

WJH 6th Anniversary Special Issues (1): Management of hepatocellular carcinoma**Management of hepatocellular carcinoma: Predictive value of immunohistochemical markers for postoperative survival**

Zhao-Shan Niu, Xiao-Jun Niu, Mei Wang

Zhao-Shan Niu, Lab of Micromorphology, Medical College of Qingdao University, Qingdao 266071, Shandong Province, China
Xiao-Jun Niu, Clinical Medicine Specialty, Medical College of Qingdao University, Qingdao 266071, Shandong Province, China
Mei Wang, Institute of Education, the University of Reading, RG6 1HY Reading, United Kingdom

Author contributions: Niu ZS designed the review's objectives; Niu XJ and Wang M searched the literature for latest developments in the field; Niu ZS and Wang M were involved in reviewing the literature, writing and editing the manuscript.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Zhao-Shan Niu, MD, Lab of Micromorphology, Medical College of Qingdao University, 308 Ningxia Road, Qingdao 266071, Shandong Province, China. niumiao1993@hotmail.com

Telephone: +86-532-83780012

Fax: +86-532-83780012

Received: July 22, 2014

Peer-review started: July 23, 2014

First decision: August 14, 2014

Revised: September 2, 2014

Accepted: November 7, 2014

Article in press: November 10, 2014

Published online: January 27, 2015

HCC. In clinical practice, there exists an urgent need for valid prognostic markers to identify patients with prognosis, hence the importance of studies on prognostic markers in improving the prediction of HCC prognosis. This review focuses on the most promising immunohistochemical prognostic markers in predicting the postoperative survival of HCC patients.

Key words: Hepatocellular carcinoma; Management; Immunohistochemical; Prognostic marker; Predictive marker

© The Author(s) 2015. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Hepatocellular carcinoma (HCC) is a leading cause of cancer-related death worldwide. Hepatic resection is generally considered to be one of the most effective therapies for HCC patients, however, the overall post-hepatic resection survival of HCC patients remains unsatisfactory as indicated by the high recurrence rate. Therefore, there is an urgent need to identify prognostic biomarkers for the prediction of postoperative recurrence or metastasis, and to develop better strategies for HCC management. The purpose of this paper is to review the most promising immunohistochemical prognostic markers so far for predicting the postoperative survival of HCC patients.

Niu ZS, Niu XJ, Wang M. Management of hepatocellular carcinoma: Predictive value of immunohistochemical markers for postoperative survival. *World J Hepatol* 2015; 7(1): 7-27 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v7/i1/7.htm> DOI: <http://dx.doi.org/10.4254/wjh.v7.i1.7>

Abstract

Hepatocellular carcinoma (HCC) accounts for over 90% of all primary liver cancers. With an ever increasing incidence trend year by year, it has become the third most common cause of death from cancer worldwide. Hepatic resection is generally considered to be one of the most effective therapies for HCC patients, however, there is a high risk of recurrence in postoperative

INTRODUCTION

Hepatocellular carcinoma (HCC) is a leading cause

of cancer-related death worldwide, with an increasing incidence^[1]. The major risk factor associated with HCC is liver cirrhosis, which is predominantly caused by chronic B virus (HBV) and/or hepatitis C virus (HCV) infections, aflatoxin B1 exposure, and alcoholic liver disease. It is estimated that HBV and HCV account for approximately 75%-80% of HCC cases worldwide. In particular, chronic HBV infection is a predominant risk factor for HCC in Asia and Africa^[2]. Hepatic resection (HR) is a potentially curative and popular therapy for HCC patients^[3], however, the postoperative outcome remains unsatisfactory, with a 5-year post-HR recurrence rate of approximately 80%^[4,5]. In fact, the high postoperative incidence of recurrence is the most frequent cause of postoperative death in HCC patients, and the main reason for the low postoperative survival rate is either intrahepatic metastasis or metachronous multicentric HCC^[6].

So far, it has been still difficult to predict the probability of HCC metastasis and post-HR recurrence. There have been many studies on the risk factors contributing to post-HR recurrence of HCC where a number of prognostic factors related to clinicopathological parameters of HCC have been considered, including tumor size, stage, and grade. Due to lack of a systemic/uniformed approach, these researches results are not consistent. More investigations are required in the search for better markers for HCC prognosis, as a better prediction of postoperative recurrence or metastasis ultimately helps develop better strategies for HCC management.

Immunohistochemistry (IHC) is the most widely applied pathological technique in determining the expression status of tumor-associated proteins and in studying the prognostic and clinical relevance of biomarkers^[7-9]. In spite of the paramount importance of IHC in determining the utility of a biomarker in clinical practice, the lack of universally accepted standardization guidelines has rendered the translation of promising biomarkers into clinical application. Having elaborated on nearly all the promising biomarkers so far in the main body of this review, we will discuss in conclusion the various limitations and technical challenges that need to be addressed when validating *via* IHC a predictive biomarker for clinical endpoint. More specific to HCC, although many immunohistochemical markers have been reported to have a prognostic value for HCC patients, some of which are also validated as independent prognostic markers, so far, there has been no consensus on how these markers could add prognostic value to the clinical parameters. An ideal IHC biomarker for HCC needs to be repeatable, with strong localized staining, valid across a number of patient groups and HCC subtypes, easily quantifiable, and associated with clear clinical outcome measures. Based on our extensive review of relevant literature (Table 1), this review intends to find out why no immunohistochemical markers are applicable in clinical practice, and focuses on the most promising immunohistochemical markers among existing ones in

predicting the postoperative survival of HCC patients.

TUMOR SUPPRESSORS

Tumor suppressor p53

Alteration of p53 is one of the most frequent genetic changes found in HCC, and the biological function of p53 in tumor initiation and progression has been well characterized^[10]. Numerous studies have investigated the prognostic value of p53 protein expression in HCC patients, but reports on the prognostic significance of p53 protein in HCC are often inconsistent and even conflicting, making it difficult to assess the clinical benefit of p53. So far, many studies have demonstrated that p53 protein expression is closely related to the occurrence, progression, metastasis, and survival of HCC. The over-expression of p53 protein is not only closely related to clinicopathological parameters, such as poorly-differentiated HCC, advanced HCC stages^[11,12], but also to microvascular invasion, portal vein invasion, and high risk of tumor recurrence, and overall survival (OS) as well as recurrence-free survival (RFS) post-HR^[13-16], especially within the first year post-HR in HCC patients^[17]. Collectively, these findings indicate that the presence of p53 over-expression in HCC is identified as a major risk factor associated with the aggressive behavior of tumor, as well as a significant predictive marker for postoperative recurrence and survival in HCC patients^[18].

Nevertheless, in some reports with either univariate or multivariate analysis, p53 protein expression in HCC has not been found to be an independent prognostic indicator of survival, despite that the over-expression of p53 protein is more frequent in tumors with poor cellular differentiation^[19], > 5 cm in diameter^[20], and vascular invasion^[21]. Having said that, tumor differentiation and tumor size \geq 5 cm and vascular invasion are reported to be at high risk of HCC recurrence postoperatively^[22-24], and they are independent poor prognostic factors for OS and disease free survival (DFS) in post-HR HCC patients^[25,26]. These findings indicate that p53 expression in HCC may serve as a marker of a more aggressive behavior, and it could have an indirect adverse impact on survival.

Aiming at establishing whether those conclusions could provide solid grounds for applying p53 protein into prognostic clinical practice, the authors of this review carefully studied and compared the included studies. To our surprise, we have noticed several drawbacks in those studies that may affect the reliability of their own conclusions.

To begin with, variation in the immunohistochemical methods with respect to specific antibody clones, dilutions, antigen retrieval methods, as well as the cut-off values for positive expression, could have significant impact on the analysis of the prognostic value of p53 detection in HCC. Most studies used the monoclonal DO-7 antibody, with dilution ranging from 1:50 to 1:100, and citrate buffer for antigen retrieval, neither of which seems to have

Table 1 Immunohistochemical markers of hepatocellular carcinoma associated with prognosis in this review

Marker	Association with poor prognosis	Quoted literature examples
Tumor suppressors		
Mutant p53	Increased expression	Schöniger-Hekele <i>et al</i> ^[18]
Proliferation associated proteins		
Ki67 (detected by Mib1)	Increased expression	Schmilovitz-Weiss <i>et al</i> ^[40]
Proteins associated with angiogenesis		
CD105	Increased microvessel density	Yao <i>et al</i> ^[57]
Proteins involved in angiogenesis		Tseng <i>et al</i> ^[111]
VEGF	Increased expression	
MMPs (matrix metalloproteinases)		
MMP-2 and MMP-9	Increased expression	Xiang <i>et al</i> ^[74] ; Nanashima <i>et al</i> ^[48]
Molecules involved in cell adhesion		
E-Cadherin	Decreased expression	Cho <i>et al</i> ^[94]
CD44 (CD44s and CD44v6)	Increased expression	Ryu <i>et al</i> ^[112] ; Endo K <i>et al</i> ^[113]
OPN	Increased expression	Huang <i>et al</i> ^[130]
Cell cycle regulators		
p27 (Kip1)	Decreased expression	Wan <i>et al</i> ^[137]
DNA-binding nuclear protein		
HMGB1	Increased expression	Xiao <i>et al</i> ^[171]
Cancer stem cells		
CD133	Increased expression	Chan <i>et al</i> ^[188]
EpCAM	Increased expression	Chan <i>et al</i> ^[188]
CK19	Increased expression	Xu <i>et al</i> ^[197]
Cell surface proteins		
GPC3	Increased expression	Fu <i>et al</i> ^[211]
mTOR Pathway	Increased expression	Baba <i>et al</i> ^[223]

VEGF: Vascular endothelial growth factor; MMP-2: Matrix metalloproteinases 2; CD44s: CD44 standard isoform; CD44v6: CD44 variant isoforms; OPN: Osteopontin; HMGB1: High-mobility group box 1 protein; EpCAM: Epithelial cell adhesion molecule; CK19: Cytokeratin19; GPC3: Glypican-3; mTOR: Mammalian target of rapamycin.

Table 2 p53 antibody used in different studies in this review

Ref.	Clone	Source	Dilution	Antigen retrieval	Cut-off value ¹
Tseng <i>et al</i> ^[111]	DO-7	DAKO	1:100	Citrate buffer	> 5% nuclear p53 staining
Hu <i>et al</i> ^[123]	DO-7	DAKO	1:1000	Citrate buffer	> 10% nuclear p53 staining
Kang <i>et al</i> ^[14]	DO-7	DAKO	1:100	Citrate buffer	> 5% nuclear p53 staining
Stroescu <i>et al</i> ^[16]	DO-7	DAKO	Not reported	Citrate buffer	< 24% nuclear p53 staining
Sung <i>et al</i> ^[17]	Bp53-12	Zymed	1:80	Citrate buffer	> 5% nuclear p53 staining
Qin <i>et al</i> ^[19]	DO-7	DAKO	Not reported	Citrate buffer	≥ 10% nuclear p53 staining
Guo <i>et al</i> ^[20]	CM1	SDC	1:2000	Citrate buffer	> 5% nuclear p53 staining
Umemura <i>et al</i> ^[21]	DO-7	DAKO	1:50	Citrate buffer	≥ 10% nuclear p53 staining

¹Immunohistochemical cut-off value indicates the percentage of cells with p53 positively staining nuclei. DAKO: Dako Denmark A/S, Glostrup, Denmark; Zymed: Zymed Lab Inc, CA, United States; SDC: San Diego, CA, United States.

any impact on the association between p53 expression and prognosis. In the meantime, we have noticed that different researchers adopted different cut-off values for determining positive p53 expression without any explanation or justification, which has significantly affected the association between p53 expression and prognosis in HCC (Table 2). Since p53 protein expression as detected by IHC does not always reflect the presence of mutant p53 protein, the predictive value of p53 IHC in detecting *TP53* mutations is currently under debate. So far, an optimal threshold is yet to be defined.

In general, a cut-off value of > 10% p53 immunopositive cells appears to be predictive of *TP53* mutations in HCC^[27].

What's more, some studies used retrospective analyses in small series of patients. Naturally, without sufficient resolution and reproducibility, it is unlikely to accurately predict disease progression by means of these study designs.

Furthermore, inappropriate proportion of important variables was included in some studies, such as tumor grade, tumor size, tumor stage. For example, too many cases for Edmondson-Steiner Grade I, tumor-node-metastasis (TNM) stage I, or tumors ≤ 5 cm in diameter were selected, which easily resulted in the comparatively low positive rate of p53. And the reliability of their conclusions suffers.

Finally, we have noticed that compared with HCV

Table 3 Clinicopathological parameters affecting the association between p53 expression and prognosis in this review (*n*)

Ref.	Number of patients	Positive rate (%)	HBsAg/HCVAb positive	Edmondson grade		TNM stage		Tumor size	
				I + II	III + IV (1)	I + II	III + IV	≤ 5 cm	> 5 cm
Tseng <i>et al</i> ^[11]	113	37.1	79/34	84	29 (1)	54	59	Not reported	
Hu <i>et al</i> ^[13]	124	41.9	83/30	20/38/13 (2)		61	63	Not reported	
Kang <i>et al</i> ^[14]	83	96.4	59/8	27	56 (1)	Not reported		57	26
Stroescu <i>et al</i> ^[16]	47	68	40/0	19	28 (1)	Not reported		20	27
Sung <i>et al</i> ^[17]	105	19	82/6	78	27 (1)	Not reported		> 3 cm	52
Qin <i>et al</i> ^[19]	113	22	40/25	55	58 (1)	Not reported		48	55
Guo <i>et al</i> ^[20]	104	34.6	14/55	18/56/31 (2)		67	37	Not reported	
Umemura <i>et al</i> ^[21]	90	33.3	Not reported	65	25 (1)	Not reported		37	53

TNM: Tumor-node-metastasis; HBsAg: Hepatitis B surface antigen; HCVAb: Hepatitis C virus antibody.

infection, where HCCs were caused mainly by the synergistic effect of HBV infection and aflatoxin B1, studies are more likely to confirm the over-expression of p53 and its prognostic value in HCC (Table 3). This has been partly echoed by studies on the relationship between p53 and pathogenic factors. HBV infection and exposure to AFB1 have been demonstrated to induce the point mutation of p53 in HCC tissue^[28], especially exposure to AFB1 can affect the over-expression of p53 in the development of HBV-associated HCC^[29]. Other studies also reported p53 protein expression in HCC has racial and regional differences^[30]. Therefore, there is a higher chance of reaching a more reliable conclusion on the prognostic value of p53 protein in HCC, researchers should consider HCC cases induced by the same or similar pathogenic factors.

The detection of p53 expression by using IHC has another noteworthy problem. The p53 protein expression as detected by IHC does not always reflect the mutation status of TP53, with one cause being that not all mutations always result in stable protein formation, and another being that some tumors may also express wild-type p53. Nevertheless, in fact, lack of standardized IHC may be partly responsible for the inconsistencies in frequency of p53 mutations and p53 protein levels. TP53 most often has missense rather than truncating mutations, and IHC antibodies will always have difficulty in detecting proteins with a small number of missense amino acid substitutions. Therefore, the studies with high p53 expression by IHC may reflect both high wild-type and mutant p53. Given this, when determining p53 status in HCC, we should analyze it by standardized IHC in combination with p53 mutation analysis.

In conclusion, p53 protein expression comes short to be recommended as a universal predictive marker for survival in HCC patients, speaking from the available evidence. The prognostic value of p53 protein expression in HCC may vary according to different racial and regional groups. In area where HBV infection and AFB1 account for the major attributive risk of HCC, such as western Africa and south-east China, p53 protein tends to be high expression, and could be considered as a predictive marker for survival in HCC patients. Nevertheless, in order to

identify the actual prognostic value of p53 expression in HCC, further studies are required by standardized IHC with larger populations, uniform pathological samples, homogeneous patient populations. It is also worthwhile to point out that it would help us lead to a sound conclusion the studies should include a > 10% nuclear staining as a cut-off value of p53 expression.

Due to the diversity and complexity in the research conclusions on p53, Tables 2 and 3 have been created to help with understanding. These two tables are of reference value in the following discussions on the rest of markers in this review, and hence will not be repeated.

PROLIFERATION MARKERS

The proliferative activity of tumor cells is an important indicator for assessing aggressiveness and could be useful for predicting clinicopathological and prognostic significance. Many antigens, such as proliferating cell nuclear antigen (PCNA) and Ki-67, have been used as proliferation markers for cancer cells. Compared with assessments by Ki-67, cell growth fraction is often overestimated when assessed by PCNA. Thus Ki-67 is considered a more accurate marker for the proliferative stage of tumor cells than PCNA^[31,32].

Ki67

Ki-67 is a nuclear non-histone protein initially expressed in cell-cycle phases G₁, S, G₂ and mitosis, and absent in the G₀ phase. The expression of the Ki-67 protein in humans is closely associated with cell proliferation. Naturally, Ki-67 is an excellent marker for proliferating cells^[33]. MIB-1 is a monoclonal antibody that identifies Ki-67 protein in paraffin-embedded tissue. Numerous studies have shown that Ki-67 immunohistochemical staining is an effective method to predict prognosis in various tumors.

Ki-67 expression is significantly associated with histological grade of HCC patients^[34,35], in other words, the increased expression of Ki-67 in poorly differentiated tissues implies that the single fact of tumor cells losing growth control in hepatocarcinogenesis is a reflection of malignant behavior of tumor cells. Therefore, Ki-67

is an objective indicator of the proliferative ability of HCC cells, and can serve as an important index of the proliferation and differentiation of HCC cells. In addition, Ki-67 expression is significantly higher in HCC cases with shorter DFS; The same applies to the HCC cases with biologically aggressive features such as advanced stages, portal invasion and intra-hepatic metastasis^[36]. Therefore, Ki-67 expression could serve as a useful marker for evaluating the progressive activity and predicting DFS in HCC patients. Furthermore, multivariate analysis shows that Ki-67 expression is an independent prognostic factor for DFS and OS^[35]. Hence it's been concluded that the expression of Ki-67 is an independent prognostic indicator for post-HR HCC patients^[37-40].

In short, Ki-67 expression is an objective factor for predicting survival for post-HR HCC patients, and it could be considered a promising independent prognostic immunohistochemical marker in HCC patients. Therefore, Ki-67 should be taken into consideration when making decisions on adjuvant therapy. HCC patients with high expression of Ki-67 protein may need intensive surveillance and adjuvant therapy.

In spite of the above discussions, lack of standardized IHC and cut-off value has hindered Ki-67 from routine clinical application. Different studies use different methods of antigen retrieval, antibodies concentrations; In addition, the time of incubation varies from study to study; as to cut-off value, some studies have chosen median values while others an arbitrary value (*e.g.*, 10%, 20% and so on) without any explanation or justification. All of these significantly influence the final results. Ironically, the choice of the cut-off value has a major impact on clinical practice, simply because it determines which patients are classified as "high Ki-67 expression"-those who in turn have a poorer prognosis should generally receive more aggressive therapy. We believe future researchers should work towards a standardized IHC and validated cut-off level before Ki-67 could be established as a reproducible and robust prognostic factor in HCC.

To throw in some light, a study has demonstrated that when determining the clinically relevant threshold for immunohistochemical tumor positivity, receiver operating characteristic (ROC) curve analysis could be a reproducible and reliable alternative in selecting and validating cut-off scores^[41]. The term "ROC" came from tests of the ability of World War II radar operators to determine whether a blip on the radar screen represented an object (signal) or noise. At present, ROC curve analysis is a well established analytic tool and has been widely applied in various fields, including Medicine. Applications in a number of cancers have proved that cut-off scores based on ROC curve analysis guarantee maximum sensitivity and specificity, and therefore allow the greatest number of tumors to be correctly classified as carrying or not carrying the clinical outcomes^[42,43].

Therefore, we propose that Ki-67 cut-off value

should be set up according to ROC curve analysis.

MARKERS OF ANGIOGENESIS

Markers of Microvascular Density

Angiogenesis is critical for the growth, invasion and metastasis of cancers. Microvascular density (MVD) is commonly used to assess tumor neovascularization. This is especially true in HCC, characteristically a highly vascular tumor. However, there are conflicting reports in regard to whether MVD in HCC is associated with prognosis. This could be explained by the fact that different studies use different antibodies to calculate MVD.

The evaluation of MVD is generally identified by immunohistochemical staining of endothelial cells with the so-called pan-endothelial cell markers, such as CD34, CD31, and von Willebrand factor. Among them: Firstly, MVD appears to be better assessed by CD31 than by von Willebrand factor (vWF)^[44]. Secondly, antibody against CD31 fails to stain sinusoid endothelial cells in many HCC cases, therefore the prognostic value of CD31 could at most be used as a marker of vascular changes in the liver^[45]. Thirdly, although CD34 has proven to be a more sensitive and specific endothelial cell marker for microvessels in HCC^[46], MVD determined by CD34 appears to be closely correlated with the prognosis of HCC^[11,47] in some studies, while such correlation is not identified by others^[48,49]. Differences in methodology, *i.e.*, different counting techniques, selection of microvessels, *etc.*, contribute to the conflicting results. The non-specificity in CD34 determines that CD34 can not be an ideal marker for neovascularization. In addition, all the above mentioned markers react with not only newly formed vessels but also normal vessels trapped within tumor tissues.

The conclusion is the MVD identified by anti-pan-endothelial antibodies is not an ideal prognostic marker^[50].

The good news is MVD assessment using CD105 as marker (CD105-MVD) has demonstrated a higher MVD specificity in tumor tissues, and it has been more widely adopted, compared with vWF, CD31, or CD34^[51-53], as a predictor for progression and prognosis in a variety of cancers.

Endoglin

Endoglin (CD105) is a transforming growth factor- β co-receptor mainly expressed in the endothelium of tissues' blood vessels, particularly in de novo formed blood vessels within tumor. It has been used as a marker for tumor angiogenesis, with a potential for prognostic prediction^[54,55].

In HCC, some studies have demonstrated CD105 excels CD34 in marking new microvessels in HCC^[56,57]. When median scores of MVD are used as cut-off points, patients with higher score of MVD-CD105 have a significantly poorer prognosis in either DFS or OS analysis, whereas similar prognostic significance of MVD-CD34 is

observed only in DFS analysis^[57]. One study reveals that no prognostic significance is observed when median values are used as cut-off points using either IMVD-CD105 or IMVD-CD34, however, the presence of the diffuse pattern of CD105 expression in the adjacent non-tumorous liver tissues can predict a poorer DFS^[58]. Collectively, compared with CD34-MVD, CD105-MVD is a significant and independent prognostic indicator for recurrence and metastasis in HCC patients. Having said that, some study found that MVD-CD105 did not show prognostic influence in a cohort of predominantly large HCCs (> 5 cm)^[58]. Further studies need to be conducted in larger cohorts of patients with a longer follow-up period.

In summary, CD105, by specifically staining newly formed tumor vessels, is a promising and independent prognostic marker for HCC, which could in turn lead to future therapeutic trials with antiangiogenic therapy. To date, however, the lack of commonly accepted objective criteria in counting microvessels under light microscopy has hampered the clinical use of tumor MVD for prognostication. The authors of this review propose that, before other better microvessel counting methods have been established, microvessel counting should be performed in accordance with Weidner's standards^[59].

MARKERS OF PROTEINS INVOLVED IN ANGIOGENESIS

Vascular endothelial growth factor

Angiogenesis is crucial for tumor growth and metastasis, and could be stimulated by several regulators, among which vascular endothelial growth factor (VEGF) seems to be most important^[60]. The VEGF family comprises six glycoproteins: VEGF-A, VEGF-B, VEGF-C, VEGF-D, VEGF-E, and placental growth factor. These major VEGF subtypes are in the nature of multiple isoforms. The best representative of VEGF family is VEGF-A (commonly referred to as VEGF). VEGF mediates its angiogenic effects *via* several different receptors, including VEGFR1 and VEGFR2^[61].

VEGF plays an important role in tumor angiogenesis and progression, including HCC, and elevated VEGF levels in serum and tissues are related to poor prognosis in HCC patients^[62]. So far, numerous studies have explored and confirmed the prognostic value of VEGF for survival in HCC patients. Some studies found the VEGF over-expression was closely correlated with MVD, high alpha-fetoprotein levels, tumor size, dedifferentiation, advanced TNM stage, vascular invasion, capsular invasion, intrahepatic metastasis, and lymph node metastasis (LNM)^[63,64]. These findings suggested that VEGF over-expression was useful in predicting progression, metastasis, and recurrence of post-HR HCC^[65,66]. In addition, survival analyses indicated that VEGF over-expression was an independent factor for poor-prognosis DFS and OS^[11]. Therefore, VEGF expression in HCC tissues could be regarded as a valuable indicator in estimating prognosis of HCC patients^[20].

More recent studies suggested that co-expression of platelet-derived growth factor receptors- α , PDGFR- β and VEGF could be considered an independent prognostic biomarker for predicting DFS and OS in HCC patients, and that this co-expression could be used to identify patients at a higher risk of tumor recurrence and poor prognosis, and to select therapeutic schemes for HCC treatment^[67]. In addition, the co-index [VEGF/platelet-derived endothelial cell growth factor (PD-ECGF)] was an independent prognostic factor for OS and RFS; Furthermore, the co-index of VEGF and PD-ECGF was a promising independent predictor for recurrence and survival of alpha-fetoprotein (AFP)-negative HCC patients after curative resection^[68].

In spite of all the research efforts in establishing VEGF expression status as a promising prognostic marker, there is still a long way to go before the findings could have any impact on clinical practice.

Metalloproteinase

Matrix metalloproteinases (MMPs) comprise a large family of zinc- and calcium-dependent proteolytic enzymes that have been repeatedly implicated in invasion and metastasis. MMPs are capable of degrading most components of the extracellular matrix (ECM), including the basement membrane which serves as a barrier between tissue compartments^[69]. Type IV collagen (Col IV) is a major component of the ECM and basement membrane, and plays an important role in regulating and limiting tumor invasion and metastasis^[70]. Among MMPs, MMP-2 (gelatinase A) and MMP-9 (gelatinase B) are of particular importance as far as tumor invasion and metastasis are concerned, because they are capable of degrading ColIV^[71,72]. Furthermore, high MMP-2 or MMP-9 expression in tumor or stromal cells might serve as a poor-prognosis predictor in various cancers^[72,73]. This applies to HCC according to some researches.

MMP-2: Quite a few researches concluded that high intratumoral MMP-2 expression in HCC was correlated with high Edmondson grade, advanced TNM stage, and barcelona clinic liver cancer stage^[66,74], and that MMP-2 was related to HCC invasion, metastasis and recurrence^[75]. As a result of these research findings, it is widely acknowledged that MMP-2 expression could serve as a predictive marker for HCC progression, metastasis, and recurrence, and that MMP-2 expression is an independent prognostic factor for DFS and OS in HCC patients with LNM^[74].

MMP-9: The expression of MMP-9 in HCC was proved by a number of researches to be closely correlated to tumor nodule, vein invasion, advanced TNM stage, extrahepatic metastasis, and the formation of portal vein tumor thrombus^[48,76,77]. These researches suggest that the expression of MMP-9 reflects the biologically aggressive behavior of HCC, and that MMP-9 is an important molecule which participates in the progression,

metastasis and invasion of HCC. Some studies demonstrated that MMP-9 expression was up-regulated in HBV-associated HCC compared with HCV-associated HCC^[78]. Other studies concluded that MMP-9 expression was a significant predictive factor for post-HR recurrence in HCC patients with the background of HBV^[79]. Still other studies found that increased expression of MMP-9 protein was an independent prognostic factor after HCC resection^[48].

It is worthwhile to note that, when both MMP-2 and MMP-9 were analyzed in the same set of patients, MMP-2 was predominantly involved in hepatocarcinogenesis and progression, while MMP-9 was predominantly involved in the capsular infiltration and portal vein invasion^[80]; and that the MMP-2 expression only had weak correlations to HCC recurrence, while positive MMP-9 expression was an independent recurrence-risk factor^[25]. Moreover, multivariate analysis confirmed that MMP-9 expression was an independent predictor of time to recurrence (TTR) and OS, whereas high MMP-2 expression was only correlated with TTR^[81]. This suggests that MMP-9 is superior to MMP-2 in predicting tumor recurrence and survival in post-HR HCC patients.

One study concluded that high expression of MMP-9 and MMP-2 in peritumoral stromal cells was related to poorer prognosis in HCC patients^[82]; However, this was overturned by another study^[83]. Still, one study found that MMP-2 or MMP-9 expression was not related with the histological differentiation of HCC^[84]; And yet another study claimed that MMP-2 and MMP-9 protein could serve as independent prognostic factors for poor survival regardless of the age, tumor size, tumor grades, TNM classification^[85].

The question is: What has contributed to the discrepancies in those research findings? It could be a long list that includes the differences in pathological samples, antibodies used, different IHC methods, different patient populations and different cut-off values. It is advisable that further studies enroll larger scale of clinical HCC samples and use standardized IHC. This has an add-on value of ultimately benefiting the clinical application of MMP inhibitors as chemopreventive and antiangiogenic drugs.

ADHESION MOLECULES

E-cadherin

Tumor progression is characterized by loss of cell adhesion and increase of invasion and metastasis. Cell adhesion molecules play a significant role in cancer progression and metastasis^[86]. E-cadherin is a key molecule for the maintenance of intracellular adhesion, and down-regulation of this protein has been associated with tumor progression in diverse human cancer types^[87,88].

Many researches have concluded that E-cadherin expression is very weak in HCC tumors but very strong in the cell membranes of non-tumor tissues, and E-cadherin expression is significantly correlated inversely

with histological grade, *i.e.*, with the highest in well-differentiated^[89] as put in one study, or the increased loss of E-cadherin expression is observed particularly in poorly-differentiated^[90] as put in another; In addition, low expression of E-cadherin in HCC is also related to pathological stage and later TNM stage^[91]. Therefore, it is safe to say that low expression of E-cadherin is a strong indicator of malignant HCC progression. There are also some researches that suggest low expression of E-cadherin is significantly associated with intrahepatic metastasis and regional lymph node metastasis^[92,93]. When you combine the findings of these two types of researches, it seems natural to conclude that loss of E-cadherin expression in HCC could predict a high risk of post-HR recurrence^[94]. Taken together, these findings indicate that detection of E-cadherin expression could be useful in predicting HCC prognosis.

On the opposite side, two studies revealed low expression of E-cadherin had no direct correlation with the post-HR recurrence^[95], and it did not predict poor survival even when there was increased loss of E-cadherin in tumors of higher histologic grade^[96]. The researchers themselves admitted insufficient number of and lack of homogeneity in the included patients could have contributed to the opposite findings^[95]. Another two studies confirmed atypical cytosolic expression of E-cadherin or high E-cadherin membrane/cytoplasm ratio was correlated with a poorer patient prognosis^[97,98].

Decreased expression of E-cadherin has been found in all three types of epithelial-mesenchymal transition (EMT) and is thought to be the prototypical marker of EMT^[99]. EMT has been shown to be a pivotal mechanism contributing to cancer invasion and metastasis, as epithelial cells lose their polarity and acquire the migratory properties of mesenchymal cells. The characteristic changes during EMT include the downregulation of epithelial markers such as E-cadherin and the upregulation of mesenchymal markers such as vimentin^[100]. The EMT of HCC cells is thought to be a key event in intrahepatic dissemination and distal metastasis^[101]. A recent study suggests that the loss of E-cadherin followed by the overexpression of vimentin may play a vital role in the invasive and metastatic phenotype and in the process of EMT, leading to unfavorable outcomes in patients with HCC^[102].

The authors of this review carefully studied all the related articles, in the course of which differences in antibodies, cut-off values, or race stood out. Further investigation is necessary for assessing these discrepancies.

CD44

CD44, is a transmembrane glycoprotein and has been implicated in numerous biological processes, including cell-cell interactions, cell adhesion, and cell migration^[103]. Through alternative mRNA splicing, cells produce numerous CD44 protein isoforms: standard isoform (CD44s) and variant isoforms (CD44v). CD44s is a cell adhesion molecule known for mediating cellular adhesion

to the extracellular matrix—a prerequisite for tumor cell migration. Some researchers argue CD44s is involved in invasion and metastasis of various cancers^[104]. Among the CD44 variant isoforms, CD44v6 has reportedly been associated with increased invasion, metastasis, and poor prognosis^[105,106]. Recent studies also suggest CD44 is one of the cancer stem cell markers associated with poor prognosis^[107].

CD44s: Many studies have indicated that high CD44s expression in HCC is correlated with high AFP level, large tumor size, multiple tumors, poor tumor differentiation, advanced tumor stage, portal vein tumor thrombus, and early tumor recurrence or metastasis^[108-110]. These findings suggest that CD44s expression may serve as a predictive marker for HCC progression, metastasis, and recurrence. However, it is not always the case. For example, one study found that there was a significant correlation between CD44s expression and the presence of vascular invasion, but not between CD44s expression and tumor grade, from which the author concluded that high CD44s expression may have implications relating to metastasis and prognosis in HCC patients^[111]. Another study found that statistically Edmondson grades had a significant correlation with CD44s expression, and yet such correlation did not exist between CD44s expression and vascular invasion, from which the conclusion is CD44s expression was significantly correlated with DFS and independent factor in multivariate analysis^[112].

Some other studies suggested either high CD44s expression was a poor prognostic factor following curative HR of primary HCC, including reduced DFS and OS^[108,109], or high CD44s expression was an independent factor for OS^[113]. One study failed to present a significant correlation between patient survival and CD44s expression, however, it did show expression of CD44s as a significant predictable marker for LNM^[110].

The inconsistency in CD44s expression and clinicopathological parameters is obvious, however, all relevant studies endorse that CD44s expression could serve as a predictive marker for HCC metastasis and survival.

CD44v6: Some studies suggest that high expression of CD44v6 is related to aggressive clinical behavior in HCC, more specifically it is correlated with high tumor grades, advanced TNM stage^[114,115]. In addition, CD44v6 overexpression presents a positive correlation with HCC metastatic potential^[116]. These findings indicate that high expression of CD44v6 may serve as a predictive marker for HCC progression and metastasis. As to its relationship with vascular invasion, some studies concluded that high CD44v6 expression significantly correlated with the presence of vascular invasion^[113,115], while one study demonstrated that a low expression level of CD44v6 tended to be associated with vascular invasion^[117]. These studies adopted different scoring systems and cut-off values, which could have contributed to the discrepancy in the results.

In multivariate survival analysis, some studies demonstrated high expression of CD44v6 was significantly correlated with OS and TTR^[115], or that it was an independent factor for OS^[113]. Thus, detection of CD44v6 expression could be useful in predicting prognosis of HCC.

The authors of this review would tentatively recommend that, for CD44v6 expression evaluation, cut-off value be selected on the basis of ROC curve analysis. In addition to a valid cut-off value, future studies should consider a larger sample and a longer follow-up period. Only then could relevant studies add clinical value to CD44v6 expression in HCC.

Osteopontin

Osteopontin (OPN) is a multifunctional secreted phosphorylated glycoprotein that belongs to the small integrin-binding ligand N-linked glycoprotein family, and it is implicated in promoting malignant cell proliferation, detachment, invasive and metastatic progression of many carcinomas^[118-120]. The expression level of OPN is elevated in a variety of human cancers, particularly those that metastasize preferentially to the skeleton^[121]. Recent studies have indicated that OPN is involved in HCC progression and metastasis.

It is widely acknowledged that OPN expression is localized predominantly in the cytoplasm, and OPN expression in HCC is stronger than those in paracarcinoma tissues and normal liver tissues^[122]. And that higher expression of OPN in HCC is closely associated with poor differentiation and advanced tumor stage^[123,124]. And that it is positively correlated with tumor size, capsular invasion, portal vein tumor thrombus, lymph node metastasis^[122,125,126]. Therefore, it is safe to say OPN could serve as novel biomarker for monitoring HCC progression and metastasis.

In addition, numerous studies have suggested OPN could serve as a useful marker for predicting early recurrence in HCC patients^[122,127], and that OPN could help determine whether individual patient needs adjuvant therapy to prevent early post-HR recurrence^[128], and that OPN expression is an independent prognostic factor either for DFS in HBV-positive small HCC (< 5 cm)^[129], or for OS and DFS in patients with the TNM stage I HCC^[127]. These findings suggest that OPN could be solely identified as an independent prognostic biomarker for post-HR HCC patients^[130].

Recent studies have suggested that the combination of OPN and some other markers seem promising for HCC prognosis. For example, the combinations of tumor OPN with either caspase-3, or Bcl-2, or CD44, have all been announced as promising independent predictors of tumor recurrence and survival in HCC patients^[130,131]. It is especially true for those with early-stage disease when tumor OPN is combined with microenvironment-associated peritumoral macrophages^[132]. Nevertheless, the interaction between tumor OPN and these markers, which facilitates tumor progression and metastasis, still

remains unclear in clinical practice. Further large-scale studies are required to confirm their clinical value^[133].

CELL CYCLE REGULATORS

p27

The functional alterations of cell-cycle regulators, such as Cyclin Dependent Kinases (CDK) and their inhibitors, occur frequently in cancers. As a critical CDK inhibitor, p27 (Kip1) is involved in G1 phase progression, and is widely regarded as adverse prognostic biomarker for various types of cancers, since decreased or absent expression of p27 (Kip1) is frequently observed in various types of human cancers with poor prognoses^[134,135]. It has been reported that p27 (Kip1) is exclusively inactivated by proteasome-mediated protein degradation^[136]. p27 (Kip1) is frequently inactivated in HCC and is considered a potent tumor suppressor.

So far, many studies have reported that decreased p27 expression is significantly lower in HCC than those in the adjacent noncancerous tissues or in normal liver tissues, and it is a risk factor in HCC^[137-139]. Furthermore, some studies have indicated decreased p27 expression is closely related to the aggressive HCC tumor behaviors^[139,140]. In addition, some studies indicated that p27 expression was decreased in advanced cases in a series of curatively resected HCCs^[141], and the p27 labeling index was significantly decreased in the cases with advanced tumor stages, portal invasion, poor differentiation, larger size, and intrahepatic metastasis^[142,143]. As concluded by a researcher, p27 expression in HCC could act as an independent predictor of post-HR recurrence^[142].

It has been reported that in multivariate analysis, p27 expression could be recognized as an independent prognostic marker for OS^[144], and OS and loco-regional recurrence-free^[145], which suggests low expression level of p27 is associated with significantly worse prognosis in HCC patients^[137,146]. Similar findings have been reported that high expression of p27 is a favorable independent prognostic parameter^[147]. Taken together, p27 could be regarded as a powerful clinical indicator for prognosis prediction in individual HCC patient.

An interesting point is that it is in both nucleus and cytoplasm that tumor cells were found to have expressed p27 protein^[148]. The significance of cytoplasmic p27 protein is still under debate, and cytoplasmic p27 protein is rarely considered in assessing p27 IHC score. Decreased or absent expression of p27 (Kip1) in nucleus is frequently observed in various types of human cancers with poor prognoses^[149-153]; however, some researchers argue over-expression of cytoplasmic p27 may also serve as a marker for poor prognosis in several types of human cancers^[154-156]. Further studies suggest that the nuclear localization of p27 is essential for its growth-inhibiting function^[157]. When narrowing down to HCC, the expression of p27 is mainly found in nucleus and cytoplasm^[144]. It has been generally accepted that low expression of nuclear p27 protein is associated with

poorer prognosis, while cytoplasmic expression of p27 is positively associated with poor cellular differentiation—the higher the expression, the higher incidence in HCC patients^[140]. This is echoed by a study that concluded cytoplasmic localization of p27 could be an early event during hepatocarcinogenesis^[158].

It remains unclear whether the cytoplasmic staining represents a methodological artifact or a finding of biological and/or prognostic importance. In view of this uncertainty, the authors of this review propose that only nuclear p27 (kip-1) staining for HCC survival analyses be considered in staining evaluation.

Taken together, IHC detection of p27 on routine tissue sections could be useful in predicting survival of individual HCC patient and in determining future therapeutic strategies. Therefore, p27 is worthy of further evaluation as a potential prognostic marker in clinical trial samples of large cohorts.

DNA-BINDING NUCLEAR PROTEIN

High-mobility group box 1 protein

High-mobility group box (HMGB) proteins are non-histone nuclear proteins with different functions in the cell^[159]. HMGB1, HMGB2, and HMGB3 are the members of the HMGB protein family, with HMGB1 being the most important one. While the expressions of HMGB2 and HMGB3 are limited, HMGB1 plays a role in cancer progression, angiogenesis, invasion, and metastasis development^[160]. The function of HMGB1 is complicated by its cellular localization. In nucleus, HMGB1 binds with DNA and serves as a structural component^[161]. Cytoplasmic localization of HMGB1 is associated with the proliferation and metastasis of different tumor types. The process could be dramatically sped up when cytoplasmic localization of HMGB1 binds with the receptor for advanced glycation end products^[162]. As for the “sped up” process, one study has deduced that the interaction between receptor for advanced glycan endproducts and HMGB1 activates mitogen-activated protein kinases, nuclear factor kappa B, and phosphoinositide 3-kinases (PI3K)/AKT signaling pathways to promote cellular proliferation and metastasis^[163].

There are many relevant studies that focus on HCC and their findings include: In HCC cells, downregulation of HMGB1 could remarkably inhibit the growth of HCLM3 cells, as well as their migration and invasion ability^[164]; HMGB1 knockdown inhibited the proliferative activities and metastatic potential of SMMC-7721 cells. That is to say, the expression of HMGB1 was closely correlated with pathological grade and distant metastases of liver cancer, and HMGB1 knockdown inhibited liver cancer growth and metastasis^[165]. In addition, HMGB1-siRNA could inhibit the invasion and migration abilities of human hepatoma cell line HepG2^[166]. In the liver tumor model, stable knockdown of HMGB1 suppressed HCC invasion and metastasis^[167]. In detection of serum

HMGB1, serum HMGB1 was positively correlated with clinicopathological features in HCC patients, higher serum HMGB1 level was correlated with bigger tumor size, poor Edmondson grade and advanced TNM stage^[168]. Collectively, these findings suggest that HMGB1 in HCC is significant in tumor progression, invasion and metastasis.

In recent years, many studies have explored the clinical significance of HMGB1 expression in various human tumors, including HCC. Some study reported that over-expression of HMGB1 was significantly associated with HCC incomplete encapsulation and advanced TNM stage^[169]; similarly, another study demonstrated that, by detecting fresh samples, over-expression of MGB1 mRNA was correlated with HCC high Edmondson grade, advanced TNM stage, vascular invasion and capsule invasion^[170]. These findings indicate that over-expression of HMGB1 is associated with HCC tumor growth and invasion.

Recent studies have also demonstrated the expression of HMGB1 could serve as an independent prognostic factor for poor OS and DFS for post-HR HCC patients; more importantly, subgroup analysis showed the expression of HMGB1 was significantly associated with poor prognosis in HCC patients > 5 cm, but not in HCC patients ≤ 5 cm^[169]. This trend suggests that HMGB1 could be an important prognostic marker for late stage HCC; in addition, multivariate analysis has also concluded that HMGB1 expression is a key independent prognostic factor that could be associated with OS of HCC patients^[171]. Therefore, HMGB1 expression could be taken as an independent predictor of prognosis for post-HR HCC patients. However, further studies are necessary before we could tell for sure whether HMGB1 is a reliable clinical predictor of survival for individual post-HR HCC patient.

STEM CELL MARKERS

In recent years, many findings have suggested that tumors are comprised of heterogeneous cell populations, only a small fraction of which are tumorigenic with the ability to self-renew and produce phenotypically diverse tumor cell populations^[172]. Cells in this fraction are called cancer stem cells (CSCs) or tumor-initiating cells or cancer progenitor cells, and they have the ability to self-renew, proliferate, and maintain the neoplastic clone. Accumulating evidence has shown that these CSCs have long-term proliferative potential and the ability to regenerate tumors with phenotypically heterogeneous cell types, and that these CSCs are important mediators of tumor metastasis and cancer relapse^[173].

So far, various cell surface and transmembrane proteins expressed by CSCs have been identified, including CD44, CD47, CD123, epithelial cell adhesive molecule (EpCAM) (CD326), CD133^[174]. In HCC, the three major types of liver CSCs (LCSCs) are dedifferentiated hepatocytes, hepatic oval cells, and bone marrow cells.

To date, CD133, CD90, and EpCAM, CD44, CD24, and CD13 have been identified as specific antigenic markers for HCC stem cells^[175]; The oval cell-specific marker (OV6) is identified as a marker for hepatic oval cells^[176], in addition, cytokeratin 7 (CK7) and CK19 are identified as markers for dedifferentiated hepatocytes^[177].

LCSCs can be observed by IHC and electron microscope. In HCC, the phenotypes of LCSCs express as OV6, CK7, CK19, CD133 and EpCAM^[178]. There have been a number of studies reporting that the expression of LCSCs markers in HCC is associated with poor clinical outcome after surgical resection^[179,180]. Among them, the expression of EpCAM, CK19 and CD 133 has demonstrated association with intrahepatic recurrence in HCC patients^[181].

To our knowledge, EpCAM, CK19, and CD 133 have been so far the most widely studied LCSCs markers in HCC using IHC.

CD133

Prominin 1 (CD133) is a pentaspan transmembrane glycoprotein with uncertain physiological function, and it is often expressed by various epithelial and non-epithelial cells, notably by stem and cancer stem cells. CD133 is currently recognized as a marker for LCSCs^[182-184]. A number of studies have demonstrated *via* IHC that CD133 expression is associated with poorer tumor grade and advanced tumor stage^[185]; Moreover, CD133 expression is associated with the absence of tumor capsule; and CD133 tends to be expressed in tumors showing stronger potential for invasion and metastasis^[186]. These findings suggest that CD133 expression is associated with HCC progression, invasion and metastasis. In addition, several studies have demonstrated that CD133 expression is a significant risk factor for the OS of HCC patients, especially patients with Stage III and IVA HCC^[187]; And that Cox proportional hazard model has shown that CD133 expression is an independent predictor for DFS^[177]; and the multivariate survival analysis has demonstrated that CD133 expression is an independent adverse prognostic factor for OS and DFS, especially for patients with early-stage HCC^[188]. All the above mentioned studies agree to the basic point that increased CD133 expression could serve as an independent prognostic factor for survival in HCC patients.

EpCAM

EpCAM, also known as 17-1A, GA733-2, KSA, ESA, and EGP-40, is a type I transmembrane glycoprotein and acts as a homotypic calcium-independent cell adhesion molecule. It is expressed in almost all carcinomas. EpCAM is currently recognized as a marker for LCSCs^[189-191]. Many studies have demonstrated *via* IHC that EpCAM expression is associated with younger age^[181], poorer histological differentiation, vascular invasion and/or more advanced stage^[180,188,192]. These findings suggest that EpCAM expression is associated with HCC progression. Furthermore, several studies have demonstrated that EpCAM expression could serve as an independent factor

for DFS in HCCs at all stages^[188]; And the multivariate survival analysis has demonstrated that EpCAM expression is a significant predictor for shorter survival time in HCC patients^[186], especially patients with T1 HCC^[180]. Taken together, increased EpCAM expression could serve as an independent prognostic factor for survival in HCC patients.

CK19

CK19 has been considered as a marker for the biliary phenotype^[193], and it is not expressed in normal hepatocytes^[194]. CK19 is currently recognized as a marker for LCSCs^[181,183,195,196]. Increased CK19 expression is correlated with high histological differentiation, advanced BCLC stage, TNM stage^[197], tumor non-encapsulation^[198], the presence of satellite lesions^[74], number of tumor foci, and vascular tumor embolism^[199]. These findings dictate that increased CK19 expression could serve as a new biomarker predicting HCC progression and recurrence. In addition, some studies have identified association between CK19 expression in HCC and increased vascular invasion, lymph node metastasis, and intrahepatic spread^[200,201], dictating that CK19 expression is an independent risk factor for developing LNM, and that it is an important risk factor for early tumor recurrence. In addition, increased CK19 expression has also been found to be both an independent poor prognostic factor for OS, DFS, and RFS in post-HR HCC patients^[74,197], and an independent prognostic factor for HCC with LNM^[202]. However, other studies have come to a different conclusion. Some studies have demonstrated that CK19 is an independent prognosticator for OS, but not for DFS^[194,199]. Still some studies have suggested that CK19 expression has prognostic significance for DFS, though CK19 fails to offer independent prognostic value^[188].

Taken together, the expressions of CD133, EpCAM and CK19 could be readily assessed by IHC and they are clinically significant biomarkers for surgically resected HCCs. However, predictive values of single LCSCs markers remain controversial and further validation is required in independent cohorts ahead of any clinical utilization^[203]. More importantly, because of high degree of HCC heterogeneity, the predictive range of a single marker is limited to a very small subpopulation. A combination of several LCSCs markers may provide greater specificity and reliability in predicting HCC prognosis^[178].

CELL SURFACE PROTEINS

Glypican-3

Glypican-3 (GPC3) is an oncofetal protein considered as a relatively specific HCC biomarker that is not detectable in hepatic para-carcinomatous and cirrhotic tissues^[204], and it is over-expressed in HCC using IHC^[205,206]. Recently, much evidence has shown that GPC3 could be a useful tool to identify early HCC development. More recently, GPC3 has been reported to be a new prognostic factor after curative hepatectomy in HCC patients.

In addition to being a marker for HCC, GPC3 plays a role in the progression of the disease^[207]. GPC3 expression has been less frequently observed in well-differentiated HCC than in moderately and poorly differentiated HCC^[205,208,209]; furthermore, it has been found significantly correlated with serum AFP level, tumor number and presence of satellite nodules, and TNM stage^[210,211]; in addition, GPC3 expression has also been found to be associated with postoperative metastasis/recurrence in HCC patients^[129,208,212]. These findings indicate that GPC3 expression might be a valuable marker closely related with post-operative progression, metastasis/recurrence in HCC patients. Multivariate analysis has identified GPC3 expression as an independent prognostic factor for OS^[129]. However, in other studies, for HCC patients with HCV infection in particular, the high membranous GPC3 immunoreactivity has been identified as an independent prognostic factor for DFS^[213]; one study has even suggested that over-expression of GPC3 is an independent prognostic factor for DFS in HBV-positive small HCC (< 5 cm)^[129]. Recently an extensive study has shown that high GPC3 expression is an independent risk factor for poor postoperative tumor recurrence, DFS, and OS^[211], again suggesting that GPC3 expression is a potential and reliable biomarker for predicting tumor recurrence and OS in post-HR HCC patients.

Overall, these studies indicate that GPC3 expression has the potential to serve as a valuable predictive marker for survival in post-HR HCC patients. Further studies are required to confirm GPC3 is one of the reliable clinical predictors of survival for individual post-HR HCC patient.

MAMMALIAN TARGET OF RAPAMYCIN PATHWAY

Currently there is evidence suggesting that phospho-specific antibodies could serve as potential biomarkers for HCC. These markers provide useful reagents for analysis of signaling pathways in clinical samples, and therefore has the potential for actionable targets^[214]. So far, the molecular biology of hepatocarcinogenesis and HCC progression has been widely investigated. Many studies have indicated that signaling pathways dysregulated in HCC are important steps towards understanding HCC pathogenesis and developing new therapeutic approaches. Over recent years, several molecular pathways have been identified contributing to the molecular pathogenesis of HCC, among which the mammalian target of rapamycin (mTOR) signaling pathway has been identified to play a critical role in the pathogenesis of HCC^[215]. And many studies have investigated the relationship between mTOR pathway and HCC prognosis.

mTOR pathway, an important regulator of multiple cellular functions including proliferation, differentiation, tumorigenesis, and apoptosis, is up-regulated in many cancers^[216]. Deregulation of the mTOR signaling pathway has been reported in many malignancies, and some of the

signaling molecules in this pathway could predict prognosis in different cancers. PI3K/AKT is considered a critical upstream mediator of the mTOR signaling pathway. Recent data from a genomic sequence of HCC samples identified mutations in PIK3CA, an upstream regulator of AKT, in 50% of patients with poor prognosis and survival length of < 3 years following partial liver resection, whereas only 10% of the HCC patients with a good prognosis had a mutation in PIK3CA^[217]. Activation of AKT is a risk factor for early disease recurrence and poor prognosis in patients with HCC^[218]. Activated AKT positively modulates mTOR function. mTOR is a key component of PI3K and AKT pathways that activate downstream kinases required for G1 to S phase transition^[219]. mTOR acts by directly activating p70S6 kinase (p70S6K/S6K1) and inhibiting 4E binding protein 1 (4E-BP1)^[220], both regulating the translation of important factors involved in cell proliferation (such as c-myc, cyclic D1 and pRb) and angiogenesis (such as HIF1- α)^[221]. The p70S6 kinase and 4E-BP1 have shown to be independent predictors of prognosis in several types of solid tumors including liver^[216,222,223]. Therefore, the expression of mTOR pathway could be used as prognostic indicator in HCC.

In addition, one study has indicated that c-Jun N-Terminal Protein Kinase 1 (JNK1) activation contributes to poorer HCC prognosis, and there is similarity in gene expression patterns between the HCC with abnormal mTOR signaling and JNK1 HCC^[224], which further supports the assumption that HCCs with abnormal mTOR signaling are tumors of a highly aggressive nature and with poorer prognosis.

Recently, mTOR has emerged as an exciting target for cancer therapy including HCC. mTOR inhibitors have been tested successfully in clinical trials for their antineoplastic potency and good tolerability^[225]. A second generation of mTOR pathway inhibitors has been utilized in preclinical HCC models^[226] and the results suggest that mTOR inhibitors in HCC treatment could have a bright future.

Noticeably, although phospho-specific antibodies used in IHC are expected to detect phosphorylated proteins^[227-229], some preanalytic variables (such as fixation technique and duration) may critically affect the signal^[230], and in some cases these antibodies may also cross-react with nonphosphorylated proteins^[231]. Therefore, it is of ultimate importance to standardize preanalytic variables and to employ a control in determining whether the staining pattern is specific.

CONCLUSION

In this review, we give an overview of the literature published on immunohistochemical prognostic markers in HCC. Out of 17 markers that have been investigated by ten groups (summarized in Table 1), there are twelve markers (over-expression of Ki67, VEGF, MMP-2, MMP-9, CD44s, CD44v6, OPN, HMGB1, CD133,

EpCAM, CK19, GPC3 and mTOR pathway, and increased microvascular density of CD105) that have shown to be independent prognostic factors for survival in HCC patients. However, studies on some markers, such as p53, E-cadherin and p27, have all reported inconsistent results. Lack of standardized IHC has contributed to these discrepancies; other possible contributors include small sample sizes, pathological differences in samples, heterogeneous patient populations, various follow-up periods of the patients, and different racial and regional groups.

So far, numerous investigations have demonstrated many immunohistochemical markers could be potential prognostic/predictive indicators of HCC. However, their clinical utilization is severely hindered by the lack of standardized IHC methodology.

Although IHC is the most widely applied technique in pathology to determine the expression status of tumor-associated proteins and to study the clinical prognostic relevance of biomarkers, IHC results are subject to a variety of pre-analytical variables (*e.g.*, fixation method or the duration of fixation, methods of tissue processing), analytic variables (*e.g.*, antibodies, dilutions, antigen retrieval, time of incubation), and post-analytic variables, most importantly, subjectivity in determining scoring system for protein expression (cut-off values, *i.e.*, thresholds for positivity and interpretation criteria). Throughout IHC, each and every variable may greatly affect the accuracy and reliability of IHC results.

In view of the urgent demand from clinical practice, it is prerequisite to rigorously standardize IHC methodology, and this standardization should include all aspects of pre-analytical, analytic and post-analytic variables.

It sounds like a mission impossible to exercise full control over all pre-analytic variables, not to mention a complete standardization. Having said that, the collaboration among laboratories in Europe and the States has proven to be effective in tackling them. Analytic variables could to some degree be compensated for by using a large sample series. It is worthwhile to highlight that, because polyclonal antibodies have higher chances to cross-react with other antigens, it is important to further validate if the results presented in the study are specific by comparing staining patterns obtained with polyclonal antibodies with staining patterns generated by monoclonal antibodies. In addition, in order to improve reliability and interpretability of immunohistochemical markers, it has been advocated that standardized reporting criteria be used for biomarker studies^[232]. A wide-spread adoption of these recommendations will help overcome some of these methodological issues.

Nevertheless, subjectivity in applying a scoring system for protein expression is probably the biggest obstacle for the pathology laboratories. Therefore, we put strong emphasis on post-analytic variables, *i.e.*, cut-off values and interpretation criteria.

Prognostic significance of immunohistochemical marker fluctuates sharply with different cut-off values,

which in itself makes it difficult to determine a valid cut-off value for clinical use. ROC curve analysis could be used as an alternative method in the selection and validation of cut-off scores for determining the most clinically relevant threshold for immunohistochemical tumor positivity^[41]. Where contradictory results have been yielded from researches on established biomarkers, this tool should be adopted to re-evaluate protein expression. In addition, the authors of this review would tentatively recommend future investigations on novel tumor markers use ROC curve analysis.

No IHC scoring methods have been strictly agreed on. Researchers have been relying on percentage of positivity or intensity of positive staining, or a combination of these two, to estimate protein level. The intensity of positive staining in liver tissue sections could be easily affected by such pigments as iron deposition or brown granules in Kupffer cells, and therefore is not a valid indicator of specific immunostaining. Additionally, IHC is a technique that detects specific antigens present in the target cells by labeling them with antibodies against them which are tagged with enzymes to convert a soluble colorless substrate to a colored insoluble precipitate which can be detected under the microscope. The intensity of positive staining is easily affected by individual researcher's skill and experience both in operating IHC and in reading slides, as well as technical conditions for IHC operation. Therefore, IHC intensity is not an appropriate criterion to be used in HCC research. The authors of this review would tentatively recommend that, for protein positive expression evaluation in liver tissue sections, percentage of stained area/field be selected as a quantitative method for IHC results. To ensure objectivity, the scoring methods of immunohistochemical markers should be assessed by independent observers.

It is worthwhile to highlight that IHC in itself could never tell us about the mutation status of these proteins. That is to say, in order to better understand the relevance between immunohistochemical markers and clinical outcomes, standardized IHC should be combined with gene mutation analysis using polymerase chain reaction methods in the same patients.

A number of studies have demonstrated that although single marker could provide useful information on the prediction of patients' survival and treatment outcomes, and could monitor efficacy of individualization of therapy, the heterogeneity of HCC tumors requires a combination of biomarkers in order to yield better clinical performance. In the foreseeable future it is likely that multiple markers need to be integrated into a prognostic signature to accurately predict outcomes. In fact, the HCC biomarkers in combination are increasingly becoming part of surveillance protocols in United States clinics^[235]. Still a further long way to go before their routine use in clinical practice becomes a reality, which requires immunohistochemical markers of prognosis and prediction to be validated in carefully designed large-scale, prospective clinical trials, using standardized IHC

techniques. Then, and only until then, could the validation of prognostic and predictive markers eventually guide our clinical decision making in regard to follow-up scheduling and treatment choice.

REFERENCES

- 1 **Iliescu L**, Mindrut E, Grasu M, Orban C, Tanase A, Toma L. Management of hepatocellular carcinoma -- experience of a single center. *Chirurgia* (Bucur) 2014; **109**: 204-207 [PMID: 24742411]
- 2 **Ishikawa T**. Anti-viral therapy to reduce recurrence and improve survival in hepatitis B virus-related hepatocellular carcinoma. *World J Gastroenterol* 2013; **19**: 8861-8866 [PMID: 24379608 DOI: 10.3748/wjg.v19.i47.8861]
- 3 **Cucchetti A**, Piscaglia F, Cescon M, Ercolani G, Pinna AD. Systematic review of surgical resection vs radiofrequency ablation for hepatocellular carcinoma. *World J Gastroenterol* 2013; **19**: 4106-4118 [PMID: 23864773 DOI: 10.3748/wjg.v19.i26.4106]
- 4 **Furtado R**, Crawford M, Sandroussi C. Systematic review and meta-analysis of adjuvant i(131) lipiodol after excision of hepatocellular carcinoma. *Ann Surg Oncol* 2014; **21**: 2700-2707 [PMID: 24743904 DOI: 10.1245/s10434-014-3511-2]
- 5 **Ishikawa T**. Strategy for improving survival and reducing recurrence of HCV-related hepatocellular carcinoma. *World J Gastroenterol* 2013; **19**: 6127-6130 [PMID: 24115808 DOI: 10.3748/wjg.v19.i37.6127]
- 6 **Yang Z**, Zhang Y, Wang L. A feedback inhibition between miRNA-127 and TGF β /c-Jun cascade in HCC cell migration via MMP13. *PLoS One* 2013; **8**: e65256 [PMID: 23762330 DOI: 10.1371/journal.pone.0065256]
- 7 **Matsumoto A**, Ishibashi Y, Urashima M, Omura N, Nakada K, Nishikawa K, Shida A, Takada K, Kashiwagi H, Yanaga K. High UBCH10 protein expression as a marker of poor prognosis in esophageal squamous cell carcinoma. *Anticancer Res* 2014; **34**: 955-961 [PMID: 24511039]
- 8 **Nosrati M**, Kashani-Sabet M. Immunohistochemical diagnostic and prognostic markers for melanoma. *Methods Mol Biol* 2014; **1102**: 259-273 [PMID: 24258983 DOI: 10.1007/978-1-62703-727-3_14]
- 9 **Zeestraten EC**, Benard A, Reimers MS, Schouten PC, Liefers GJ, van de Velde CJ, Kuppen PJ. The prognostic value of the apoptosis pathway in colorectal cancer: a review of the literature on biomarkers identified by immunohistochemistry. *Biomark Cancer* 2013; **5**: 13-29 [PMID: 24179395 DOI: 10.4137/BIC.S11475]
- 10 **Wang Z**, Jiang Y, Guan D, Li J, Yin H, Pan Y, Xie D, Chen Y. Critical roles of p53 in epithelial-mesenchymal transition and metastasis of hepatocellular carcinoma cells. *PLoS One* 2013; **8**: e72846 [PMID: 24023784 DOI: 10.1371/journal.pone.0072846]
- 11 **Tseng PL**, Tai MH, Huang CC, Wang CC, Lin JW, Hung CH, Chen CH, Wang JH, Lu SN, Lee CM, Changchien CS, Hu TH. Overexpression of VEGF is associated with positive p53 immunostaining in hepatocellular carcinoma (HCC) and adverse outcome of HCC patients. *J Surg Oncol* 2008; **98**: 349-357 [PMID: 18646041 DOI: 10.1002/jso.21109]
- 12 **Tu K**, Zheng X, Zan X, Han S, Yao Y, Liu Q. Evaluation of Fbxw7 expression and its correlation with the expression of c-Myc, cyclin E and p53 in human hepatocellular carcinoma. *Hepatol Res* 2012; **42**: 904-910 [PMID: 22548670 DOI: 10.1111/j.1872-034X.2012.01005.x]
- 13 **Hu TH**, Wang CC, Huang CC, Chen CL, Hung CH, Chen CH, Wang JH, Lu SN, Lee CM, Changchien CS, Tai MH. Down-regulation of tumor suppressor gene PTEN, overexpression of p53, plus high proliferating cell nuclear

- antigen index predict poor patient outcome of hepatocellular carcinoma after resection. *Oncol Rep* 2007; **18**: 1417-1426 [PMID: 17982625]
- 14 **Kang GH**, Lee BS, Lee ES, Kim SH, Lee HY, Kang DY. Prognostic significance of p53, mTOR, c-Met, IGF-1R, and HSP70 overexpression after the resection of hepatocellular carcinoma. *Gut Liver* 2014; **8**: 79-87 [PMID: 24516705 DOI: 10.5009/gnl.2014.8.1.79]
 - 15 **Srivastava S**, Wong KF, Ong CW, Huak CY, Yeoh KG, Teh M, Luk JM, Salto-Tellez M. A morpho-molecular prognostic model for hepatocellular carcinoma. *Br J Cancer* 2012; **107**: 334-339 [PMID: 22713659 DOI: 10.1038/bjc.2012.230]
 - 16 **Stroescu C**, Dragnea A, Ivanov B, Pechianu C, Herlea V, Sgarbura O, Popescu A, Popescu I. Expression of p53, Bcl-2, VEGF, Ki67 and PCNA and prognostic significance in hepatocellular carcinoma. *J Gastrointest Liver Dis* 2008; **17**: 411-417 [PMID: 19104702]
 - 17 **Sung CO**, Yoo BC, Koh KC, Cho JW, Park CK. Prognostic significance of p53 overexpression after hepatic resection of hepatocellular carcinoma. *Korean J Gastroenterol* 2005; **45**: 425-430 [PMID: 15973077]
 - 18 **Schöniger-Hekele M**, Hänel S, Wrba F, Müller C. Hepatocellular carcinoma--survival and clinical characteristics in relation to various histologic molecular markers in Western patients. *Liver Int* 2005; **25**: 62-69 [PMID: 15698400 DOI: 10.1111/j.1478-3231.2004.0997.x]
 - 19 **Qin HX**, Nan KJ, Yang G, Jing Z, Ruan ZP, Li CL, Xu R, Guo H, Sui CG, Wei YC. Expression and clinical significance of TAp73alpha, p53, PCNA and apoptosis in hepatocellular carcinoma. *World J Gastroenterol* 2005; **11**: 2709-2713 [PMID: 15884108]
 - 20 **Guo RP**, Zhong C, Shi M, Zhang CQ, Wei W, Zhang YQ, Li JQ. Clinical value of apoptosis and angiogenesis factors in estimating the prognosis of hepatocellular carcinoma. *J Cancer Res Clin Oncol* 2006; **132**: 547-555 [PMID: 16763805 DOI: 10.1007/s00432-006-0097-5]
 - 21 **Umamura A**, Itoh Y, Itoh K, Yamaguchi K, Nakajima T, Higashitsuji H, Onoue H, Fukumoto M, Okanoue T, Fujita J. Association of gankyrin protein expression with early clinical stages and insulin-like growth factor-binding protein 5 expression in human hepatocellular carcinoma. *Hepatology* 2008; **47**: 493-502 [PMID: 18161051 DOI: 10.1002/hep.22027]
 - 22 **Eguchi S**, Kanematsu T, Arii S, Omata M, Kudo M, Sakamoto M, Takayasu K, Makuuchi M, Matsuyama Y, Monden M. Recurrence-free survival more than 10 years after liver resection for hepatocellular carcinoma. *Br J Surg* 2011; **98**: 552-557 [PMID: 21267990 DOI: 10.1002/bjs.7393]
 - 23 **Liao KF**, Lai SW, Lin CY, Huang CH, Lin YY. Risk factors of recurrence after curative resection of hepatocellular carcinoma in Taiwan. *Am J Med Sci* 2011; **341**: 301-304 [PMID: 21441859 DOI: 10.1097/MAJ.0b013e3181ff5d93]
 - 24 **Yang S**, Yan HL, Tao QF, Yuan SX, Tang GN, Yang Y, Wang LL, Zhang YL, Sun SH, Zhou WP. Low CADM2 expression predicts high recurrence risk of hepatocellular carcinoma patients after hepatectomy. *J Cancer Res Clin Oncol* 2014; **140**: 109-116 [PMID: 24240726 DOI: 10.1007/s00432-013-1536-8]
 - 25 **Hu T**, Guo H, Wang W, Yu S, Han L, Jiang L, Ma J, Yang C, Guo Q, Nan K. Loss of p57 expression and RhoA overexpression are associated with poor survival of patients with hepatocellular carcinoma. *Oncol Rep* 2013; **30**: 1707-1714 [PMID: 23842948 DOI: 10.3892/or.2013.2608]
 - 26 **Park SK**, Jung YK, Chung DH, Kim KK, Park YH, Lee JN, Kwon OS, Kim YS, Choi DJ, Kim JH. Factors influencing hepatocellular carcinoma prognosis after hepatectomy: a single-center experience. *Korean J Intern Med* 2013; **28**: 428-438 [PMID: 23864801 DOI: 10.3904/kjim.2013.28.4.428]
 - 27 **Liu J**, Ma Q, Zhang M, Wang X, Zhang D, Li W, Wang F, Wu E. Alterations of TP53 are associated with a poor outcome for patients with hepatocellular carcinoma: evidence from a systematic review and meta-analysis. *Eur J Cancer* 2012; **48**: 2328-2338 [PMID: 22459764 DOI: 10.1016/j.ejca.2012.03.001]
 - 28 **Qi LN**, Bai T, Chen ZS, Wu FX, Chen YY, De Xiang B, Peng T, Han ZG, Li LQ. The p53 mutation spectrum in hepatocellular carcinoma from Guangxi, China : role of chronic hepatitis B virus infection and aflatoxin B1 exposure. *Liver Int* 2014; Epub ahead of print [PMID: 24461059 DOI: 10.1111/liv.12460]
 - 29 **Chen Ban K**, Singh H, Krishnan R, Fong Seow H. Comparison of the expression of beta-catenin in hepatocellular carcinoma in areas with high and low levels of exposure to aflatoxin B1. *J Surg Oncol* 2004; **86**: 157-163 [PMID: 15170655 DOI: 10.1002/jso.20051]
 - 30 **Song TJ**, Fong Y, Cho SJ, Gönen M, Hezel M, Tuorto S, Choi SY, Kim YC, Suh SO, Koo BH, Chae YS, Jarnagin WR, Klimstra DS. Comparison of hepatocellular carcinoma in American and Asian patients by tissue array analysis. *J Surg Oncol* 2012; **106**: 84-88 [PMID: 22234941 DOI: 10.1002/jso.23036]
 - 31 **Jiang YH**, Cheng B, Ge MH, Zhang G. The prognostic significance of p63 and Ki-67 expression in myoepithelial carcinoma. *Head Neck Oncol* 2012; **4**: 9 [PMID: 22452794 DOI: 10.1186/1758-3284-4-9]
 - 32 **Le Page C**, Huntsman DG, Provencher DM, Mes-Masson AM. Predictive and prognostic protein biomarkers in epithelial ovarian cancer: recommendation for future studies. *Cancers (Basel)* 2010; **2**: 913-954 [PMID: 24281100 DOI: 10.3390/cancers2020913]
 - 33 **Bologna-Molina R**, Mosqueda-Taylor A, Molina-Frechero N, Mori-Estevéz AD, Sánchez-Acuña G. Comparison of the value of PCNA and Ki-67 as markers of cell proliferation in ameloblastic tumors. *Med Oral Patol Oral Cir Bucal* 2013; **18**: e174-e179 [PMID: 23229269 DOI: 10.4317/medoral.18573]
 - 34 **Hsu HT**, Wu PR, Chen CJ, Hsu LS, Yeh CM, Hsing MT, Chiang YS, Lai MT, Yeh KT. High cytoplasmic expression of Krüppel-like factor 4 is an independent prognostic factor of better survival in hepatocellular carcinoma. *Int J Mol Sci* 2014; **15**: 9894-9906 [PMID: 24897024 DOI: 10.3390/ijms15069894]
 - 35 **Huang X**, Liu F, Zhu C, Cai J, Wang H, Wang X, He S, Liu C, Yao L, Ding Z, Zhang Y, Zhang T. Suppression of KIF3B expression inhibits human hepatocellular carcinoma proliferation. *Dig Dis Sci* 2014; **59**: 795-806 [PMID: 24368420 DOI: 10.1007/s10620-013-2969-2]
 - 36 **Ito Y**, Matsuura N, Sakon M, Takeda T, Umeshita K, Nagano H, Nakamori S, Dono K, Tsujimoto M, Nakahara M, Nakao K, Monden M. Both cell proliferation and apoptosis significantly predict shortened disease-free survival in hepatocellular carcinoma. *Br J Cancer* 1999; **81**: 747-751 [PMID: 10574266 DOI: 10.1038/sj.bjc.6690758]
 - 37 **Chen H**, Miao J, Li H, Wang C, Li J, Zhu Y, Wang J, Wu X, Qiao H. Expression and prognostic significance of p21-activated kinase 6 in hepatocellular carcinoma. *J Surg Res* 2014; **189**: 81-88 [PMID: 24576777 DOI: 10.1016/j.jss.2014.01.049]
 - 38 **Cao X**, Xia Y, Yang J, Jiang J, Chen L, Ni R, Li L, Gu Z. Clinical and biological significance of never in mitosis gene A-related kinase 6 (NEK6) expression in hepatic cell cancer. *Pathol Oncol Res* 2012; **18**: 201-207 [PMID: 21725899 DOI: 10.1007/s12253-011-9429-0]
 - 39 **Ke Q**, Ji J, Cheng C, Zhang Y, Lu M, Wang Y, Zhang L, Li P, Cui X, Chen L, He S, Shen A. Expression and prognostic role of Spy1 as a novel cell cycle protein in hepatocellular carcinoma. *Exp Mol Pathol* 2009; **87**: 167-172 [PMID: 19686732 DOI: 10.1016/j.yexmp.2009.07.011]
 - 40 **Schmilovitz-Weiss H**, Tobar A, Halpern M, Levy I, Shabtai E, Ben-Ari Z. Tissue expression of squamous cellular carcinoma antigen and Ki67 in hepatocellular carcinoma-correlation with prognosis: a historical prospective study. *Diagn Pathol* 2011; **6**: 121 [PMID: 22151825 DOI: 10.1186/1746-1596-6-121]
 - 41 **Zlobec I**, Steele R, Terracciano L, Jass JR, Lugli A. Selecting immunohistochemical cut-off scores for novel biomarkers

- of progression and survival in colorectal cancer. *J Clin Pathol* 2007; **60**: 1112-1116 [PMID: 17182662 DOI: 10.1136/jcp.2006.044537]
- 42 **Yang H**, Liu J, Yu H, Sun P, Hu Y, Zhong J, Zhu Z. Expression and association of CD44v6 with prognosis in T2-3N0M0 esophageal squamous cell carcinoma. *J Thorac Dis* 2014; **6**: 91-98 [PMID: 24605222 DOI: 10.3978/j.issn.2072-1439.2013.11.16]
- 43 **Spira A**, Ettinger DS. Multidisciplinary management of lung cancer. *N Engl J Med* 2004; **350**: 379-392 [PMID: 14736930 DOI: 10.1056/NEJMra035536]
- 44 **Wang D**, Stockard CR, Harkins L, Lott P, Salih C, Yuan K, Buchsbaum D, Hashim A, Zayzafoon M, Hardy RW, Hameed O, Grizzle W, Siegal GP. Immunohistochemistry in the evaluation of neovascularization in tumor xenografts. *Biotech Histochem* 2008; **83**: 179-189 [PMID: 18846440 DOI: 10.1080/10520290802451085]
- 45 **Wang SN**, Chuang SC, Yeh YT, Yang SF, Chai CY, Chen WT, Kuo KK, Chen JS, Lee KT. Potential prognostic value of leptin receptor in hepatocellular carcinoma. *J Clin Pathol* 2006; **59**: 1267-1271 [PMID: 16565226 DOI: 10.1136/jcp.2005.033464]
- 46 **Chebibi I**, Shabani-Rad MT, Chow MS, Zhang J, Gao ZH. Microvessel density and clinicopathologic characteristics in hepatocellular carcinoma with and without cirrhosis. *Biomark Insights* 2007; **2**: 59-68 [PMID: 19662192]
- 47 **Yang P**, Yuan W, He J, Wang J, Yu L, Jin X, Hu Y, Liao M, Chen Z, Zhang Y. Overexpression of EphA2, MMP-9, and MVD-CD34 in hepatocellular carcinoma: Implications for tumor progression and prognosis. *Hepatol Res* 2009; **39**: 1169-1177 [PMID: 19788698 DOI: 10.1111/j.1872-034X.2009.00563.x]
- 48 **Nanashima A**, Nakayama T, Sumida Y, Abo T, Takeshita H, Shibata K, Hidaka S, Sawai T, Yasutake T, Nagayasu T. Relationship between microvessel count and post-hepatectomy survival in patients with hepatocellular carcinoma. *World J Gastroenterol* 2008; **14**: 4915-4922 [PMID: 18756600 DOI: 10.3748/wjg.14.4915]
- 49 **Zeng W**, Gouw AS, van den Heuvel MC, Molema G, Poppema S, van der Jagt EJ, de Jong KP. Hepatocellular carcinomas in cirrhotic and noncirrhotic human livers share angiogenic characteristics. *Ann Surg Oncol* 2010; **17**: 1564-1571 [PMID: 20087783 DOI: 10.1245/s10434-009-0900-z]
- 50 **Yang LY**, Lu WQ, Huang GW, Wang W. Correlation between CD105 expression and postoperative recurrence and metastasis of hepatocellular carcinoma. *BMC Cancer* 2006; **6**: 110 [PMID: 16650286 DOI: 10.1186/1471-2407-6-110]
- 51 **Miyata Y**, Sagara Y, Watanabe S, Asai A, Matsuo T, Ohba K, Hayashi T, Sakai H. CD105 is a more appropriate marker for evaluating angiogenesis in urothelial cancer of the upper urinary tract than CD31 or CD34. *Virchows Arch* 2013; **463**: 673-679 [PMID: 23975255 DOI: 10.1007/s00428-013-1463-8]
- 52 **Saroufim A**, Messai Y, Hasmim M, Rioux N, Iacovelli R, Verhoest G, Bensalah K, Patard JJ, Albiges L, Azzarone B, Escudier B, Chouaib S. Tumoral CD105 is a novel independent prognostic marker for prognosis in clear-cell renal cell carcinoma. *Br J Cancer* 2014; **110**: 1778-1784 [PMID: 24594997 DOI: 10.1038/bjc.2014.71]
- 53 **Gurzu S**, Cimpean AM, Kovacs J, Jung I. Counting of angiogenesis in colorectal carcinomas using double immunostain. *Tumori* 2012; **98**: 485-490 [PMID: 23052166 DOI: 10.1700/1146.12644]
- 54 **Pappa CA**, Alexandrakis MG, Boula A, Psarakis FE, Kolovou A, Bantouna V, Stavroulaki E, Tsirakis G. Emerging roles of endoglin/CD105 and angiogenic cytokines for disease development and progression in multiple myeloma patients. *Hematol Oncol* 2013; **31**: 201-205 [PMID: 23576184 DOI: 10.1002/hon.2044]
- 55 **Bodnar M**, Szyłberg Ł, Kaźmierczak W, Marszałek A. [Evaluation of microvessel density (MVD) in laryngeal squamous cell carcinoma]. *Przegl Lek* 2012; **69**: 726-730 [PMID: 23421020]
- 56 **Wang Y**, Zhang XH, Guo P, Yan LN, He D. [Tumor microvascular density detected by anti-CD105 and anti-CD34 in hepatocellular carcinoma patients and its predictive value of tumor recurrence after liver transplantation]. *Sichuan Daxue Xuebao Yixueban* 2010; **41**: 818-821 [PMID: 21302449]
- 57 **Yao Y**, Pan Y, Chen J, Sun X, Qiu Y, Ding Y. Endoglin (CD105) expression in angiogenesis of primary hepatocellular carcinomas: analysis using tissue microarrays and comparisons with CD34 and VEGF. *Ann Clin Lab Sci* 2007; **37**: 39-48 [PMID: 17311868]
- 58 **Ho JW**, Poon RT, Sun CK, Xue WC, Fan ST. Clinicopathological and prognostic implications of endoglin (CD105) expression in hepatocellular carcinoma and its adjacent non-tumorous liver. *World J Gastroenterol* 2005; **11**: 176-181 [PMID: 15633211 DOI: 10.3748/wjg.v11.i2.176]
- 59 **Weidner N**. Current pathologic methods for measuring intratumoral microvessel density within breast carcinoma and other solid tumors. *Breast Cancer Res Treat* 1995; **36**: 169-180 [PMID: 8534865 DOI: 10.1007/BF00666038]
- 60 **Sobczyńska-Rak A**, Polkowska I, Silmanowicz P. Elevated Vascular Endothelial Growth Factor (VEGF) levels in the blood serum of dogs with malignant neoplasms of the oral cavity. *Acta Vet Hung* 2014; **62**: 362-371 [PMID: 24659713 DOI: 10.1556/AVet.2014.009]
- 61 **Kaseb AO**, Hanbali A, Cotant M, Hassan MM, Wollner I, Philip PA. Vascular endothelial growth factor in the management of hepatocellular carcinoma: a review of literature. *Cancer* 2009; **115**: 4895-4906 [PMID: 19637355 DOI: 10.1002/cncr.24537]
- 62 **Kong SY**, Park JW, Lee JA, Park JE, Park KW, Hong EK, Kim CM. Association between vascular endothelial growth factor gene polymorphisms and survival in hepatocellular carcinoma patients. *Hepatology* 2007; **46**: 446-455 [PMID: 17659575 DOI: 10.1002/hep.21720]
- 63 **Wang D**, Luo L, Chen W, Chen LZ, Zeng WT, Li W, Huang XH. Significance of the vascular endothelial growth factor and the macrophage migration inhibitory factor in the progression of hepatocellular carcinoma. *Oncol Rep* 2014; **31**: 1199-1204 [PMID: 24366206 DOI: 10.3892/or.2013.2946]
- 64 **Thelen A**, Scholz A, Benckert C, von Marschall Z, Schröder M, Wiedenmann B, Neuhaus P, Rosewicz S, Jonas S. VEGF-D promotes tumor growth and lymphatic spread in a mouse model of hepatocellular carcinoma. *Int J Cancer* 2008; **122**: 2471-2481 [PMID: 18338756 DOI: 10.1002/ijc.23439]
- 65 **Minata M**, Harada KH, Kudo M, Ikai I, Nishida N. The prognostic value of vascular endothelial growth factor in hepatocellular carcinoma for predicting metastasis after curative resection. *Oncology* 2013; **84** Suppl 1: 75-81 [PMID: 23428863 DOI: 10.1159/000345894]
- 66 **Xiang ZL**, Zeng ZC, Fan J, Tang ZY, Zeng HY, Gao DM. Gene expression profiling of fixed tissues identified hypoxia-inducible factor-1 α , VEGF, and matrix metalloproteinase-2 as biomarkers of lymph node metastasis in hepatocellular carcinoma. *Clin Cancer Res* 2011; **17**: 5463-5472 [PMID: 21712445]
- 67 **Chen L**, Shi Y, Jiang CY, Wei LX, Lv YL, Wang YL, Dai GH. Coexpression of PDGFR- α , PDGFR- β and VEGF as a prognostic factor in patients with hepatocellular carcinoma. *Int J Biol Markers* 2011; **26**: 108-116 [PMID: 21574155 DOI: 10.5301/IJBM.2011.8322]
- 68 **Hu J**, Xu Y, Shen ZZ, Wang Z, Lu Q, Yang GH, Ding ZB, Fan J, Zhou J. High expressions of vascular endothelial growth factor and platelet-derived endothelial cell growth factor predict poor prognosis in alpha-fetoprotein-negative hepatocellular carcinoma patients after curative resection. *J Cancer Res Clin Oncol* 2009; **135**: 1359-1367 [PMID: 19350273 DOI: 10.1007/s00432-009-0577-5]
- 69 **Herszényi L**, Hritz I, Lakatos G, Varga MZ, Tulassay Z. The behavior of matrix metalloproteinases and their inhibitors in

- colorectal cancer. *Int J Mol Sci* 2012; **13**: 13240-13263 [PMID: 23202950 DOI: 10.3390/ijms131013240]
- 70 **Fan HX**, Li HX, Chen D, Gao ZX, Zheng JH. Changes in the expression of MMP2, MMP9, and ColIV in stromal cells in oral squamous tongue cell carcinoma: relationships and prognostic implications. *J Exp Clin Cancer Res* 2012; **31**: 90 [PMID: 23107277 DOI: 10.1186/1756-9966-31-90]
- 71 **Dodd T**, Jadhav R, Wiggins L, Stewart J, Smith E, Russell JC, Rocic P. MMPs 2 and 9 are essential for coronary collateral growth and are prominently regulated by p38 MAPK. *J Mol Cell Cardiol* 2011; **51**: 1015-1025 [PMID: 21884701 DOI: 10.1016/j.yjmcc.2011.08.012]
- 72 **Puzovic V**, Brcic I, Ranogajec I, Jakic-Razumovic J. Prognostic values of ETS-1, MMP-2 and MMP-9 expression and co-expression in breast cancer patients. *Neoplasma* 2014; **61**: 439-446 [PMID: 24645837 DOI: 10.4149/neo_2014_054]
- 73 **Puljiz M**, Puljiz Z, Vucemilo T, Ramić S, Knezević F, Culo B, Alvir I, Tomica D, Danolić D. Prognostic significance of matrix metalloproteinases 2 and 9 in endometrial cancer. *Coll Antropol* 2012; **36**: 1367-1372 [PMID: 23390835]
- 74 **Xiang ZL**, Zeng ZC, Tang ZY, Fan J, Sun HC, Tan YS. Expression of cytokeratin 19 and matrix metalloproteinase 2 predicts lymph node metastasis in hepatocellular carcinoma. *Mol Biol Rep* 2011; **38**: 3531-3539 [PMID: 21104440 DOI: 10.1007/s11033-010-0463-x]
- 75 **Guo RP**, Zhong C, Shi M, Zhang CQ, Wei W, Zhang YQ, Li JQ. [Expression and clinical impact of vascular endothelial growth factor and matrix metalloproteinase-2 in hepatocellular carcinoma]. *Zhonghua Zhongliu Zazhi* 2006; **28**: 285-288 [PMID: 16875630]
- 76 **Nart D**, Yaman B, Yilmaz F, Zeytinlu M, Karasu Z, Kiliç M. Expression of matrix metalloproteinase-9 in predicting prognosis of hepatocellular carcinoma after liver transplantation. *Liver Transpl* 2010; **16**: 621-630 [PMID: 20440771 DOI: 10.1002/lt.22028]
- 77 **Hou YK**, Wang Y, Cong WM, Wu MC. [Expression of tumor metastasis-suppressor gene KiSS-1 and matrix metalloproteinase-9 in portal vein tumor thrombus of hepatocellular carcinoma]. *Ai Zheng* 2007; **26**: 591-595 [PMID: 17562263]
- 78 **Lee CF**, Ling ZQ, Zhao T, Lee KR. Distinct expression patterns in hepatitis B virus- and hepatitis C virus-infected hepatocellular carcinoma. *World J Gastroenterol* 2008; **14**: 6072-6077 [PMID: 18932288 DOI: 10.3748/wjg.14.6072]
- 79 **Chen ZB**, Shen SQ, Ding YM, Wang WX, Tao JP, Liang LJ, Hu WJ. The angiogenic and prognostic implications of VEGF, Ang-1, Ang-2, and MMP-9 for hepatocellular carcinoma with background of hepatitis B virus. *Med Oncol* 2009; **26**: 365-371 [PMID: 19082771 DOI: 10.1007/s12032-008-9130-7]
- 80 **Ishii Y**, Nakasato Y, Kobayashi S, Yamazaki Y, Aoki T. A study on angiogenesis-related matrix metalloproteinase networks in primary hepatocellular carcinoma. *J Exp Clin Cancer Res* 2003; **22**: 461-470 [PMID: 14582707]
- 81 **Chen R**, Cui J, Xu C, Xue T, Guo K, Gao D, Liu Y, Ye S, Ren Z. The significance of MMP-9 over MMP-2 in HCC invasiveness and recurrence of hepatocellular carcinoma after curative resection. *Ann Surg Oncol* 2012; **19** Suppl 3: S375-S384 [PMID: 21681378 DOI: 10.1245/s10434-011-1836-7]
- 82 **Altadill A**, Rodríguez M, González LO, Junquera S, Corte MD, González-Dieguez ML, Linares A, Barbón E, Fresno-Forcelledo M, Rodrigo L, Vizoso FJ. Liver expression of matrix metalloproteases and their inhibitors in hepatocellular carcinoma. *Dig Liver Dis* 2009; **41**: 740-748 [PMID: 19372066 DOI: 10.1016/j.dld.2009.01.016]
- 83 **Gao ZH**, Tretiakova MS, Liu WH, Gong C, Farris PD, Hart J. Association of E-cadherin, matrix metalloproteinases, and tissue inhibitors of metalloproteinases with the progression and metastasis of hepatocellular carcinoma. *Mod Pathol* 2006; **19**: 533-540 [PMID: 16474379 DOI: 10.1038/modpathol.3800554]
- 84 **Matsunaga Y**, Koda M, Murawaki Y. Expression of matrix metalloproteinases (MMPs) and tissue inhibitors of metalloproteinases (TIMPs) in hepatocellular carcinoma tissue, compared with the surrounding non-tumor tissue. *Res Commun Mol Pathol Pharmacol* 2004; **115-116**: 143-150 [PMID: 17564313]
- 85 **Wei QY**, Wu YQ, Fan SQ. [Expression of matrix metalloproteinases and tissue inhibitors of matrix metalloproteinases in the hepatocellular carcinomas]. *Hunan Yike Daxue Xuebao* 2003; **28**: 212-216 [PMID: 14653069]
- 86 **Bendas G**, Borsig L. Cancer cell adhesion and metastasis: selectins, integrins, and the inhibitory potential of heparins. *Int J Cell Biol* 2012; **2012**: 676731 [PMID: 22505933 DOI: 10.1155/2012/676731]
- 87 **Techasen A**, Loilome W, Namwat N, Khuntikeo N, Puapairoj A, Jearanaikoon P, Saya H, Yongvanit P. Loss of E-cadherin promotes migration and invasion of cholangiocarcinoma cells and serves as a potential marker of metastasis. *Tumour Biol* 2014; **35**: 8645-8652 [PMID: 24867095 DOI: 10.1007/s13277-014-2087-6]
- 88 **Pectasides E**, Rampias T, Sasaki C, Perisanidis C, Kouloulis V, Burtness B, Zamboukas T, Rimm D, Fountzilias G, Psyrri A. Markers of epithelial to mesenchymal transition in association with survival in head and neck squamous cell carcinoma (HNSCC). *PLoS One* 2014; **9**: e94273 [PMID: 24722213 DOI: 10.1371/journal.pone.0094273]
- 89 **Endo K**, Ueda T, Ueyama J, Ohta T, Terada T. Immunoreactive E-cadherin, alpha-catenin, beta-catenin, and gamma-catenin proteins in hepatocellular carcinoma: relationships with tumor grade, clinicopathologic parameters, and patients' survival. *Hum Pathol* 2000; **31**: 558-565 [PMID: 10836294 DOI: 10.1053/hp.2000.6683]
- 90 **Wei Y**, Van Nhieu JT, Prigent S, Srivatanakul P, Tiollais P, Buendia MA. Altered expression of E-cadherin in hepatocellular carcinoma: correlations with genetic alterations, beta-catenin expression, and clinical features. *Hepatology* 2002; **36**: 692-701 [PMID: 12198663 DOI: 10.1053/jhep.2002.35342]
- 91 **Guo C**, Liu QG, Yang W, Zhang ZL, Yao YM. Relation among p130Cas, E-cadherin and beta-catenin expression, clinicopathologic significance and prognosis in human hepatocellular carcinoma. *Hepatobiliary Pancreat Dis Int* 2008; **7**: 490-496 [PMID: 18842495]
- 92 **Hashiguchi M**, Ueno S, Sakoda M, Iino S, Hiwatashi K, Minami K, Ando K, Mataka Y, Maemura K, Shinchi H, Ishigami S, Natsugoe S. Clinical implication of ZEB-1 and E-cadherin expression in hepatocellular carcinoma (HCC). *BMC Cancer* 2013; **13**: 572 [PMID: 24304617 DOI: 10.1186/1471-2407-13-572]
- 93 **Minata M**, Kudo M, Harada KH, Ikai I, Nishida N. Expression of E-cadherin and vascular endothelial growth factor in noncancerous liver is associated with recurrence of hepatocellular carcinoma after curative resection. *Oncology* 2013; **84** Suppl 1: 88-92 [PMID: 23428865 DOI: 10.1159/000345896]
- 94 **Cho SB**, Lee KH, Lee JH, Park SY, Lee WS, Park CH, Kim HS, Choi SK, Rew JS. Expression of E- and N-cadherin and clinicopathology in hepatocellular carcinoma. *Pathol Int* 2008; **58**: 635-642 [PMID: 18801083 DOI: 10.1111/j.1440-1827.2008.02282.x]
- 95 **Woo HY**, Min AL, Choi JY, Bae SH, Yoon SK, Jung CK. Clinicopathologic significance of the expression of Snail in hepatocellular carcinoma. *Korean J Hepatol* 2011; **17**: 12-18 [PMID: 21494073 DOI: 10.3350/kjhep.2011.17.1.12]
- 96 **Korita PV**, Wakai T, Shirai Y, Matsuda Y, Sakata J, Cui X, Ajioka Y, Hatakeyama K. Overexpression of osteopontin independently correlates with vascular invasion and poor prognosis in patients with hepatocellular carcinoma. *Hum Pathol* 2008; **39**: 1777-1783 [PMID: 18701136 DOI: 10.1016/j.humpath.2008.05.006]
- 97 **Schneider MR**, Hiltwein F, Grill J, Blum H, Krebs S, Klanner A, Bauersachs S, Bruns C, Longerich T, Horst D, Brandl

- L, de Toni E, Herbst A, Kolligs FT. Evidence for a role of E-cadherin in suppressing liver carcinogenesis in mice and men. *Carcinogenesis* 2014; **35**: 1855-1862 [PMID: 24840851 DOI: 10.1093/carcin/bgu109]
- 98 **Jiang XM**, Zhang JB, Xiong J, Huang XX, Ren ZG. Altered distribution and expression pattern of E-cadherin in hepatocellular carcinomas: correlations with prognosis and clinical features. *Asian Pac J Cancer Prev* 2012; **13**: 6455-6461 [PMID: 23464474 DOI: 10.7314/APJCP.2012.13.12.6455]
- 99 **Kalluri R**, Neilson EG. Epithelial-mesenchymal transition and its implications for fibrosis. *J Clin Invest* 2003; **112**: 1776-1784 [PMID: 14679171 DOI: 10.1172/JCI200320530]
- 100 **Thiery JP**, Acloque H, Huang RY, Nieto MA. Epithelial-mesenchymal transitions in development and disease. *Cell* 2009; **139**: 871-890 [PMID: 19945376 DOI: 10.1016/j.cell.2009.11.007]
- 101 **Zucchini-Pascal N**, Peyre L, Rahmani R. Crosstalk between beta-catenin and snail in the induction of epithelial to mesenchymal transition in hepatocarcinoma: role of the ERK1/2 pathway. *Int J Mol Sci* 2013; **14**: 20768-20792 [PMID: 24135872 DOI: 10.3390/ijms141020768]
- 102 **Mima K**, Hayashi H, Kuroki H, Nakagawa S, Okabe H, Chikamoto A, Watanabe M, Beppu T, Baba H. Epithelial-mesenchymal transition expression profiles as a prognostic factor for disease-free survival in hepatocellular carcinoma: Clinical significance of transforming growth factor- β signaling. *Oncol Lett* 2013; **5**: 149-154 [PMID: 23255911]
- 103 **Xiao S**, Zhou Y, Jiang J, Yuan L, Xue M. CD44 affects the expression level of FOS-like antigen 1 in cervical cancer tissues. *Mol Med Rep* 2014; **9**: 1667-1674 [PMID: 24604526 DOI: 10.3892/mmr.2014.2010]
- 104 **Ko YH**, Won HS, Jeon EK, Hong SH, Roh SY, Hong YS, Byun JH, Jung CK, Kang JH. Prognostic significance of CD44s expression in resected non-small cell lung cancer. *BMC Cancer* 2011; **11**: 340 [PMID: 21819617 DOI: 10.1186/1471-2407-11-340]
- 105 **Okada T**, Nakamura T, Watanabe T, Onoda N, Ashida A, Okuyama R, Ito K. Coexpression of EpCAM, CD44 variant isoforms and claudin-7 in anaplastic thyroid carcinoma. *PLoS One* 2014; **9**: e94487 [PMID: 24727741 DOI: 10.1371/journal.pone.0094487]
- 106 **Ni J**, Cozzi PJ, Hao JL, Beretov J, Chang L, Duan W, Shigdar S, Delprado WJ, Graham PH, Bucci J, Kearsley JH, Li Y. CD44 variant 6 is associated with prostate cancer metastasis and chemo-/radioresistance. *Prostate* 2014; **74**: 602-617 [PMID: 24615685 DOI: 10.1002/pros.22775]
- 107 **Dan T**, Hewitt SM, Ohri N, Ly D, Soule BP, Smith SL, Matsuda K, Council C, Shankavaram U, Lippman ME, Mitchell JB, Camphausen K, Simone NL. CD44 is prognostic for overall survival in the NCI randomized trial on breast conservation with 25 year follow-up. *Breast Cancer Res Treat* 2014; **143**: 11-18 [PMID: 24276281 DOI: 10.1007/s10549-013-2758-9]
- 108 **Hu S**, Wu X, Zhou B, Xu Z, Qin J, Lu H, Lv L, Gao Y, Deng L, Yin J, Li G. IMP3 combined with CD44s, a novel predictor for prognosis of patients with hepatocellular carcinoma. *J Cancer Res Clin Oncol* 2014; **140**: 883-893 [PMID: 24647926 DOI: 10.1007/s00432-014-1639-x]
- 109 **Mima K**, Okabe H, Ishimoto T, Hayashi H, Nakagawa S, Kuroki H, Watanabe M, Beppu T, Tamada N, Nagano O, Saya H, Baba H. CD44s regulates the TGF- β -mediated mesenchymal phenotype and is associated with poor prognosis in patients with hepatocellular carcinoma. *Cancer Res* 2012; **72**: 3414-3423 [PMID: 22552294 DOI: 10.1158/0008-5472.CAN-12-0299]
- 110 **Beckebaum S**, Chen X, Sotiropoulos GC, Radtke A, Daoudaki M, Baba HA, Wohlschlaeger J, Broelsch CE, Gerken G, Cicinnati VR. Role of osteopontin and CD44s expression for patients with hepatocellular carcinoma undergoing liver transplantation or resection. *Transplant Proc* 2008; **40**: 3182-3184 [PMID: 19010227 DOI: 10.1016/j.transproceed.2008.08.034]
- 111 **Mathew J**, Hines JE, Obafunwa JO, Burr AW, Toole K, Burt AD. CD44 is expressed in hepatocellular carcinomas showing vascular invasion. *J Pathol* 1996; **179**: 74-79 [PMID: 8691349 DOI: 10.1002/(SICI)1096-9896(199605)179:1<74::AID-PATH531>3.0.CO;2-E]
- 112 **Ryu HS**, Park SH, Lee KB, Shin E, Jang JJ. Expression of the Transmembrane Glycoprotein CD44s Is Potentially an Independent Predictor of Recurrence in Hepatocellular Carcinoma. *Gut Liver* 2011; **5**: 204-209 [PMID: 21814602 DOI: 10.5009/gnl.2011.5.2.204]
- 113 **Endo K**, Terada T. Protein expression of CD44 (standard and variant isoforms) in hepatocellular carcinoma: relationships with tumor grade, clinicopathologic parameters, p53 expression, and patient survival. *J Hepatol* 2000; **32**: 78-84 [PMID: 10673070 DOI: 10.1016/S0168-8278(00)80192-0]
- 114 **Zhang BY**, Dai XW, Chen QY, Fang L, Qian B, Sun GY, Cui HH. [Expression of epithelial-cadherin, CD44v6 and connexin43 in hepatocellular carcinoma]. *Zhonghua Binglixue Zazhi* 2006; **35**: 616-619 [PMID: 17134571]
- 115 **Zhou ZJ**, Dai Z, Zhou SL, Fu XT, Zhao YM, Shi YH, Zhou J, Fan J. Overexpression of HnRNP A1 promotes tumor invasion through regulating CD44v6 and indicates poor prognosis for hepatocellular carcinoma. *Int J Cancer* 2013; **132**: 1080-1089 [PMID: 22821376 DOI: 10.1002/ijc.27742]
- 116 **Chen BL**, Guo K, Liu YK. [Relationship between CD44 expression or glycosylation and hepatocellular carcinoma metastasis]. *Zhonghua Ganzangbing Zazhi* 2011; **19**: 898-903 [PMID: 22525501 DOI: 10.3760/cma.j.issn.1007-3418.2011.12.005]
- 117 **Mima K**, Okabe H, Ishimoto T, Hayashi H, Nakagawa S, Kuroki H, Miyake K, Takamori H, Beppu T, Baba H. The expression levels of CD44v6 are correlated with the invasiveness of hepatocellular carcinoma in vitro, but do not appear to be clinically significant. *Oncol Lett* 2012; **3**: 1047-1051 [PMID: 22783389]
- 118 **Etiz D**, Ataizi FC, Bayman E, Akcay M, Acikalin MF, Colak E, Ciftci E. Prognostic value of osteopontin in patients treated with primary radiotherapy for head and neck cancer. *Asian Pac J Cancer Prev* 2013; **14**: 5175-5178 [PMID: 24175796 DOI: 10.7314/APJCP.2013.14.9.5175]
- 119 **Thorat D**, Sahu A, Behera R, Lohite K, Deshmukh S, Mane A, Karnik S, Doke S, Kundu GC. Association of osteopontin and cyclooxygenase-2 expression with breast cancer subtypes and their use as potential biomarkers. *Oncol Lett* 2013; **6**: 1559-1564 [PMID: 24260046]
- 120 **Gimba ER**, Tilli TM. Human osteopontin splicing isoforms: known roles, potential clinical applications and activated signaling pathways. *Cancer Lett* 2013; **331**: 11-17 [PMID: 23246372 DOI: 10.1016/j.canlet.2012.12.003]
- 121 **Kruger TE**, Miller AH, Godwin AK, Wang J. Bone sialoprotein and osteopontin in bone metastasis of osteotropic cancers. *Crit Rev Oncol Hematol* 2014; **89**: 330-341 [PMID: 24071501 DOI: 10.1016/j.critrevonc.2013.08.013]
- 122 **Lin F**, Li Y, Cao J, Fan S, Wen J, Zhu G, Du H, Liang Y. Overexpression of osteopontin in hepatocellular carcinoma and its relationships with metastasis, invasion of tumor cells. *Mol Biol Rep* 2011; **38**: 5205-5210 [PMID: 21188534 DOI: 10.1007/s11033-010-0671-4]
- 123 **Hua Z**, Chen J, Sun B, Zhao G, Zhang Y, Fong Y, Jia Z, Yao L. Specific expression of osteopontin and S100A6 in hepatocellular carcinoma. *Surgery* 2011; **149**: 783-791 [PMID: 21310450 DOI: 10.1016/j.surg.2010.12.007]
- 124 **Tsai WC**, Tsai WC, Lee HS, Jin JS, Gao HW, Chao TK, Chen A, Nieh S, Chan DC, Chang FN, Lin CK. Association between Osteopontin and EGFR Expression with Clinicopathological Parameters in Hepatocellular Carcinoma. *Chin J Physiol* 2012; **55**: 412-420 [PMID: 23286449 DOI: 10.4077/CJP.2012.BAA082]

- 125 **Qin L.** Osteopontin is a promoter for hepatocellular carcinoma metastasis: a summary of 10 years of studies. *Front Med* 2014; **8**: 24-32 [PMID: 24464486 DOI: 10.1007/s11684-014-0312-8]
- 126 **Jin Y,** Chen JN, Feng ZY, Zhang ZG, Fan WZ, Wang Y, Li JP. OPN and $\alpha v\beta 3$ expression are predictors of disease severity and worse prognosis in hepatocellular carcinoma. *PLoS One* 2014; **9**: e87930 [PMID: 24498405 DOI: 10.1371/journal.pone.0087930]
- 127 **Chen RX,** Xia YH, Cui JF, Xue TC, Ye SL. Osteopontin, a single marker for predicting the prognosis of patients with tumor-node-metastasis stage I hepatocellular carcinoma after surgical resection. *J Gastroenterol Hepatol* 2010; **25**: 1435-1442 [PMID: 20659235 DOI: 10.1111/j.1440-1746.2010.06277.x]
- 128 **Cao DX,** Li ZJ, Jiang XO, Lum YL, Khin E, Lee NP, Wu GH, Luk JM. Osteopontin as potential biomarker and therapeutic target in gastric and liver cancers. *World J Gastroenterol* 2012; **18**: 3923-3930 [PMID: 22912540 DOI: 10.3748/wjg.v18.i30.3923]
- 129 **Yu MC,** Lee YS, Lin SE, Wu HY, Chen TC, Lee WC, Chen MF, Tsai CN. Recurrence and poor prognosis following resection of small hepatitis B-related hepatocellular carcinoma lesions are associated with aberrant tumor expression profiles of glypican 3 and osteopontin. *Ann Surg Oncol* 2012; **19** Suppl 3: S455-S463 [PMID: 21822558 DOI: 10.1245/s10434-011-1946-2]
- 130 **Huang H,** Zhang XF, Zhou HJ, Xue YH, Dong QZ, Ye QH, Qin LX. Expression and prognostic significance of osteopontin and caspase-3 in hepatocellular carcinoma patients after curative resection. *Cancer Sci* 2010; **101**: 1314-1319 [PMID: 20345480 DOI: 10.1111/j.1349-7006.2010.01524.x]
- 131 **Deng B,** Zhang XF, Zhu XC, Huang H, Jia HL, Ye QH, Dong QZ, Qin LX. Correlation and prognostic value of osteopontin and Bcl-2 in hepatocellular carcinoma patients after curative resection. *Oncol Rep* 2013; **30**: 2795-2803 [PMID: 24065086 DOI: 10.3892/or.2013.2737]
- 132 **Zhu W,** Guo L, Zhang B, Lou L, Lin Z, Zhu X, Ren N, Dong Q, Ye Q, Qin L. Combination of osteopontin with peritumoral infiltrating macrophages is associated with poor prognosis of early-stage hepatocellular carcinoma after curative resection. *Ann Surg Oncol* 2014; **21**: 1304-1313 [PMID: 24366422 DOI: 10.1245/s10434-013-3445-0]
- 133 **Weber GF.** The cancer biomarker osteopontin: combination with other markers. *Cancer Genomics Proteomics* 2011; **8**: 263-288 [PMID: 22086896]
- 134 **Kim N,** Kim JE, Choung HK, Lee MJ, Khwarg SI. Expression of cell cycle regulatory proteins in eyelid sebaceous gland carcinoma: low p27 expression predicts poor prognosis. *Exp Eye Res* 2014; **118**: 46-52 [PMID: 24216315 DOI: 10.1016/j.exer.2013.10.022]
- 135 **Aoyagi K,** Kouhiji K, Miyagi M, Imaizumi T, Kizaki J, Isobe T, Shirouzu K. Expression of p27Kip1 protein in gastric carcinoma. *Hepatogastroenterology* 2013; **60**: 390-394 [PMID: 23858559]
- 136 **Matsuda Y,** Wakai T, Kubota M, Takamura M, Yamagiwa S, Aoyagi Y, Osawa M, Fujimaki S, Sanpei A, Genda T, Ichida T. Clinical significance of cell cycle inhibitors in hepatocellular carcinoma. *Med Mol Morphol* 2013; **46**: 185-192 [PMID: 23640750 DOI: 10.1007/s00795-013-0047-7]
- 137 **Wan C,** Hou S, Ni R, Lv L, Ding Z, Huang X, Hang Q, He S, Wang Y, Cheng C, Gu XX, Xu G, Shen A. MIF4G domain containing protein regulates cell cycle and hepatic carcinogenesis by antagonizing CDK2-dependent p27 stability. *Oncogene* 2013; Epub ahead of print [PMID: 24336329 DOI: 10.1038/onc.2013.536]
- 138 **Fu X,** Wang Q, Chen J, Huang X, Chen X, Cao L, Tan H, Li W, Zhang L, Bi J, Su Q, Chen L. Clinical significance of miR-221 and its inverse correlation with p27Kip1 in hepatocellular carcinoma. *Mol Biol Rep* 2011; **38**: 3029-3035 [PMID: 20146005 DOI: 10.1007/s11033-010-9969-5]
- 139 **Shen DY,** Fang ZX, You P, Liu PG, Wang F, Huang CL, Yao XB, Chen ZX, Zhang ZY. Clinical significance and expression of cyclin kinase subunits 1 and 2 in hepatocellular carcinoma. *Liver Int* 2010; **30**: 119-125 [PMID: 19845855 DOI: 10.1111/j.1478-3231.2009.02106.x]
- 140 **Shehata MA,** Nosseir HR, Nagy HM, Farouk G. Cyclin dependent kinase inhibitor p27(kip1) expression and subcellular localization in relation to cell proliferation in hepatocellular carcinoma. *Egypt J Immunol* 2006; **13**: 115-130 [PMID: 17974156]
- 141 **Tannapfel A,** Grund D, Katalinic A, Uhlmann D, Köckerling F, Haugwitz U, Wasner M, Hauss J, Engeland K, Wittekind C. Decreased expression of p27 protein is associated with advanced tumor stage in hepatocellular carcinoma. *Int J Cancer* 2000; **89**: 350-355 [PMID: 10956409 DOI: 10.1002/1097-0215(20000720)89:4<350::AID-IJC6>3.0.CO;2-3]
- 142 **Armengol C,** Boix L, Bachs O, Solé M, Fuster J, Sala M, Llovet JM, Rodés J, Bruix J. p27(Kip1) is an independent predictor of recurrence after surgical resection in patients with small hepatocellular carcinoma. *J Hepatol* 2003; **38**: 591-597 [PMID: 12713869 DOI: 10.1016/S0168-8278(03)00025-4]
- 143 **Zhou Q,** He Q, Liang LJ. Expression of p27, cyclin E and cyclin A in hepatocellular carcinoma and its clinical significance. *World J Gastroenterol* 2003; **9**: 2450-2454 [PMID: 14606074]
- 144 **Chen L,** Yuan D, Wang GL, Wang Y, Wu YY, Zhu J. Clinicopathological significance of expression of Tspan-1, Jab1 and p27 in human hepatocellular carcinoma. *J Korean Med Sci* 2010; **25**: 1438-1442 [PMID: 20890423 DOI: 10.3346/jkms.2010.25.10.1438]
- 145 **Huang CW,** Lin CY, Huang HY, Liu HW, Chen YJ, Shih DF, Chen HY, Juan CC, Ker CG, Huang CY, Li CF, Shiue YL. CKS1B overexpression implicates clinical aggressiveness of hepatocellular carcinomas but not p27(Kip1) protein turnover: an independent prognosticator with potential p27 (Kip1)-independent oncogenic attributes? *Ann Surg Oncol* 2010; **17**: 907-922 [PMID: 19866239 DOI: 10.1245/s10434-009-0779-8]
- 146 **Matsuda Y.** Molecular mechanism underlying the functional loss of cyclindependent kinase inhibitors p16 and p27 in hepatocellular carcinoma. *World J Gastroenterol* 2008; **14**: 1734-1740 [PMID: 18350604 DOI: 10.3748/wjg.14.1734]
- 147 **Fiorentino M,** Altimari A, D'Errico A, Cukor B, Barozzi C, Loda M, Grigioni WF. Acquired expression of p27 is a favorable prognostic indicator in patients with hepatocellular carcinoma. *Clin Cancer Res* 2000; **6**: 3966-3972 [PMID: 11051245]
- 148 **Wander SA,** Zhao D, Slingerland JM. p27: a barometer of signaling deregulation and potential predictor of response to targeted therapies. *Clin Cancer Res* 2011; **17**: 12-18 [PMID: 20966355 DOI: 10.1158/1078-0432.CCR-10-0752]
- 149 **Liu Z,** Long Y, Zhang Y, Huang W, Long X, Yang H, Long J, Cheng C, Fang W. Nuclear p27 expression confers a favorable outcome for nasopharyngeal carcinoma patients. *Dis Markers* 2013; **35**: 925-932 [PMID: 24427780 DOI: 10.1155/2013/251209]
- 150 **Shen J,** Yin JY, Li XP, Liu ZQ, Wang Y, Chen J, Qu J, Xu XJ, McLeod HL, He YJ, Xia K, Jia YW, Zhou HH. The prognostic value of altered eIF3a and its association with p27 in non-small cell lung cancers. *PLoS One* 2014; **9**: e96008 [PMID: 24789280 DOI: 10.1371/journal.pone.0096008]
- 151 **Watanabe A,** Suzuki H, Yokobori T, Tsukagoshi M, Altan B, Kubo N, Suzuki S, Araki K, Wada S, Kashiwabara K, Hosouchi Y, Kuwano H. Stathmin1 regulates p27 expression, proliferation and drug resistance, resulting in poor clinical prognosis in cholangiocarcinoma. *Cancer Sci* 2014; **105**: 690-696 [PMID: 24708177 DOI: 10.1111/cas.12417]
- 152 **Al-Maghrabi J,** Al-Ahwal M, Buhmeida A, Syrjänen K, Sibyani A, Emam E, Ghanim A, Al-Qahtani M. Expression of cell cycle regulators p21 and p27 as predictors of disease outcome in colorectal carcinoma. *J Gastrointest Cancer* 2012;

- 43: 279-287 [PMID: 21637966 DOI: 10.1007/s12029-011-9292-y]
- 153 **Farley J**, Smith LM, Darcy KM, Brady MF, Bell J, McGuire W, Birrer MJ. Nuclear P27 expression in benign, borderline (LMP) and invasive tumors of the ovary and its association with prognosis: a gynecologic oncology group study. *Gynecol Oncol* 2011; **121**: 395-401 [PMID: 21310472 DOI: 10.1016/j.ygyno.2010.11.023]
- 154 **Kruck S**, Merseburger AS, Hennenlotter J, Scharpf M, Eyrich C, Amend B, Sievert KD, Stenzl A, Bedke J. High cytoplasmic expression of p27(Kip1) is associated with a worse cancer-specific survival in clear cell renal cell carcinoma. *BJU Int* 2012; **109**: 1565-1570 [PMID: 21981759 DOI: 10.1111/j.1464-410X.2011.10649.x]
- 155 **Chen G**, Cheng Y, Zhang Z, Martinka M, Li G. Prognostic significance of cytoplasmic p27 expression in human melanoma. *Cancer Epidemiol Biomarkers Prev* 2011; **20**: 2212-2221 [PMID: 21828232 DOI: 10.1158/1055-9965.EPI-11-0472]
- 156 **Lee YH**, Heo JH, Kim TH, Kang H, Kim G, Kim J, Cho SH, An HJ. Significance of cell cycle regulatory proteins as malignant and prognostic biomarkers in ovarian epithelial tumors. *Int J Gynecol Pathol* 2011; **30**: 205-217 [PMID: 21464733 DOI: 10.1097/PGP.0b013e3182063e71]
- 157 **Singh SP**, Lipman J, Goldman H, Ellis FH, Aizenman L, Cangi MG, Signoretti S, Chiaur DS, Pagano M, Loda M. Loss or altered subcellular localization of p27 in Barrett's associated adenocarcinoma. *Cancer Res* 1998; **58**: 1730-1735 [PMID: 9563491]
- 158 **Nan KJ**, Jing Z, Gong L. Expression and altered subcellular localization of the cyclin-dependent kinase inhibitor p27Kip1 in hepatocellular carcinoma. *World J Gastroenterol* 2004; **10**: 1425-1430 [PMID: 15133847]
- 159 **Zhang J**, McCauley MJ, Maher LJ, Williams MC, Israeloff NE. Mechanism of DNA flexibility enhancement by HMGB proteins. *Nucleic Acids Res* 2009; **37**: 1107-1114 [PMID: 19129233 DOI: 10.1093/nar/gkn1011]
- 160 **Süren D**, Yıldırım M, Demirpençe Ö, Kaya V, Alikanoğlu AS, Bülbüller N, Yıldız M, Sezer C. The role of high mobility group box 1 (HMGB1) in colorectal cancer. *Med Sci Monit* 2014; **20**: 530-537 [PMID: 24681824 DOI: 10.12659/MSM.890531]
- 161 **Chen RC**, Yi PP, Zhou RR, Xiao MF, Huang ZB, Tang DL, Huang Y, Fan XG. The role of HMGB1-RAGE axis in migration and invasion of hepatocellular carcinoma cell lines. *Mol Cell Biochem* 2014; **390**: 271-280 [PMID: 24510323 DOI: 10.1007/s11010-014-1978-6]
- 162 **Tang D**, Kang R, Zeh HJ, Lotze MT. High-mobility group box 1 and cancer. *Biochim Biophys Acta* 2010; **1799**: 131-140 [PMID: 20123075 DOI: 10.1016/j.bbagr.2009.11.014]
- 163 **Tang D**, Kang R, Zeh HJ, Lotze MT. High-mobility group box 1, oxidative stress, and disease. *Antioxid Redox Signal* 2011; **14**: 1315-1335 [PMID: 20969478 DOI: 10.1089/ars.2010.3356]
- 164 **Jiang W**, Wang Z, Li X, Li J, Huang Y, Fan X, Duan Y. Reduced high-mobility group box 1 expression induced by RNA interference inhibits the bioactivity of hepatocellular carcinoma cell line HCCLM3. *Dig Dis Sci* 2012; **57**: 92-98 [PMID: 22038506 DOI: 10.1007/s10620-011-1944-z]
- 165 **Dong YD**, Cui L, Peng CH, Cheng DF, Han BS, Huang F. Expression and clinical significance of HMGB1 in human liver cancer: Knockdown inhibits tumor growth and metastasis in vitro and in vivo. *Oncol Rep* 2013; **29**: 87-94 [PMID: 23042506 DOI: 10.3892/or.2012.2070]
- 166 **Wang C**, Tang C, Chang X, Li Z. [Effect of HMGB1 on invasion and migration of human hepatoma cell line HepG2 and its mechanism]. *Xibao Yu Fenzi Mianyixue Zazhi* 2013; **29**: 1159-1162 [PMID: 24200063]
- 167 **Yan W**, Chang Y, Liang X, Cardinal JS, Huang H, Thorne SH, Monga SP, Geller DA, Lotze MT, Tsung A. High-mobility group box 1 activates caspase-1 and promotes hepatocellular carcinoma invasiveness and metastases. *Hepatology* 2012; **55**: 1863-1875 [PMID: 22234969 DOI: 10.1002/hep.25572]
- 168 **Cheng BQ**, Jia CQ, Liu CT, Lu XF, Zhong N, Zhang ZL, Fan W, Li YQ. Serum high mobility group box chromosomal protein 1 is associated with clinicopathologic features in patients with hepatocellular carcinoma. *Dig Liver Dis* 2008; **40**: 446-452 [PMID: 18294942 DOI: 10.1016/j.dld.2007.11.024]
- 169 **Liu F**, Zhang Y, Peng Z, Gao H, Xu L, Chen M. High expression of high mobility group box 1 (hmgbl) predicts poor prognosis for hepatocellular carcinoma after curative hepatectomy. *J Transl Med* 2012; **10**: 135 [PMID: 22747650 DOI: 10.1186/1479-5876-10-135]
- 170 **Jiang W**, Wang Z, Li X, Fan X, Duan Y. High-mobility group box 1 is associated with clinicopathologic features in patients with hepatocellular carcinoma. *Pathol Oncol Res* 2012; **18**: 293-298 [PMID: 21953322 DOI: 10.1007/s12253-011-9442-3]
- 171 **Xiao J**, Ding Y, Huang J, Li Q, Liu Y, Ni W, Zhang Y, Zhu Y, Chen L, Chen B. The association of HMGB1 gene with the prognosis of HCC. *PLoS One* 2014; **9**: e89097 [PMID: 24586525 DOI: 10.1371/journal.pone.0089097]
- 172 **Adhikari AS**, Agarwal N, Iwakuma T. Metastatic potential of tumor-initiating cells in solid tumors. *Front Biosci (Landmark Ed)* 2011; **16**: 1927-1938 [PMID: 21196274 DOI: 10.2741/3831]
- 173 **Guo W**. Concise review: breast cancer stem cells: regulatory networks, stem cell niches, and disease relevance. *Stem Cells Transl Med* 2014; **3**: 942-948 [PMID: 24904174 DOI: 10.5966/sctm.2014-0020]
- 174 **Naujokat C**. Monoclonal antibodies against human cancer stem cells. *Immunotherapy* 2014; **6**: 290-308 [PMID: 24762074 DOI: 10.2217/imt.14.4]
- 175 **Shen Y**, Cao D. Hepatocellular carcinoma stem cells: origins and roles in hepatocarcinogenesis and disease progression. *Front Biosci (Elite Ed)* 2012; **4**: 1157-1169 [PMID: 22201943 DOI: 10.2741/E448]
- 176 **Yamashita T**, Wang XW. Cancer stem cells in the development of liver cancer. *J Clin Invest* 2013; **123**: 1911-1918 [PMID: 23635789 DOI: 10.1172/JCI66024]
- 177 **Yeh CT**, Kuo CJ, Lai MW, Chen TC, Lin CY, Yeh TS, Lee WC. CD133-positive hepatocellular carcinoma in an area endemic for hepatitis B virus infection. *BMC Cancer* 2009; **9**: 324 [PMID: 19744348 DOI: 10.1186/1471-2407-9-324]
- 178 **Yang XR**, Xu Y, Yu B, Zhou J, Qiu SJ, Shi GM, Zhang BH, Wu WZ, Shi YH, Wu B, Yang GH, Ji Y, Fan J. High expression levels of putative hepatic stem/progenitor cell biomarkers related to tumour angiogenesis and poor prognosis of hepatocellular carcinoma. *Gut* 2010; **59**: 953-962 [PMID: 20442200 DOI: 10.1136/gut.2008.176271]
- 179 **Bae JS**, Noh SJ, Jang KY, Park HS, Chung MJ, Park CK, Moon WS. Expression and role of epithelial cell adhesion molecule in dysplastic nodule and hepatocellular carcinoma. *Int J Oncol* 2012; **41**: 2150-2158 [PMID: 22993038 DOI: 10.3892/ijo.2012.1631]
- 180 **Kim H**, Choi GH, Na DC, Ahn EY, Kim GI, Lee JE, Cho JY, Yoo JE, Choi JS, Park YN. Human hepatocellular carcinomas with "Stemness"-related marker expression: keratin 19 expression and a poor prognosis. *Hepatology* 2011; **54**: 1707-1717 [PMID: 22045674 DOI: 10.1002/hep.24559]
- 181 **Izumi N**. Prediction and prevention of intrahepatic recurrence of hepatocellular carcinoma. *Hepatol Res* 2012; **42**: 226-232 [PMID: 22181559 DOI: 10.1111/j.1872-034X.2011.00922.x]
- 182 **Zhang KZ**, Zhang QB, Zhang QB, Sun HC, Ao JY, Chai ZT, Zhu XD, Lu L, Zhang YY, Bu Y, Kong LQ, Tang ZY. Arsenic trioxide induces differentiation of CD133+ hepatocellular carcinoma cells and prolongs posthepatectomy survival by targeting GLI1 expression in a mouse model. *J Hematol Oncol* 2014; **7**: 28 [PMID: 24678763 DOI: 10.1186/1756-8722-7-28]
- 183 **Shi JH**, Scholz H, Huitfeldt HS, Line PD. The effect of hepatic progenitor cells on experimental hepatocellular carcinoma in the regenerating liver. *Scand J Gastroenterol* 2014; **49**: 99-108 [PMID: 24188385 DOI: 10.3109/00365521.201

- 3.854406]
- 184 **Tsuchiya A**, Kamimura H, Takamura M, Yamagiwa S, Matsuda Y, Sato Y, Nomoto M, Ichida T, Aoyagi Y. Clinicopathological analysis of CD133 and NCAM human hepatic stem/progenitor cells in damaged livers and hepatocellular carcinomas. *Hepatol Res* 2009; **39**: 1080-1090 [PMID: 19619253 DOI: 10.1111/j.1872-034X.2009.00559.x]
 - 185 **Song W**, Li H, Tao K, Li R, Song Z, Zhao Q, Zhang F, Dou K. Expression and clinical significance of the stem cell marker CD133 in hepatocellular carcinoma. *Int J Clin Pract* 2008; **62**: 1212-1218 [PMID: 18479363 DOI: 10.1111/j.1742-1241.2008.01777.x]
 - 186 **Guo Z**, Li LQ, Jiang JH, Ou C, Zeng LX, Xiang BD. Cancer stem cell markers correlate with early recurrence and survival in hepatocellular carcinoma. *World J Gastroenterol* 2014; **20**: 2098-2106 [PMID: 24616575 DOI: 10.3748/wjg.v20.i8.2098]
 - 187 **Sasaki A**, Kamiyama T, Yokoo H, Nakanishi K, Kubota K, Haga H, Matsushita M, Ozaki M, Matsuno Y, Todo S. Cytoplasmic expression of CD133 is an important risk factor for overall survival in hepatocellular carcinoma. *Oncol Rep* 2010; **24**: 537-546 [PMID: 20596644 DOI: 10.3892/or.00000890]
 - 188 **Chan AW**, Tong JH, Chan SL, Lai PB, To KF. Expression of stemness markers (CD133 and EpCAM) in prognostication of hepatocellular carcinoma. *Histopathology* 2014; **64**: 935-950 [PMID: 24506513 DOI: 10.1111/his.12342]
 - 189 **Kimura O**, Kondo Y, Kogure T, Kakazu E, Ninomiya M, Iwata T, Morosawa T, Shimosegawa T. Expression of EpCAM increases in the hepatitis B related and the treatment-resistant hepatocellular carcinoma. *Biomed Res Int* 2014; **2014**: 172913 [PMID: 24696843 DOI: 10.1155/2014/172913]
 - 190 **Terris B**, Cavard C, Perret C. EpCAM, a new marker for cancer stem cells in hepatocellular carcinoma. *J Hepatol* 2010; **52**: 280-281 [PMID: 20006402 DOI: 10.1016/j.jhep.2009.10.026]
 - 191 **Kimura O**, Takahashi T, Ishii N, Inoue Y, Ueno Y, Kogure T, Fukushima K, Shiina M, Yamagiwa Y, Kondo Y, Inoue J, Kakazu E, Iwasaki T, Kawagishi N, Shimosegawa T, Sugamura K. Characterization of the epithelial cell adhesion molecule (EpCAM)+ cell population in hepatocellular carcinoma cell lines. *Cancer Sci* 2010; **101**: 2145-2155 [PMID: 20707805 DOI: 10.1111/j.1349-7006.2010.01661.x]
 - 192 **Shan YF**, Huang YL, Xie YK, Tan YH, Chen BC, Zhou MT, Shi HQ, Yu ZP, Song QT, Zhang QY. Angiogenesis and clinicopathologic characteristics in different hepatocellular carcinoma subtypes defined by EpCAM and α -fetoprotein expression status. *Med Oncol* 2011; **28**: 1012-1016 [PMID: 20571936 DOI: 10.1007/s12032-010-9600-6]
 - 193 **Chung GE**, Lee JH, Yoon JH, Myung SJ, Lee K, Jang JJ, Lee JM, Kim SH, Suh KS, Kim YJ, Lee HS. Prognostic implications of tumor vascularity and its relationship to cytokeratin 19 expression in patients with hepatocellular carcinoma. *Abdom Imaging* 2012; **37**: 439-446 [PMID: 21584634 DOI: 10.1007/s00261-011-9756-3]
 - 194 **Yang XR**, Xu Y, Shi GM, Fan J, Zhou J, Ji Y, Sun HC, Qiu SJ, Yu B, Gao Q, He YZ, Qin WZ, Chen RX, Yang GH, Wu B, Lu Q, Wu ZQ, Tang ZY. Cytokeratin 10 and cytokeratin 19: predictive markers for poor prognosis in hepatocellular carcinoma patients after curative resection. *Clin Cancer Res* 2008; **14**: 3850-3859 [PMID: 18559605 DOI: 10.1158/1078-0432.CCR-07-4338]
 - 195 **Ariizumi S**, Kotera Y, Katagiri S, Nakano M, Yamamoto M. Combined hepatocellular-cholangiocarcinoma had poor outcomes after hepatectomy regardless of Allen and Lisa class or the predominance of intrahepatic cholangiocarcinoma cells within the tumor. *Ann Surg Oncol* 2012; **19**: 1628-1636 [PMID: 22113592 DOI: 10.1245/s10434-011-2150-0]
 - 196 **Andersen JB**, Loi R, Perra A, Factor VM, Ledda-Columbano GM, Columbano A, Thorgerirsson SS. Progenitor-derived hepatocellular carcinoma model in the rat. *Hepatology* 2010; **51**: 1401-1409 [PMID: 20054870 DOI: 10.1002/hep.23488]
 - 197 **Xu M**, Xie F, Qian G, Jing Y, Zhang S, Gao L, Zheng T, Wu M, Yang J, Wei L. Peritumoral ductular reaction: a poor postoperative prognostic factor for hepatocellular carcinoma. *BMC Cancer* 2014; **14**: 65 [PMID: 24495509 DOI: 10.1186/1471-2407-14-65]
 - 198 **Lee CW**, Kuo WL, Yu MC, Chen TC, Tsai CN, Lee WC, Chen MF. The expression of cytokeratin 19 in lymph nodes was a poor prognostic factor for hepatocellular carcinoma after hepatic resection. *World J Surg Oncol* 2013; **11**: 136 [PMID: 23758804 DOI: 10.1186/1477-7819-11-136]
 - 199 **Wang ZS**, Wu LQ, Yi X, Geng C, Li YJ, Yao RY, Hu WY, Han B. [CK19 can be used to predict the early recurrence and prognosis of HBV-related hepatocellular carcinoma patients with low AFP serum concentration after R0 radical hepatectomy]. *Zhonghua Zhongliu Zazhi* 2012; **34**: 753-758 [PMID: 23291069 DOI: 10.3760/cma.j.issn.0253-3766.2012.10.008]
 - 200 **Yuan RH**, Jeng YM, Hu RH, Lai PL, Lee PH, Cheng CC, Hsu HC. Role of p53 and β -catenin mutations in conjunction with CK19 expression on early tumor recurrence and prognosis of hepatocellular carcinoma. *J Gastrointest Surg* 2011; **15**: 321-329 [PMID: 21061181 DOI: 10.1007/s11605-010-1373-x]
 - 201 **Xiang ZL**, Zeng ZC, Tang ZY, Fan J, Sun HC, Wu WZ, Tan YS. [Nuclear accumulation of CXCR4 and overexpressions of VEGF-C and CK19 are associated with a higher risk of lymph node metastasis in hepatocellular carcinoma]. *Zhonghua Zhongliu Zazhi* 2010; **32**: 344-349 [PMID: 20723431]
 - 202 **Zhuang PY**, Zhang JB, Zhu XD, Zhang W, Wu WZ, Tan YS, Hou J, Tang ZY, Qin LX, Sun HC. Two pathologic types of hepatocellular carcinoma with lymph node metastasis with distinct prognosis on the basis of CK19 expression in tumor. *Cancer* 2008; **112**: 2740-2748 [PMID: 18412155 DOI: 10.1002/cncr.23488]
 - 203 **Ji J**, Wang XW. Clinical implications of cancer stem cell biology in hepatocellular carcinoma. *Semin Oncol* 2012; **39**: 461-472 [PMID: 22846863 DOI: 10.1053/j.seminoncol.2012.05.011]
 - 204 **Zhang L**, Liu H, Sun L, Li N, Ding H, Zheng J. Glypican-3 as a potential differential diagnosis marker for hepatocellular carcinoma: a tissue microarray-based study. *Acta Histochem* 2012; **114**: 547-552 [PMID: 22119409 DOI: 10.1016/j.acthis.2011.10.003]
 - 205 **Nassar A**, Cohen C, Siddiqui MT. Utility of glypican-3 and survivin in differentiating hepatocellular carcinoma from benign and preneoplastic hepatic lesions and metastatic carcinomas in liver fine-needle aspiration biopsies. *Diagn Cytopathol* 2009; **37**: 629-635 [PMID: 19405109 DOI: 10.1002/dc.21075]
 - 206 **Liu H**, Li P, Zhai Y, Qu CF, Zhang LJ, Tan YF, Li N, Ding HG. Diagnostic value of glypican-3 in serum and liver for primary hepatocellular carcinoma. *World J Gastroenterol* 2010; **16**: 4410-4415 [PMID: 20845507 DOI: 10.3748/wjg.v16.i35.4410]
 - 207 **Filmus J**, Capurro M. Glypican-3: a marker and a therapeutic target in hepatocellular carcinoma. *FEBS J* 2013; **280**: 2471-2476 [PMID: 23305321 DOI: 10.1111/febs.12126]
 - 208 **Shirakawa H**, Suzuki H, Shimomura M, Kojima M, Gotohda N, Takahashi S, Nakagohri T, Konishi M, Kobayashi N, Kinoshita T, Nakatsura T. Glypican-3 expression is correlated with poor prognosis in hepatocellular carcinoma. *Cancer Sci* 2009; **100**: 1403-1407 [PMID: 19496787 DOI: 10.1111/j.1349-7006.2009.01206.x]
 - 209 **Shirakawa H**, Kuronuma T, Nishimura Y, Hasebe T, Nakano M, Gotohda N, Takahashi S, Nakagohri T, Konishi M, Kobayashi N, Kinoshita T, Nakatsura T. Glypican-3 is a useful diagnostic marker for a component of hepatocellular carcinoma in human liver cancer. *Int J Oncol* 2009; **34**: 649-656 [PMID: 19212669]
 - 210 **Du JL**, Wang YL, Shi HY, Guo AT, Wei LX. [Expression of

- glypican-3, hepatocyte antigen, alpha-fetoprotein, CD34 and CD10 in hepatocellular carcinoma: a clinicopathologic analysis of 375 cases]. *Zhonghua Binglixue Zazhi* 2012; **41**: 309-313 [PMID: 22883669 DOI: 10.3760/cma.j.issn.0529-5807.2012.05.006]
- 211 **Fu SJ**, Qi CY, Xiao WK, Li SQ, Peng BG, Liang LJ. Glypican-3 is a potential prognostic biomarker for hepatocellular carcinoma after curative resection. *Surgery* 2013; **154**: 536-544 [PMID: 23601901 DOI: 10.1016/j.surg.2013.02.014]
- 212 **Ning S**, Bin C, Na H, Peng S, Yi D, Xiang-hua Y, Fang-yin Z, Da-yong Z, Rong-cheng L. Glypican-3, a novel prognostic marker of hepatocellular cancer, is related with postoperative metastasis and recurrence in hepatocellular cancer patients. *Mol Biol Rep* 2012; **39**: 351-357 [PMID: 21655958 DOI: 10.1007/s11033-011-0745-y]
- 213 **Yorita K**, Takahashi N, Takai H, Kato A, Suzuki M, Ishiguro T, Ohtomo T, Nagaike K, Kondo K, Chijiwa K, Kataoka H. Prognostic significance of circumferential cell surface immunoreactivity of glypican-3 in hepatocellular carcinoma. *Liver Int* 2011; **31**: 120-131 [PMID: 20964802 DOI: 10.1111/j.1478-3231.2010.02359.x]
- 214 **Serreels A**, Macpherson IR, Evans TR, Lee FY, Clark EA, Sansom OJ, Ashton GH, Frame MC, Brunton VG. Identification of potential biomarkers for measuring inhibition of Src kinase activity in colon cancer cells following treatment with dasatinib. *Mol Cancer Ther* 2006; **5**: 3014-3022 [PMID: 17148760 DOI: 10.1158/1535-7163.MCT-06-0382]
- 215 **Villanueva A**, Chiang DY, Newell P, Peix J, Thung S, Alsinet C, Tovar V, Roayaie S, Minguez B, Sole M, Battiston C, Van Laarhoven S, Fiel MI, Di Feo A, Hoshida Y, Yea S, Toffanin S, Ramos A, Martignetti JA, Mazzaferro V, Bruix J, Waxman S, Schwartz M, Meyerson M, Friedman SL, Llovet JM. Pivotal role of mTOR signaling in hepatocellular carcinoma. *Gastroenterology* 2008; **135**: 1972-1983, 1983.e1-11 [PMID: 18929564 DOI: 10.1053/j.gastro.2008.08.008]
- 216 **Zhou L**, Huang Y, Li J, Wang Z. The mTOR pathway is associated with the poor prognosis of human hepatocellular carcinoma. *Med Oncol* 2010; **27**: 255-261 [PMID: 19301157 DOI: 10.1007/s12032-009-9201-4]
- 217 **Calvisi DF**, Ladu S, Gorden A, Farina M, Lee JS, Conner EA, Schroeder I, Factor VM, Thorgeirsson SS. Mechanistic and prognostic significance of aberrant methylation in the molecular pathogenesis of human hepatocellular carcinoma. *J Clin Invest* 2007; **117**: 2713-2722 [PMID: 17717605 DOI: 10.1172/JCI31457]
- 218 **Nakanishi K**, Sakamoto M, Yamasaki S, Todo S, Hirohashi S. Akt phosphorylation is a risk factor for early disease recurrence and poor prognosis in hepatocellular carcinoma. *Cancer* 2005; **103**: 307-312 [PMID: 15593087 DOI: 10.1002/cncr.20774]
- 219 **Rowinsky EK**. Targeting the molecular target of rapamycin (mTOR). *Curr Opin Oncol* 2004; **16**: 564-575 [PMID: 15627018 DOI: 10.1097/01.cco.0000143964.74936.d1]
- 220 **Vignot S**, Faivre S, Aguirre D, Raymond E. mTOR-targeted therapy of cancer with rapamycin derivatives. *Ann Oncol* 2005; **16**: 525-537 [PMID: 15728109 DOI: 10.1093/annonc/mdi113]
- 221 **Cervello M**, McCubrey JA, Cusimano A, Lampiasi N, Azzolina A, Montalto G. Targeted therapy for hepatocellular carcinoma: novel agents on the horizon. *Oncotarget* 2012; **3**: 236-260 [PMID: 22470194]
- 222 **Sahin F**, Kannangai R, Adegbola O, Wang J, Su G, Torbenson M. mTOR and P70 S6 kinase expression in primary liver neoplasms. *Clin Cancer Res* 2004; **10**: 8421-8425 [PMID: 15623621 DOI: 10.1158/1078-0432.CCR-04-0941]
- 223 **Baba HA**, Wohlschlaeger J, Cicinnati VR, Hilgard P, Lang H, Sotiropoulos GC, Takeda A, Beckebaum S, Schmitz KJ. Phosphorylation of p70S6 kinase predicts overall survival in patients with clear margin-resected hepatocellular carcinoma. *Liver Int* 2009; **29**: 399-405 [PMID: 18492014 DOI: 10.1111/j.1478-3231.2008.01798.x]
- 224 **Chang Q**, Chen J, Beezhold KJ, Castranova V, Shi X, Chen F. JNK1 activation predicts the prognostic outcome of the human hepatocellular carcinoma. *Mol Cancer* 2009; **8**: 64 [PMID: 19686584 DOI: 10.1186/1476-4598-8-64]
- 225 **Wang Z**, Jin W, Jin H, Wang X. mTOR in viral hepatitis and hepatocellular carcinoma: function and treatment. *Biomed Res Int* 2014; **2014**: 735672 [PMID: 24804240 DOI: 10.1155/2014/735672]
- 226 **Matter MS**, Decaens T, Andersen JB, Thorgeirsson SS. Targeting the mTOR pathway in hepatocellular carcinoma: current state and future trends. *J Hepatol* 2014; **60**: 855-865 [PMID: 24308993 DOI: 10.1016/j.jhep.2013.11.031]
- 227 **Fleming S**, Mayer NJ, Vlatkovic LJ, McLean J, McConachie M, Baty D. Signalling pathways in succinate dehydrogenase B-associated renal carcinoma. *Histopathology* 2014; **64**: 477-483 [PMID: 24236567 DOI: 10.1111/his.12250]
- 228 **Prodromidis G**, Nikitakis NG, Sklavounou A. Immunohistochemical Analysis of the Activation Status of the Akt/mTOR/pS6 Signaling Pathway in Oral Lichen Planus. *Int J Dent* 2013; **2013**: 743456 [PMID: 24228033]
- 229 **Rouleau C**, Rico C, Hapkova I, de Santa Barbara P. Immunohistochemical analysis of bone morphological protein signaling pathway in human myometrium. *Exp Mol Pathol* 2012; **93**: 56-60 [PMID: 22537545 DOI: 10.1016/j.yexmp.2012.04.007]
- 230 **Siddiqui S**, Rimm DL. Pre-analytic variables and phospho-specific antibodies: the Achilles heel of immunohistochemistry. *Breast Cancer Res* 2010; **12**: 113 [PMID: 21176180 DOI: 10.1186/bcr2782]
- 231 **Schoephoerster J**, Frisch J, Grahek M, Wu C, He Y, Wang W, Nguyen J, Schwartz D, Kalyuzhny AE. Absorption control in immunohistochemistry using phospho-peptides immobilized on magnetic beads. *Methods Mol Biol* 2011; **717**: 291-300 [PMID: 21370038 DOI: 10.1007/978-1-61779-024-9_17]
- 232 **O'Hurley G**, Sjöstedt E, Rahman A, Li B, Kampf C, Pontén F, Gallagher WM, Lindskog C. Garbage in, garbage out: a critical evaluation of strategies used for validation of immunohistochemical biomarkers. *Mol Oncol* 2014; **8**: 783-798 [PMID: 24725481 DOI: 10.1016/j.molonc.2014.03.008]
- 233 **Gish RG**. Early detection of hepatocellular carcinoma through surveillance using biomarkers. *Gastroenterol Hepatol (N Y)* 2014; **10**: 121-123 [PMID: 24803876]

P- Reviewer: Hoare M, Lin ZY, Minuk G, Zhu X
S- Editor: Tian YL **L- Editor:** A **E- Editor:** Liu SQ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

