiation. *Carcinogenesis* 1998; **19**: 81-85 [PMID: 9472697 DOI: 10.1093/carcin/19.1.81]
- 21 **Pari L**, Murugan P. Tetrahydrocurcumin: effect on chloroquine-mediated oxidative damage in rat kidney. *Basic Clin Pharmacol Toxicol* 2006; **99**: 329-334 [PMID: 17076682 DOI: 10.1111/j.1742-7843.2006.pto_503.x]
- 22 **Sharma RA**, McLelland HR, Hill KA, Ireson CR, Euden SA, Manson MM, Pirmohamed M, Marnett LJ, Gescher AJ, Steward WP. Pharmacodynamic and pharmacokinetic study of oral *Curcuma* extract in patients with colorectal cancer. *Clin Cancer Res* 2001; **7**: 1894-1900 [PMID: 11448902]
- 23 **Bisht S**, Maitra A. Systemic delivery of curcumin: 21st century solutions for an ancient conundrum. *Curr Drug Discov Technol* 2009; **6**: 192-199 [PMID: 19496751 DOI: 10.2174/157016309789054933]
- 24 **Semalty A**, Semalty M, Rawat MS, Franceschi F. Supramolecular phospholipids-polyphenolics interactions: the PHY-TOSOME strategy to improve the bioavailability of phytochemicals. *Fitoterapia* 2010; **81**: 306-314 [PMID: 19919847 DOI: 10.1016/j.fitote.2009.11.001]
- 25 **Marczylo TH**, Verschoyle RD, Cooke DN, Morazzoni P, Steward WP, Gescher AJ. Comparison of systemic availability of curcumin with that of curcumin formulated with phosphatidylcholine. *Cancer Chemother Pharmacol* 2007; **60**: 171-177 [PMID: 17051370 DOI: 10.1007/s00280-006-0355-x]
- 26 **Cuomo J**, Appendino G, Dern AS, Schneider E, McKinnon TP, Brown MJ, Togni S, Dixon BM. Comparative absorption of a standardized curcuminoid mixture and its lecithin formulation. *J Nat Prod* 2011; **74**: 664-669 [PMID: 21413691 DOI: 10.1021/np1007262]
- 27 **Lao CD**, Ruffin MT, Normolle D, Heath DD, Murray SI, Bailey JM, Boggs ME, Crowell J, Rock CL, Brenner DE. Dose escalation of a curcuminoid formulation. *BMC Complement Altern Med* 2006; **6**: 10 [PMID: 16545122 DOI: 10.1186/1472-6882-6-10]

- 28 **Carroll RE**, Benya RV, Turgeon DK, Vareed S, Neuman M, Rodriguez L, Kakarala M, Carpenter PM, McLaren C, Meyskens FL, Brenner DE. Phase IIa clinical trial of curcumin for the prevention of colorectal neoplasia. *Cancer Prev Res (Phila)* 2011; **4**: 354-364 [PMID: 21372035 DOI: 10.1158/1940-6207.CAPR-10-0098]
- 29 **Goel A**, Kunnumakkara AB, Aggarwal BB. Curcumin as „Curcumin“: from kitchen to clinic. *Biochem Pharmacol* 2008; **75**: 787-809 [PMID: 17900536]
- 30 **Abe Y**, Hashimoto S, Horie T. Curcumin inhibition of inflammatory cytokine production by human peripheral blood monocytes and alveolar macrophages. *Pharmacol Res* 1999; **39**: 41-47 [PMID: 10051376 DOI: 10.1006/phrs.1998.0404]
- 31 **Surh YJ**, Chun KS, Cha HH, Han SS, Keum YS, Park KK, Lee SS. Molecular mechanisms underlying chemopreventive activities of anti-inflammatory phytochemicals: down-regulation of COX-2 and iNOS through suppression of NF-kappa B activation. *Mutat Res* 2001; **480-481**: 243-268 [PMID: 11506818]
- 32 **Satou R**, Miyata K, Katsurada A, Navar LG, Kobori H. Tumor necrosis factor- α suppresses angiotensinogen expression through formation of a p50/p50 homodimer in human renal proximal tubular cells. *Am J Physiol Cell Physiol* 2010; **299**: C750-C759 [PMID: 20592241 DOI: 10.1152/ajpcell.00078.2010]
- 33 **Brasier AR**. The NF-kappaB regulatory network. *Cardiovasc Toxicol* 2006; **6**: 111-130 [PMID: 17303919 DOI: 10.1385/CT:6:2:111]
- 34 **Jobin C**, Bradham CA, Russo MP, Juma B, Narula AS, Brenner DA, Sartor RB. Curcumin blocks cytokine-mediated NF-kappa B activation and proinflammatory gene expression by inhibiting inhibitory factor I-kappa B kinase activity. *J Immunol* 1999; **163**: 3474-3483 [PMID: 10477620]
- 35 **Huang MT**, Lysz T, Ferraro T, Abidi TF, Laskin JD, Conney AH. Inhibitory effects of curcumin on in vitro lipoxygenase and cyclooxygenase activities in mouse epidermis. *Cancer Res* 1991; **51**: 813-819 [PMID: 1899046]
- 36 **Anand P**, Sundaram C, Jhurani S, Kunnumakkara AB, Aggarwal BB. Curcumin and cancer: an “old-age” disease with an “age-old” solution. *Cancer Lett* 2008; **267**: 133-164 [PMID: 18462866 DOI: 10.1016/j.canlet.2008.03.025]
- 37 **Nowell PC**. Tumor progression: a brief historical perspective. *Semin Cancer Biol* 2002; **12**: 261-266 [PMID: 12147207 DOI: 10.1016/S1044-579X(02)00012-3]
- 38 **Balkwill F**, Charles KA, Mantovani A. Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer Cell* 2005; **7**: 211-217 [PMID: 15766659]
- 39 **Carlier-Mercier L**, Fontaine JL, Boccon-Gibod L, Girardet JP, Josset P, Gouraud F, Pinturier MF. A rare cause of neonatal exudative enteropathy: congenital Langerhans cell histiocytosis (histiocytosis X). *Arch Fr Pediatr* 1992; **49**: 199-201 [PMID: 1610278 DOI: 10.1016/j.mrfmmm.2005.04.019]
- 40 **Philip M**, Rowley DA, Schreiber H. Inflammation as a tumor promoter in cancer induction. *Semin Cancer Biol* 2004; **14**: 433-439 [PMID: 15489136 DOI: 10.1016/j.semcancer.2004.06.006]
- 41 **St-Pierre Y**, Couillard J, Van Themsche C. Regulation of MMP-9 gene expression for the development of novel molecular targets against cancer and inflammatory diseases. *Expert Opin Ther Targets* 2004; **8**: 473-489 [PMID: 15469396 DOI: 10.1517/14728222.8.5.473]
- 42 **Luo JL**, Kamata H, Karin M. IKK/NF-kappaB signaling: balancing life and death—a new approach to cancer therapy. *J Clin Invest* 2005; **115**: 2625-2632 [PMID: 16200195 DOI: 10.1172/JCI26322]
- 43 **Sumanont Y**, Murakami Y, Tohda M, Vajragupta O, Matsumoto K, Watanabe H. Evaluation of the nitric oxide radical scavenging activity of manganese complexes of curcumin and its derivative. *Biol Pharm Bull* 2004; **27**: 170-173 [PMID: 14758027 DOI: 10.1248/bpb.27.170]
- 44 **Gao X**, Kuo J, Jiang H, Deeb D, Liu Y, Divine G, Chapman RA, Dulchavsky SA, Gautam SC. Immunomodulatory activity of curcumin: suppression of lymphocyte proliferation, development of cell-mediated cytotoxicity, and cytokine production in vitro. *Biochem Pharmacol* 2004; **68**: 51-61 [PMID: 15183117 DOI: 10.1016/j.bcp.2004.03.015]
- 45 **Grandjean-Laquerriere A**, Gangloff SC, Le Naour R, Trentesaux C, Hornebeck W, Guenounou M. Relative contribution of NF-kappaB and AP-1 in the modulation by curcumin and pyrrolidine dithiocarbamate of the UVB-induced cytokine expression by keratinocytes. *Cytokine* 2002; **18**: 168-177 [PMID: 12126654 DOI: 10.1006/cyto.2002.0888]
- 46 **Feng R**, Lu Y, Bowman LL, Qian Y, Castranova V, Ding M. Inhibition of activator protein-1, NF-kappaB, and MAPKs and induction of phase 2 detoxifying enzyme activity by chlorogenic acid. *J Biol Chem* 2005; **280**: 27888-27895 [PMID: 15944151 DOI: 10.1074/jbc.M503347200]
- 47 **Aggarwal S**, Takada Y, Singh S, Myers JN, Aggarwal BB. Inhibition of growth and survival of human head and neck squamous cell carcinoma cells by curcumin via modulation of nuclear factor-kappaB signaling. *Int J Cancer* 2004; **111**: 679-692 [PMID: 15252836 DOI: 10.1002/ijc.20333]
- 48 **Notarbartolo M**, Poma P, Perri D, Dusonchet L, Cervello M, D'Alessandro N. Antitumor effects of curcumin, alone or in combination with cisplatin or doxorubicin, on human hepatic cancer cells. Analysis of their possible relationship to changes in NF-kB activation levels and in IAP gene expression. *Cancer Lett* 2005; **224**: 53-65 [PMID: 15911101 DOI: 10.1016/j.canlet.2004.10.051]
- 49 **Lee JS**, Surh YJ. Nrf2 as a novel molecular target for chemoprevention. *Cancer Lett* 2005; **224**: 171-184 [PMID: 15914268 DOI: 10.1016/j.canlet.2004.09.042]
- 50 **Kim JH**, Lee KW, Lee MW, Lee HJ, Kim SH, Surh YJ. Hirsutenone inhibits phorbol ester-induced upregulation of COX-2 and MMP-9 in cultured human mammary epithelial cells: NF-kappaB as a potential molecular target. *FEBS Lett* 2006; **580**: 385-392 [PMID: 16380122 DOI: 10.1016/j.febslet.2005.12.015]
- 51 **Goel A**, Boland CR, Chauhan DP. Specific inhibition of cyclooxygenase-2 (COX-2) expression by dietary curcumin in HT-29 human colon cancer cells. *Cancer Lett* 2001; **172**: 111-118 [PMID: 11566484 DOI: 10.1016/S0304-3835(01)00655-3]
- 52 **John A**, Tuszynski G. The role of matrix metalloproteinases in tumor angiogenesis and tumor metastasis. *Pathol Oncol Res* 2001; **7**: 14-23 [PMID: 11349215 DOI: 10.1007/BF03032599]
- 53 **Woo MS**, Jung SH, Kim SY, Hyun JW, Ko KH, Kim WK, Kim HS. Curcumin suppresses phorbol ester-induced matrix metalloproteinase-9 expression by inhibiting the PKC to MAPK signaling pathways in human astrogloma cells. *Biochem Biophys Res Commun* 2005; **335**: 1017-1025 [PMID: 16102725 DOI: 10.1016/j.bbrc.2005.07.174]
- 54 **Philip S**, Bulbule A, Kundu GC. Matrix metalloproteinase-2: mechanism and regulation of NF-kappaB-mediated activation and its role in cell motility and ECM-invasion. *Glycoconj J* 2004; **21**: 429-441 [PMID: 15750784 DOI: 10.1007/s10719-004-5533-7]
- 55 **O'Hanlon DM**, Fitzsimons H, Lynch J, Tormey S, Malone C, Given HF. Soluble adhesion molecules (E-selectin, ICAM-1 and VCAM-1) in breast carcinoma. *Eur J Cancer* 2002; **38**: 2252-2257 [PMID: 12441261 DOI: 10.1016/S0959-8049(02)00218-6]
- 56 **Lee CW**, Lin WN, Lin CC, Luo SF, Wang JS, Pouyssegur J, Yang CM. Transcriptional regulation of VCAM-1 expression by tumor necrosis factor-alpha in human tracheal smooth muscle cells: involvement of MAPKs, NF-kappaB, p300, and histone acetylation. *J Cell Physiol* 2006; **207**: 174-186 [PMID: 16288471 DOI: 10.1002/jcp.20549]
- 57 **Thangapazham RL**, Sharma A, Maheshwari RK. Multiple molecular targets in cancer chemoprevention by curcumin.

- AAPS J 2006; **8**: E443-E449 [PMID: 17025261 DOI: 10.1208/aapsj080352]
- 58 **Radhakrishna Pillai G**, Srivastava AS, Hassanein TI, Chauhan DP, Carrier E. Induction of apoptosis in human lung cancer cells by curcumin. *Cancer Lett* 2004; **208**: 163-170 [PMID: 15142674 DOI: 10.1016/j.canlet.2]
- 59 **Aggarwal BB**, Sethi G, Ahn KS, Sandur SK, Pandey MK, Kunnumakkara AB, Sung B, Ichikawa H. Targeting signal-transducer-and-activator-of-transcription-3 for prevention and therapy of cancer: modern target but ancient solution. *Ann N Y Acad Sci* 2006; **1091**: 151-169 [PMID: 17341611 DOI: 10.1196/annals.1378.063]
- 60 **Darnell JE**. STATs and gene regulation. *Science* 1997; **277**: 1630-1635 [PMID: 9287210 DOI: 10.1126/science.277.5332.1630]
- 61 **Dell'Albani P**, Santangelo R, Torrisi L, Nicoletti VG, de Vellis J, Giuffrida Stella AM. JAK/STAT signaling pathway mediates cytokine-induced iNOS expression in primary astroglial cell cultures. *J Neurosci Res* 2001; **65**: 417-424 [PMID: 11536325 DOI: 10.1002/jnr.1169]
- 62 **Kovarik P**, Mangold M, Ramsauer K, Heidari H, Steinborn R, Zotter A, Levy DE, Müller M, Decker T. Specificity of signaling by STAT1 depends on SH2 and C-terminal domains that regulate Ser727 phosphorylation, differentially affecting specific target gene expression. *EMBO J* 2001; **20**: 91-100 [PMID: 11226159 DOI: 10.1093/emboj/20.1.91]
- 63 **Cull VS**, Tilbrook PA, Bartlett EJ, Brekalo NL, James CM. Type I interferon differential therapy for erythroleukemia: specificity of STAT activation. *Blood* 2003; **101**: 2727-2735 [PMID: 12446459 DOI: 10.1182/blood-2002-05-1521]
- 64 **Krebs DL**, Hilton DJ. SOCS proteins: negative regulators of cytokine signaling. *Stem Cells* 2001; **19**: 378-387 [PMID: 11553846 DOI: 10.1634/stemcells.19-5-378]
- 65 **Shishodia S**, Chaturvedi MM, Aggarwal BB. Role of curcumin in cancer therapy. *Curr Probl Cancer* 2007; **31**: 243-305 [PMID: 17645940 DOI: 10.1016/j.currprobcancer.2007.04.001]
- 66 **Rajasingh J**, Raikwar HP, Muthian G, Johnson C, Bright JJ. Curcumin induces growth-arrest and apoptosis in association with the inhibition of constitutively active JAK-STAT pathway in T cell leukemia. *Biochem Biophys Res Commun* 2006; **340**: 359-368 [PMID: 16364242 DOI: 10.1016/j.bbrc.2005.12.014]
- 67 **Bharti AC**, Donato N, Aggarwal BB. Curcumin (diferuloylmethane) inhibits constitutive and IL-6-inducible STAT3 phosphorylation in human multiple myeloma cells. *J Immunol* 2003; **171**: 3863-3871 [PMID: 14500688]
- 68 **Weber-Nordt RM**, Egen C, Wehinger J, Ludwig W, Gouilleux-Gruart V, Mertelsmann R, Finke J. Constitutive activation of STAT proteins in primary lymphoid and myeloid leukemia cells and in Epstein-Barr virus (EBV)-related lymphoma cell lines. *Blood* 1996; **88**: 809-816 [PMID: 8704235]
- 69 **Mackenzie GG**, Queisser N, Wolfson ML, Fraga CG, Adamo AM, Oteiza PI. Curcumin induces cell-arrest and apoptosis in association with the inhibition of constitutively active NF-kappaB and STAT3 pathways in Hodgkin's lymphoma cells. *Int J Cancer* 2008; **123**: 56-65 [PMID: 18386790 DOI: 10.1002/ijc.23477]
- 70 **Seo JH**, Jeong KJ, Oh WJ, Sul HJ, Sohn JS, Kim YK, Cho do Y, Kang JK, Park CG, Lee HY. Lysophosphatidic acid induces STAT3 phosphorylation and ovarian cancer cell motility: their inhibition by curcumin. *Cancer Lett* 2010; **288**: 50-56 [PMID: 19647363 DOI: 10.1016/j.canlet.2009.06.023]
- 71 **Egwuagu CE**. STAT3 in CD4+ T helper cell differentiation and inflammatory diseases. *Cytokine* 2009; **47**: 149-156 [PMID: 19648026 DOI: 10.1016/j.cyto.2009.07.003]
- 72 **Podolsky DK**. Inflammatory bowel disease. *N Engl J Med* 2002; **347**: 417-429 [PMID: 12167685 DOI: 10.1056/NEJM199109263251306]
- 73 **Rutgeerts P**, Van Assche G, Vermeire S. Optimizing anti-TNF treatment in inflammatory bowel disease. *Gastroenterology* 2004; **126**: 1593-1610 [PMID: 15168370 DOI: 10.1053/j.gastro.2004.02.070]
- 74 **Barrett JC**, Hansoul S, Nicolae DL, Cho JH, Duerr RH, Rioux JD, Brant SR, Silverberg MS, Taylor KD, Barmada MM, Bitton A, Dassopoulos T, Datta LW, Green T, Griffiths AM, Kistner EO, Murtha MT, Regueiro MD, Rotter JL, Schumm LP, Steinhardt AH, Targan SR, Xavier RJ, Libioulle C, Sandor C, Lathrop M, Belaiche J, Dewit O, Gut I, Heath S, Laukens D, Mni M, Rutgeerts P, Van Gossum A, Zelenika D, Franchimont D, Hugot JP, de Vos M, Vermeire S, Louis E, Cardon LR, Anderson CA, Drummond H, Nimmo E, Ahmad T, Prescott NJ, Onnie CM, Fisher SA, Marchini J, Ghorji J, Bumpstead S, Gwilliam R, Tremelling M, Deloukas P, Mansfield J, Jewell D, Satsangi J, Mathew CG, Parkes M, Georges M, Daly MJ. Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat Genet* 2008; **40**: 955-962 [PMID: 18587394 DOI: 10.1038/ng.175]
- 75 **Medzhitov R**. Recognition of microorganisms and activation of the immune response. *Nature* 2007; **449**: 819-826 [PMID: 17943118 DOI: 10.1038/nature06246]
- 76 **Steinman RM**, Hemmi H. Dendritic cells: translating innate to adaptive immunity. *Curr Top Microbiol Immunol* 2006; **311**: 17-58 [PMID: 17048704 DOI: 10.1007/3-540-32636-7_2]
- 77 **Schindler C**, Levy DE, Decker T. JAK-STAT signaling: from interferons to cytokines. *J Biol Chem* 2007; **282**: 20059-20063 [PMID: 17502367 DOI: 10.1074/jbc.R700016200]
- 78 **Hunter CA**. New IL-12-family members: IL-23 and IL-27, cytokines with divergent functions. *Nat Rev Immunol* 2005; **5**: 521-531 [PMID: 15999093 DOI: 10.1038/nri1648]
- 79 **Lovato P**, Brender C, Agnholt J, Kelsen J, Kaltoft K, Svejgaard A, Eriksen KW, Woetmann A, Ødum N. Constitutive STAT3 activation in intestinal T cells from patients with Crohn's disease. *J Biol Chem* 2003; **278**: 16777-16781 [PMID: 12615922 DOI: 10.1074/jbc.M207999200]
- 80 **Takeda K**, Kaisho T, Yoshida N, Takeda J, Kishimoto T, Akira S. Stat3 activation is responsible for IL-6-dependent T cell proliferation through preventing apoptosis: generation and characterization of T cell-specific Stat3-deficient mice. *J Immunol* 1998; **161**: 4652-4660 [PMID: 9794394]
- 81 **Atreya I**, Atreya R, Neurath MF. NF-kappaB in inflammatory bowel disease. *J Intern Med* 2008; **263**: 591-596 [PMID: 18479258 DOI: 10.1111/j.1365-2796.2008.01953.x]
- 82 **Sugimoto K**. Role of STAT3 in inflammatory bowel disease. *World J Gastroenterol* 2008; **14**: 5110-5114 [PMID: 18777586 DOI: 10.3748/wjg.14.5110]
- 83 **Afif W**, Loftus EV. Safety profile of IBD therapeutics: infectious risks. *Med Clin North Am* 2010; **94**: 115-133 [PMID: 19944801 DOI: 10.1016/j.mcna.2009.08.016]
- 84 **Hilsden RJ**, Verhoef MJ, Rasmussen H, Porcino A, DeBruyn JC. Use of complementary and alternative medicine by patients with inflammatory bowel disease. *Inflamm Bowel Dis* 2011; **17**: 655-662 [PMID: 20848543 DOI: 10.1002/ibd.21360]
- 85 **Holt PR**, Katz S, Kirshoff R. Curcumin therapy in inflammatory bowel disease: a pilot study. *Dig Dis Sci* 2005; **50**: 2191-2193 [PMID: 16240238 DOI: 10.1007/s10620-005-3032-8]
- 86 **Hanai H**, Iida T, Takeuchi K, Watanabe F, Maruyama Y, Andoh A, Tsujikawa T, Fujiyama Y, Mitsuyama K, Sata M, Yamada M, Iwaoka Y, Kanke K, Hiraishi H, Hirayama K, Arai H, Yoshii S, Uchijima M, Nagata T, Koide Y. Curcumin maintenance therapy for ulcerative colitis: randomized, multicenter, double-blind, placebo-controlled trial. *Clin Gastroenterol Hepatol* 2006; **4**: 1502-1506 [PMID: 17101300 DOI: 10.1016/j.cgh.2006.08.008]
- 87 **Cruz-Correa M**, Shoskes DA, Sanchez P, Zhao R, Hyland LM, Wexner SD, Giardiello FM. Combination treatment with curcumin and quercetin of adenomas in familial adenomatous polyposis. *Clin Gastroenterol Hepatol* 2006; **4**: 1035-1038 [PMID: 16757216 DOI: 10.1016/j.cgh.2006.03.020]
- 88 **Angulo P**. Nonalcoholic fatty liver disease. *N Engl J Med* 2002; **346**: 1221-1231 [PMID: 11961152 DOI: 10.1056/NEJMr011775]

- 89 **Ratziu V**, Bellentani S, Cortez-Pinto H, Day C, Marchesini G. A position statement on NAFLD/NASH based on the EASL 2009 special conference. *J Hepatol* 2010; **53**: 372-384 [PMID: 20494470 DOI: 10.1016/j.jhep.2010.04.008]
- 90 **Rivera-Espinoza Y**, Muriel P. Pharmacological actions of curcumin in liver diseases or damage. *Liver Int* 2009; **29**: 1457-1466 [PMID: 19811613 DOI: 10.1111/j.1478-3231.2009.02086.x]
- 91 **Nanji AA**, Jokelainen K, Tipoe GL, Rahemtulla A, Thomas P, Dannenberg AJ. Curcumin prevents alcohol-induced liver disease in rats by inhibiting the expression of NF-kappa B-dependent genes. *Am J Physiol Gastrointest Liver Physiol* 2003; **284**: G321-G327 [PMID: 12388178 DOI: 10.1152/ajp-gi.00230.2002]
- 92 **Gressner AM**, Weiskirchen R. Modern pathogenetic concepts of liver fibrosis suggest stellate cells and TGF-beta as major players and therapeutic targets. *J Cell Mol Med* 2006; **10**: 76-99 [PMID: 16563223 DOI: 10.1111/j.1582-4934.2006.tb00292.x]
- 93 **Gupta SC**, Patchva S, Aggarwal BB. Therapeutic roles of curcumin: lessons learned from clinical trials. *AAPS J* 2013; **15**: 195-218 [PMID: 23143785 DOI: 10.1208/s12248-012-9432-8]
- 94 **Gaedeke J**, Noble NA, Border WA. Curcumin blocks fibrosis in anti-Thy 1 glomerulonephritis through up-regulation of heme oxygenase 1. *Kidney Int* 2005; **68**: 2042-2049 [PMID: 16221204 DOI: 10.1111/j.1523-1755.2004.00713.x]
- 95 **Kang HC**, Nan JX, Park PH, Kim JY, Lee SH, Woo SW, Zhao YZ, Park EJ, Sohn DH. Curcumin inhibits collagen synthesis and hepatic stellate cell activation in-vivo and in-vitro. *J Pharm Pharmacol* 2002; **54**: 119-126 [PMID: 11829122 DOI: 10.1211/0022357021771823]
- 96 **Miquel J**, Bernd A, Sempere JM, Díaz-Alperi J, Ramírez A. The curcuma antioxidants: pharmacological effects and prospects for future clinical use. A review. *Arch Gerontol Geriatr* 2002; **34**: 37-46 [PMID: 14764309 DOI: 10.1016/S0167-4943(01)00194-7]
- 97 **Iqbal M**, Sharma SD, Okazaki Y, Fujisawa M, Okada S. Dietary supplementation of curcumin enhances antioxidant and phase II metabolizing enzymes in ddY male mice: possible role in protection against chemical carcinogenesis and toxicity. *Pharmacol Toxicol* 2003; **92**: 33-38 [PMID: 12710595 DOI: 10.1034/j.1600-0773.2003.920106.x]
- 98 **Ak T**, Gülçin I. Antioxidant and radical scavenging properties of curcumin. *Chem Biol Interact* 2008; **174**: 27-37 [PMID: 18547552 DOI: 10.1016/j.cbi.2008.05.003]
- 99 **Motterlini R**, Foresti R, Bassi R, Green CJ. Curcumin, an antioxidant and anti-inflammatory agent, induces heme oxygenase-1 and protects endothelial cells against oxidative stress. *Free Radic Biol Med* 2000; **28**: 1303-1312 [PMID: 10889462 DOI: 10.1016/S0891-5849(00)00294-X]
- 100 **Savarino E**, Zentilin P, Savarino V. NERD: an umbrella term including heterogeneous subpopulations. *Nat Rev Gastroenterol Hepatol* 2013; **10**: 371-380 [PMID: 23528345 DOI: 10.1038/nrgastro.2013.50]
- 101 **Knowles CH**, Aziz Q. Visceral hypersensitivity in non-erosive reflux disease. *Gut* 2008; **57**: 674-683 [PMID: 18079285 DOI: 10.1136/gut.2007.127886]
- 102 **Caterina MJ**, Rosen TA, Tominaga M, Brake AJ, Julius D. A capsaicin-receptor homologue with a high threshold for noxious heat. *Nature* 1999; **398**: 436-441 [PMID: 10201375 DOI: 10.1038/18906]
- 103 **Zhi L**, Dong L, Kong D, Sun B, Sun Q, Grundy D, Zhang G, Rong W. Curcumin acts via transient receptor potential vanilloid-1 receptors to inhibit gut nociception and reverses visceral hyperalgesia. *Neurogastroenterol Motil* 2013; **25**: e429-e440 [PMID: 23638900 DOI: 10.1111/nmo.12145]
- 104 **Guarino MP**, Cheng L, Ma J, Harnett K, Biancani P, Altomare A, Panzera F, Behar J, Cicala M. Increased TRPV1 gene expression in esophageal mucosa of patients with non-erosive and erosive reflux disease. *Neurogastroenterol Motil* 2010; **22**: 746-751, e219 [PMID: 20456759 DOI: 10.1111/j.1365-2982.2010.01514.x]
- 105 **Savarino E**, Zentilin P, Tutuian R, Pohl D, Casa DD, Frazzoni M, Cestari R, Savarino V. The role of nonacid reflux in NERD: lessons learned from impedance-pH monitoring in 150 patients off therapy. *Am J Gastroenterol* 2008; **103**: 2685-2693 [PMID: 18775017 DOI: 10.1111/j.1572-0241.2008.02119.x]
- 106 **Zagari RM**, Fuccio L, Wallander MA, Johansson S, Fiocca R, Casanova S, Farahmand BY, Winchester CC, Roda E, Bazzoli F. Gastro-oesophageal reflux symptoms, oesophagitis and Barrett's oesophagus in the general population: the Loiano-Monghidoro study. *Gut* 2008; **57**: 1354-1359 [PMID: 18424568 DOI: 10.1136/gut.2007.145177]
- 107 **Barbara G**, Stanghellini V, De Giorgio R, Cremon C, Cottrell GS, Santini D, Pasquinelli G, Morselli-Labate AM, Grady EF, Bunnett NW, Collins SM, Corinaldesi R. Activated mast cells in proximity to colonic nerves correlate with abdominal pain in irritable bowel syndrome. *Gastroenterology* 2004; **126**: 693-702 [PMID: 14988823 DOI: 10.1053/j.gastro.2003.11.055]

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