

Intestinal barrier: Molecular pathways and modifiers

Min Kyung Jeon, Christina Klaus, Elke Kaemmerer, Nikolaus Gassler

Min Kyung Jeon, Christina Klaus, Elke Kaemmerer, Nikolaus Gassler, Institute of Pathology, RWTH Aachen University, 52074 Aachen, Germany

Author contributions: Jeon MK and Gassler N designed and wrote the article; Klaus C and Kaemmerer E critical reviewed the literature and made technical notes.

Correspondence to: Nikolaus Gassler, MA, Professor, Institute of Pathology, RWTH Aachen University, Pauwelsstraße 30, 52074 Aachen, Germany. ngassler@ukaachen.de

Telephone: +49-241-8088897 Fax: +49-241-8082439

Received: June 24, 2013 Revised: August 28, 2013

Accepted: September 3, 2013

Published online: November 15, 2013

Abstract

The gastrointestinal tract is frequently challenged by pathogens/antigens contained in food and water and the intestinal epithelium must be capable of rapid regeneration in the event of tissue damage. Disruption of the intestinal barrier leads to a number of immune-mediated diseases, including inflammatory bowel disease, food allergy, and celiac disease. The intestinal mucosa is composed of different types of epithelial cells in specific barrier functions. Epithelial cells control surface-associated bacterial populations without disrupting the intestinal microflora that is crucial for host health. They are also capable of modulating mucosal immune system, and are thus essential in maintaining homeostasis in the gut. Thus, the regulation of intestinal epithelial homeostasis is crucial for the maintenance of the structure of the mucosa and the defensive barrier functions. Recent studies have demonstrated that multiple molecular pathways are involved in the regulation of intestinal epithelial cell polarity. These include the Wnt, Notch, Hippo, transforming growth factor- β (TGF- β)/bone morphogenetic protein (BMP) and Hedgehog pathways, most of which were identified in lower organisms where they play important roles during embryogenesis. These pathways are also used in adult organisms to regulate multiple self-renewing organs. Understanding the interactions between these

molecular mechanisms and intestinal barrier function will therefore provide important insight into the pathogenesis of intestinal-based immune-mediated diseases.

© 2013 Baishideng Publishing Group Co., Limited. All rights reserved.

Key words: Intestinal epithelium; Mucosal barrier; Homeostasis; Molecular pathways; Immune-mediated disease

Core tip: The pathogenesis of gastrointestinal diseases is associated with important molecular pathways such as Wnt, Notch, Hippo, transforming growth factor- β /bone morphogenetic protein or Hedgehog in controlling cell-fate determination. Here, we discuss how they contribute to homeostasis of intestinal epithelium.

Jeon MK, Klaus C, Kaemmerer E, Gassler N. Intestinal barrier: Molecular pathways and modifiers. *World J Gastrointest Pathophysiol* 2013; 4(4): 94-99 Available from: URL: <http://www.wjgnet.com/2150-5330/full/v4/i4/94.htm> DOI: <http://dx.doi.org/10.4291/wjgp.v4.i4.94>

INTRODUCTION

The intestinal epithelium is a single-cell layer that serves as a protective barrier against the external environment. It supports nutrients and water transport, while maintaining a defense against intraluminal toxins, bacteria, and antigens. The epithelial barrier is governed by the expression of adherens junctions (AJs) and tight junctions (TJs), including cadherins, claudins, occludin, and junctional adhesion molecules (JAM) proteins, which seal adjacent cells together^[1]. Expression of AJ and TJ proteins is regulated by phosphorylation and it can either promote or destabilize TJ formation^[2]. The AJ and TJ complexes also play a crucial role in the regulation of cellular polarization, proliferation, and differentiation^[3].

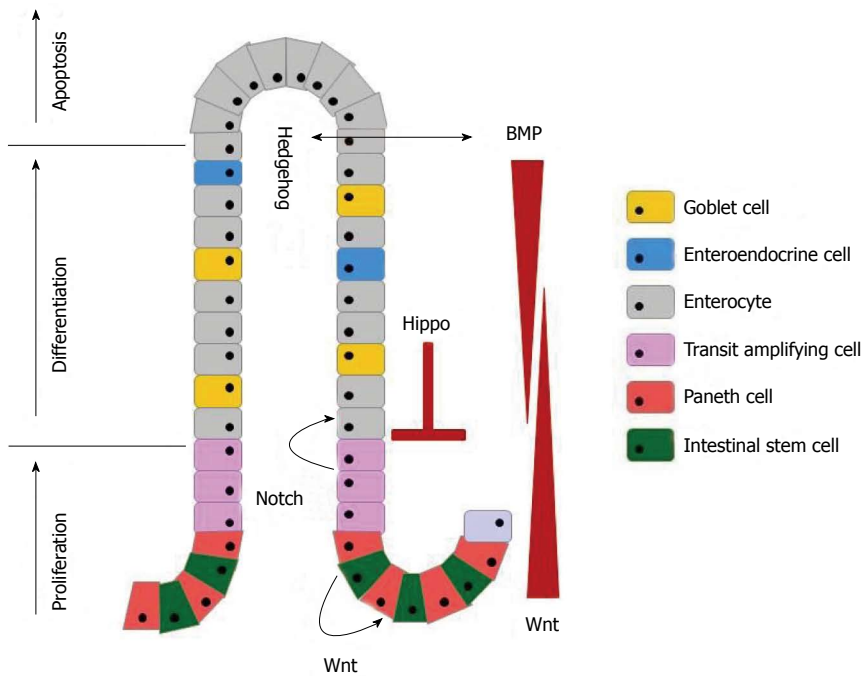


Figure 1 The molecular regulatory pathways in intestinal epithelial homeostasis. Wnt signaling promotes the proliferation of TA/stem cells and drives Paneth cell maturation. Notch signaling cooperates with Wnt to drive proliferation of intestinal stem cells and regulates maintenance of undifferentiated TA/stem cells. Hippo and bone morphogenetic protein (BMP) signaling inhibits proliferation and induces maturation of all secretory cell types. Hedgehog signaling activates the expression of BMP in the mesenchyme.

The intestinal epithelium is populated by distinct types of cells derived from stem cells, such as the absorptive cells (enterocytes) and the secretory cells (mucus-secreting goblet cells, hormone-secreting enteroendocrine cells, Tuft cells, and antimicrobial peptides-secreting Paneth cells). All but the Paneth cells differentiate into mature forms from the crypts to replace cells extruded from the tips of the villi^[4,5]. This continuous replenishment of intestinal epithelium takes generally 4-7 d and it is important for the maintenance of epithelial integrity. Several conserved signaling cascades, *i.e.*, the Wnt, transforming growth factor- β (TGF- β)/bone morphogenetic protein (BMP), Notch, Hippo, and Hedgehog pathway are associated with the maintenance of the morphological and functional features of diverse epithelial cell types (Figure 1).

Disruption of this delicate balance causes a variety of intestinal-related inflammatory and autoimmune syndromes^[6-8]. For example, healthy first-degree relatives of patients with inflammatory bowel disease (IBD) and celiac disease have increased intestinal permeability and epithelial apoptosis^[9,10]. Intestinal barrier dysfunction also contributes to the severity of food allergen-induced clinical symptoms^[11]. In this review, we summarize current studies for major molecular signaling pathways in intestinal homeostasis.

SIGNALING PATHWAYS REGULATING INTESTINAL HOMEOSTASIS

WNT PATHWAY

Wnt signaling plays multiple roles during intestinal ho-

meostasis. It contains the canonical and non-canonical pathway. The central player in the canonical Wnt signaling cascade is β -catenin, a cytoplasmic protein the stability of which is regulated by the APC tumor suppressor. Complexes of frizzled seven transmembrane molecules and low-density lipoprotein receptor related proteins (LRP-5/-6) serve as receptors for secreted Wnts^[12]. When Wnt receptor complexes are not engaged, casein kinase-1 and GSK3- β , both residing in the APC complex, sequentially phosphorylate β -catenin at a series of highly conserved Ser/Thr residues near its N-terminus and are thereby tagged for ubiquitination and proteasomal degradation. In contrast, Wnt stimulation blocks the intrinsic kinase activity of the APC complex, leading to β -catenin accumulation. Unphosphorylated β -catenin translocates into the nucleus where it engages transcription factors of the TCF/LEF family, thereby translating the Wnt signal into the transcription of a TCF target gene^[13].

The canonical Wnt pathway is essential for epithelial proliferation and crypt maintenance^[14,15]. Mutations in the *APC* gene, a negative regulator of Wnt signaling, also results in hyperproliferation of the epithelium followed by the development of adenomas^[16]. Moreover, β -catenin plays a role as an AJ component, thereby linking it to the cytoskeleton of epithelial cells. Wnt signaling is necessary for positioning and maturation of Paneth cells in the crypts, and for separating proliferating and differentiated cells. These processes are controlled by the Wnt-dependent expression of specific ephrin receptors in the intestine^[17]. Non-canonical (β -catenin

independent) Wnt signaling is termed the planar cell polarity pathway, which is activated by the GTPases Rho and Rac. These induce cytoskeletal rearrangements and help to form new crypts^[18].

NOTCH PATHWAY

Notch signaling is active in intestinal crypt compartments and assists the Wnt pathway in promoting stem cell proliferation and negatively regulates differentiation into the secretory lineages^[19]. Interaction of the Notch receptor (NOTCH 1-4) and ligand (Delta-like 1, 3, and 4; Jagged 1 and 2) between two adjacent cells results in proteolytic cleavages of the receptor by the γ -secretase enzyme complex, leading to the translocation of the Notch intracellular domain (NICD) into the nucleus. There, NICD binds to the transcription factor CSL (CBF1 in human, RBPjk in mice, Su (H) in *Drosophila*, Lag-1 in *C. elegans*) and activates target genes such as *HES-1* (Hairy and enhancer of split 1), *HES-3*, and *HES-5*, which are important for the differentiation of absorptive cells. The transcription factor MATH-1 is a downstream target of HES-1 repression in the intestine and its activity leads to generation of the secretory cell lineages^[20,21]. These results suggest that the absorptive versus secretory epithelial cell type fate decision is established through the HES/MATH1 axis. In addition, Gfi-1 and neurogenin-3 (Ngn-3), other transcription factors, compete for selection of enteroendocrine versus goblet or Paneth cell fates^[22].

The dysregulation of Notch activity is related to the pathogenesis of IBDs, such as ulcerative colitis (UC) and Crohn's disease (CD). A histological study in UC revealed that the depletion of goblet cells with loss of ATOH1 expression and CD is caused by dysregulation of secretory cell differentiation^[23-25]. Additional evidence supporting such an important role for Notch in the intestine is derived from studies of γ -secretase inhibitors (GSIs). Treatment of mice with GSIs induced colitis due to inhibition of Notch signaling^[26]. Notch activity may thus contribute to the regenerating epithelium and enhance the barrier function of the intestinal epithelium.

HIPPO PATHWAY

The Hippo pathway plays a crucial role in controlling organ size by inhibiting cell proliferation and apoptosis in response to cell-cell contact. This tumor suppressor pathway regulates intestinal regeneration and tumorigenesis^[27,28]. When the Hippo pathway is active, the downstream effector of this pathway Yes-associated protein (YAP) is phosphorylated at S127 by the LAT1/2 kinases. Phosphorylated YAP remains in the cytoplasm and inhibits its proliferative and anti-apoptotic function in the nucleus, which is mediated by its binding to TEAD1-4 transcription factors. Cytoplasmic YAP has the Wnt antagonizing effects, thereby contributing to the prevention of proliferation and intestinal stem cell expansion^[29].

In contrast, the deletion of Hippo pathway component Mst1/2 in mouse intestinal epithelial cells results in an expansion of undifferentiated stem cells and an absence of all secretory lineages^[30].

In the small intestinal and the colonic epithelium of the normal mouse, YAP protein is found in the crypts and under normal conditions YAP makes no contribution to intestinal epithelial proliferation. However, it is required for tissue regeneration caused by injury. YAP protein is overexpressed in the crypts of the recovery phase from the DSS-treated mice^[31]. Treatment of GSIs suppressed the intestinal dysplasia caused by YAP and this result suggests that YAP stimulates Notch signaling. YAP is commonly overexpressed in colorectal cancers^[32]. Therefore, the activation of YAP for regulating intestinal stem cells regeneration indicates that a deficiency of Hippo signaling may contribute to tumorigenesis in the intestine.

HEDGEHOG PATHWAY

The Hedgehog signaling is initiated through the binding of Hedgehog ligands to the patched homolog 1 (Ptch1) receptor. In the absence of ligands, Ptch1 inhibits the activity of smoothed (Smo). Hedgehog binding inactivates Ptch1 and in contrast, Smo inhibition is released and these mechanisms lead to the translocation of members of the GLI family (GLI-1, GLI-2, and GLI-3 in mammals) of Zn-finger transcription factors from the cytoplasm to the nucleus. Upon nuclear translocation, they activate the transcription of target genes, such as Ptch1, Gli-1, Bmp4, Hhip1^[33]. Three Hedgehog homologs, such as sonic hedgehog (Shh), indian hedgehog (Ihh), and desert hedgehog (Dhh) are known to be highly conserved in mammals and of these, only Shh and Ihh are expressed in epithelium during development, but are redistributed after villus formation to be localized to the intervillus region^[34]. Thus, Hedgehog signaling is required for the formation of villi. Shh has been localized to the region of the crypts and Shh plays a role in the regulation of epithelial proliferation^[35]. Furthermore, Hedgehog ligands have been reported to be anti-inflammatory epithelial modulators in the intestine^[36]. Hedgehog signaling pathway inhibitors induced hypoplasia of Paneth cells, thus this signaling pathway may influence intestinal epithelium repair partly through the regulation of Paneth cells^[37]. Hedgehog signaling decreases during the injury phase and increases during the repair phase^[38]. It confirms an important role of Hedgehog signaling in the repair of intestinal epithelium after injury.

TGF- β /BMP PATHWAY

TGF- β signaling regulates embryonic development, wound healing, proliferation, and cell differentiation^[39,40]. The TGF- β family consists of cytokines including TGF- β isoform, BMPs, and activins. Signaling is induced by ligand binding to type II serine/threonine kinase receptors, which results in the phosphorylation of

the type I receptor. Then, signaling of these activated receptors is transduced through three classes of SMAD proteins: receptor-regulated SMADs (R-SMADs: SMAD-1, -2, -3, -5, and -8), common SMAD (SMAD-4), and inhibitory SMADs (I-SMADs: SMAD-6 and -7). R-SMADs become phosphorylated by activated type I receptors and subsequent translocate to the nucleus. The SMAD complex interacts with transcription factors, thereby inducing target gene expression. TGF- β signaling components are expressed in the differentiated compartment of the intestine^[41]. Inactivation of TGF- β signaling components has also been identified at the adenoma-to-carcinoma transition^[42].

BMP signaling mediates the action of hedgehog, blocking the formation of ectopic crypts, and the expression of BMP antagonist noggin in the crypts prevents the activity of BMP, thereby enabling proliferation to continue^[43]. TGF- β ligands signal through SMAD-2 and -3, whereas BMP signaling is mediated through SMAD-1, -5 and -8. Phosphorylated SMAD-1, -5, and -8 are observed in the villus epithelium and this modification prevents the villus epithelium from adopting a crypt-like proliferative character^[44]. BMP signaling antagonizes the Wnt pathway within the differentiated compartment, thereby positioning transient amplifying cells in the crypts^[45]. In addition, humans with germline mutations in SMAD-4 or BMP receptor type 1A (BMPRI1A) are associated with up to 50% of JPS (Juvenile polyposis syndrome) cases^[46,47].

CONCLUSION

Intestinal epithelial barrier dysfunction has been implicated as a critical role in the predisposition to a number of gastrointestinal diseases such as IBD, food allergy, and celiac disease. It could lead to defects in multiple aspects of microbial, epithelial, and immune interactions, and thus injury to the epithelial lining thought to be an important factor in the pathogenesis of intestinal-based immune-mediated diseases^[48-50].

The homeostasis of the intestinal epithelium is maintained by a complex interplay of multiple regulatory mechanisms. Together, the Wnt, Hedgehog and TGF- β /BMP pathways maintain the crypt-villus architecture. The Wnt, Notch, and Hippo signaling pathways combine to control the cell fate choices from stem cells. Thus and understanding of critical signaling pathways for maintaining intestinal homeostasis is essential.

ACKNOWLEDGMENTS

We would like to thank P Akens for her help in typing and proofreading the manuscript.

REFERENCES

- 1 **Laukoetter MG**, Bruewer M, Nusrat A. Regulation of the intestinal epithelial barrier by the apical junctional complex.

- Curr Opin Gastroenterol* 2006; **22**: 85-89 [PMID: 16462161 DOI: 10.1097/01.mog.0000203864.48255.4f]
- 2 **Atkinson KJ**, Rao RK. Role of protein tyrosine phosphorylation in acetaldehyde-induced disruption of epithelial tight junctions. *Am J Physiol Gastrointest Liver Physiol* 2001; **280**: G1280-G1288 [PMID: 11352822]
- 3 **Hartsock A**, Nelson WJ. Adherens and tight junctions: structure, function and connections to the actin cytoskeleton. *Biochim Biophys Acta* 2008; **1778**: 660-669 [PMID: 17854762 DOI: 10.1016/j.bbame.2007.07.012]
- 4 **Porter EM**, Bevins CL, Ghosh D, Ganz T. The multifaceted Paneth cell. *Cell Mol Life Sci* 2002; **59**: 156-170 [PMID: 11846026]
- 5 **Sancho E**, Batlle E, Clevers H. Signaling pathways in intestinal development and cancer. *Annu Rev Cell Dev Biol* 2004; **20**: 695-723 [PMID: 15473857 DOI: 10.1146/annurev.cellbio.20.010403.092805]
- 6 **Brandt EB**, Strait RT, Hershko D, Wang Q, Muntel EE, Scribner TA, Zimmermann N, Finkelman FD, Rothenberg ME. Mast cells are required for experimental oral allergen-induced diarrhea. *J Clin Invest* 2003; **112**: 1666-1677 [PMID: 14660743 DOI: 10.1172/JCI19785]
- 7 **Schulzke JD**, Bentzel CJ, Schulzke I, Riecken EO, Fromm M. Epithelial tight junction structure in the jejunum of children with acute and treated celiac sprue. *Pediatr Res* 1998; **43**: 435-441 [PMID: 9544995 DOI: 10.1203/00006450-199804000-00001]
- 8 **D'Incà R**, Di Leo V, Corrao G, Martines D, D'Odorico A, Mestriner C, Venturi C, Longo G, Sturniolo GC. Intestinal permeability test as a predictor of clinical course in Crohn's disease. *Am J Gastroenterol* 1999; **94**: 2956-2960 [PMID: 10520851 DOI: 10.1111/j.1572-0241.1999.01444.x]
- 9 **Peeters M**, Geypens B, Claus D, Nevens H, Ghooys Y, Verbeke G, Baert F, Vermeire S, Vlietinck R, Rutgeerts P. Clustering of increased small intestinal permeability in families with Crohn's disease. *Gastroenterology* 1997; **113**: 802-807 [PMID: 9287971]
- 10 **Gitter AH**, Bendfeldt K, Schulzke JD, Fromm M. Leaks in the epithelial barrier caused by spontaneous and TNF- α -induced single-cell apoptosis. *FASEB J* 2000; **14**: 1749-1753 [PMID: 10973924 DOI: 10.1096/fj.99-0898com]
- 11 **Ventura MT**, Polimeno L, Amoroso AC, Gatti F, Annoscia E, Marinaro M, Di Leo E, Matino MG, Buquicchio R, Bonini S, Turisi A, Francavilla A. Intestinal permeability in patients with adverse reactions to food. *Dig Liver Dis* 2006; **38**: 732-736 [PMID: 16880015 DOI: 10.1016/j.dld.2006.06.012]
- 12 **Giles RH**, van Es JH, Clevers H. Caught up in a Wnt storm: Wnt signaling in cancer. *Biochim Biophys Acta* 2003; **1653**: 1-24 [PMID: 12781368]
- 13 **Aberle H**, Bauer A, Stappert J, Kispert A, Kemler R. *beta*-catenin is a target for the ubiquitin-proteasome pathway. *EMBO J* 1997; **16**: 3797-3804 [PMID: 9233789 DOI: 10.1093/emboj/16.13.3797]
- 14 **Korinek V**, Barker N, Moerer P, van Donselaar E, Huls G, Peters PJ, Clevers H. Depletion of epithelial stem-cell compartments in the small intestine of mice lacking Tcf-4. *Nat Genet* 1998; **19**: 379-383 [PMID: 9697701 DOI: 10.1038/1270]
- 15 **Pinto D**, Gregorieff A, Begthel H, Clevers H. Canonical Wnt signals are essential for homeostasis of the intestinal epithelium. *Genes Dev* 2003; **17**: 1709-1713 [PMID: 12865297 DOI: doi:]
- 16 **Oshima M**, Oshima H, Kitagawa K, Kobayashi M, Itakura C, Taketo M. Loss of Apc heterozygosity and abnormal tissue building in nascent intestinal polyps in mice carrying a truncated Apc gene. *Proc Natl Acad Sci USA* 1995; **92**: 4482-4486 [PMID: 7753829]
- 17 **Stappenbeck TS**, Mills JC, Gordon JI. Molecular features of adult mouse small intestinal epithelial progenitors. *Proc Natl Acad Sci USA* 2003; **100**: 1004-1009 [PMID: 12552106 DOI: 10.1073/pnas.242735899]

- 18 **Nusse R.** Wnt signaling. *Cold Spring Harb Perspect Biol* 2012; **4**: 1749-1753 [PMID: 22550232 DOI: 10.1101/cshperspect.a011163]
- 19 **VanDussen KL,** Carulli AJ, Keeley TM, Patel SR, Puthoff BJ, Magness ST, Tran IT, Maillard I, Siebel C, Kolterud Å, Grosse AS, Gumucio DL, Ernst SA, Tsai YH, Dempsey PJ, Samuelson LC. Notch signaling modulates proliferation and differentiation of intestinal crypt base columnar stem cells. *Development* 2012; **139**: 488-497 [PMID: 22190634 DOI: 10.1242/dev.070763]
- 20 **De Strooper B,** Annaert W, Cupers P, Saftig P, Craessaerts K, Mumm JS, Schroeter EH, Schrijvers V, Wolfe MS, Ray WJ, Goate A, Kopan R. A presenilin-1-dependent gamma-secretase-like protease mediates release of Notch intracellular domain. *Nature* 1999; **398**: 518-522 [PMID: 10206645 DOI: 10.1038/19083]
- 21 **Ueo T,** Imayoshi I, Kobayashi T, Ohtsuka T, Seno H, Nakase H, Chiba T, Kageyama R. The role of Hes genes in intestinal development, homeostasis and tumor formation. *Development* 2012; **139**: 1071-1082 [PMID: 22318232 DOI: 10.1242/dev.069070]
- 22 **Shroyer NF,** Wallis D, Venken KJ, Bellen HJ, Zoghbi HY. Gfi1 functions downstream of Math1 to control intestinal secretory cell subtype allocation and differentiation. *Genes Dev* 2005; **19**: 2412-2417 [PMID: 16230531 DOI: 10.1101/gad.1353905]
- 23 **Zheng X,** Tsuchiya K, Okamoto R, Iwasaki M, Kano Y, Sakamoto N, Nakamura T, Watanabe M. Suppression of hath1 gene expression directly regulated by hes1 via notch signaling is associated with goblet cell depletion in ulcerative colitis. *Inflamm Bowel Dis* 2011; **17**: 2251-2260 [PMID: 21987298 DOI: 10.1002/ibd.21611]
- 24 **Gersemann M,** Becker S, Kübler I, Koslowski M, Wang G, Herrlinger KR, Griger J, Fritz P, Fellermann K, Schwab M, Wehkamp J, Stange EF. Differences in goblet cell differentiation between Crohn's disease and ulcerative colitis. *Differentiation* 2009; **77**: 84-94 [PMID: 19281767 DOI: 10.1016/j.diff.2008.09.008]
- 25 **Wehkamp J,** Salzman NH, Porter E, Nuding S, Weichen-thal M, Petras RE, Shen B, Schaeffeler E, Schwab M, Linz-meier R, Feathers RW, Chu H, Lima H, Fellermann K, Ganz T, Stange EF, Bevins CL. Reduced Paneth cell alpha-defensins in ileal Crohn's disease. *Proc Natl Acad Sci USA* 2005; **102**: 18129-18134 [PMID: 16330776 DOI: 10.1073/pnas.0505256102]
- 26 **Okamoto R,** Tsuchiya K, Nemoto Y, Akiyama J, Nakamura T, Kanai T, Watanabe M. Requirement of Notch activation during regeneration of the intestinal epithelia. *Am J Physiol Gastrointest Liver Physiol* 2009; **296**: G23-G35 [PMID: 19023031 DOI: 10.1152/ajpgi.90225.2008]
- 27 **Karpowicz P,** Perez J, Perrimon N. The Hippo tumor suppressor pathway regulates intestinal stem cell regeneration. *Development* 2010; **137**: 4135-4145 [PMID: 21098564 DOI: 10.1242/dev.060483]
- 28 **Cai J,** Zhang N, Zheng Y, de Wilde RF, Maitra A, Pan D. The Hippo signaling pathway restricts the oncogenic potential of an intestinal regeneration program. *Genes Dev* 2010; **24**: 2383-2388 [PMID: 21041407 DOI: 10.1101/gad.1978810]
- 29 **Chen L,** Qin F, Deng X, Avruch J, Zhou D. Hippo pathway in intestinal homeostasis and tumorigenesis. *Protein Cell* 2012; **3**: 305-310 [PMID: 22492181 DOI: 10.1007/s13238-012-2913-9]
- 30 **Zhou D,** Zhang Y, Wu H, Barry E, Yin Y, Lawrence E, Dawson D, Willis JE, Markowitz SD, Camargo FD, Avruch J. Mst1 and Mst2 protein kinases restrain intestinal stem cell proliferation and colonic tumorigenesis by inhibition of Yes-associated protein (Yap) overabundance. *Proc Natl Acad Sci USA* 2011; **108**: E1312-E1320 [PMID: 22042863 DOI: 10.1073/pnas.1110428108]
- 31 **Ren F,** Wang B, Yue T, Yun EY, Ip YT, Jiang J. Hippo signaling regulates Drosophila intestine stem cell proliferation through multiple pathways. *Proc Natl Acad Sci USA* 2010; **107**: 21064-21069 [PMID: 21078993 DOI: 10.1073/pnas.1012759107]
- 32 **Steinhardt AA,** Gayyed MF, Klein AP, Dong J, Maitra A, Pan D, Montgomery EA, Anders RA. Expression of Yes-associated protein in common solid tumors. *Hum Pathol* 2008; **39**: 1582-1589 [PMID: 18703216 DOI: 10.1016/j.humpath.2008.04.012]
- 33 **Ingham PW,** McMahon AP. Hedgehog signaling in animal development: paradigms and principles. *Genes Dev* 2001; **15**: 3059-3087 [PMID: 11731473 DOI: 10.1101/gad.938601]
- 34 **Watt FM.** Unexpected Hedgehog-Wnt interactions in epithelial differentiation. *Trends Mol Med* 2004; **10**: 577-580 [PMID: 15567325 DOI: 10.1016/j.molmed.2004.10.008]
- 35 **Barker N,** van Es JH, Kuipers J, Kujala P, van den Born M, Cozijnsen M, Haegbarth A, Korving J, Begthel H, Peters PJ, Clevers H. Identification of stem cells in small intestine and colon by marker gene Lgr5. *Nature* 2007; **449**: 1003-1007 [PMID: 17934449 DOI: 10.1038/nature06196]
- 36 **Zacharias WJ,** Li X, Madison BB, Kretovich K, Kao JY, Merchant JL, Gumucio DL. Hedgehog is an anti-inflammatory epithelial signal for the intestinal lamina propria. *Gastroenterology* 2010; **138**: 2368-2377, 2377. e1-e4 [PMID: 20206176 DOI: 10.1053/j.gastro.2010.02.057]
- 37 **van den Brink GR.** Hedgehog signaling in development and homeostasis of the gastrointestinal tract. *Physiol Rev* 2007; **87**: 1343-1375 [PMID: 17928586 DOI: 10.1152/physrev.00054.2006]
- 38 **Liang R,** Morris P, Cho SS, Abud HE, Jin X, Cheng W. Hedgehog signaling displays a biphasic expression pattern during intestinal injury and repair. *J Pediatr Surg* 2012; **47**: 2251-2263 [PMID: 23217885 DOI: 10.1016/j.jpedsurg.2012.09.016]
- 39 **Blobe GC,** Schiemann WP, Lodish HF. Role of transforming growth factor beta in human disease. *N Engl J Med* 2000; **342**: 1350-1358 [PMID: 10793168 DOI: 10.1056/NEJM200005043421807]
- 40 **Shi Y,** Massagué J. Mechanisms of TGF- β signaling from cell membrane to the nucleus. *Cell* 2003; **113**: 685-700 [PMID: 12809600]
- 41 **Winesett MP,** Ramsey GW, Barnard JA. Type II TGF(beta) receptor expression in intestinal cell lines and in the intestinal tract. *Carcinogenesis* 1996; **17**: 989-995 [PMID: 8640948]
- 42 **Fearon ER,** Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell* 1990; **61**: 759-767 [PMID: 2188735]
- 43 **Haramis AP,** Begthel H, van den Born M, van Es J, Jonkheer S, Offerhaus GJ, Clevers H. De novo crypt formation and juvenile polyposis on BMP inhibition in mouse intestine. *Science* 2004; **303**: 1684-1686 [PMID: 15017003 DOI: 10.1126/science.1093587]
- 44 **He XC,** Zhang J, Tong WG, Tawfik O, Ross J, Scoville DH, Tian Q, Zeng X, He X, Wiedemann LM, Mishina Y, Li L. BMP signaling inhibits intestinal stem cell self-renewal through suppression of Wnt-beta-catenin signaling. *Nat Genet* 2004; **36**: 1117-1121 [PMID: 15378062 DOI: 10.1038/ng1430]
- 45 **Radtke F,** Clevers H, Riccio O. From gut homeostasis to cancer. *Curr Mol Med* 2006; **6**: 275-289 [PMID: 16712475]
- 46 **Zhou XP,** Woodford-Richens K, Lehtonen R, Kurose K, Aldred M, Hampel H, Launonen V, Virta S, Pilarski R, Salovaara R, Bodmer WF, Conrad BA, Dunlop M, Hodgson SV, Iwama T, Järvinen H, Kellokumpu I, Kim JC, Leggett B, Markie D, Mecklin JP, Neale K, Phillips R, Pirus J, Rozen P, Houlston RS, Aaltonen LA, Tomlinson IP, Eng C. Germline mutations in BMPRI1/ALK3 cause a subset of cases of juvenile polyposis syndrome and of Cowden and Bannayan-Riley-Ruvalcaba syndromes. *Am J Hum Genet* 2001; **69**: 704-711 [PMID: 11536076 DOI: 10.1086/323703]
- 47 **Howe JR,** Bair JL, Sayed MG, Anderson ME, Mitros FA, Petersen GM, Velculescu VE, Traverso G, Vogelstein B. Germline mutations of the gene encoding bone morphogenetic protein receptor 1A in juvenile polyposis. *Nat Genet* 2001;

- 28: 184-187 [PMID: 11381269 DOI: 10.1038/88919]
- 48 **Wu Y**, Cain-Hom C, Choy L, Hagenbeek TJ, de Leon GP, Chen Y, Finkle D, Venook R, Wu X, Ridgway J, Schahin-Reed D, Dow GJ, Shelton A, Stawicki S, Watts RJ, Zhang J, Choy R, Howard P, Kadyk L, Yan M, Zha J, Callahan CA, Hymowitz SG, Siebel CW. Therapeutic antibody targeting of individual Notch receptors. *Nature* 2010; **464**: 1052-1057 [PMID: 20393564 DOI: 10.1038/nature08878]
- 49 **Terzić J**, Grivennikov S, Karin E, Karin M. Inflammation and colon cancer. *Gastroenterology* 2010; **138**: 2101-2114.e5 [PMID: 20420949 DOI: 10.1053/j.gastro.2010.01.058]
- 50 **Koch S**, Nava P, Addis C, Kim W, Denning TL, Li L, Parkos CA, Nusrat A. The Wnt antagonist Dkk1 regulates intestinal epithelial homeostasis and wound repair. *Gastroenterology* 2011; **141**: 259-268, 268.e1-8 [PMID: 21440550 DOI: 10.1053/j.gastro.2011.03.043]

P- Reviewers: Jeung EB, Morini S

S- Editor: Song XX **L- Editor:** A **E- Editor:** Liu XM





百世登

Baishideng®

Published by **Baishideng Publishing Group Co., Limited**

Flat C, 23/F., Lucky Plaza,

315-321 Lockhart Road, Wan Chai, Hong Kong, China

Telephone: +852-6555-7188

Fax: +852-3177-9906

E-mail: bpgoffice@wjgnet.com

<http://www.wjgnet.com>

