# World Journal of *Gastroenterology*

World J Gastroenterol 2024 September 28; 30(36): 4014-4082





Published by Baishideng Publishing Group Inc

J G  $\mathcal{N}$ 

# World Journal of Gastroenterology

#### Contents

Weekly Volume 30 Number 36 September 28, 2024

#### **EDITORIAL**

4014	Autophagy and its role in gastrointestinal diseases	
	Shao BZ, Zhang WG, Liu ZY, Linghu EQ	
4021	Redefining hemorrhoid therapy with endoscopic polidocanol foam sclerobanding	
	Rao AG, Nashwan AJ	
4025	Dual-targeted treatment for inflammatory bowel disease: Whether fecal microbiota transplantation can be an important part of it	
	Zhang ZN, Sang LX	
4031	Reconceptualization of immune checkpoint inhibitor-associated gastritis	
	Deng YF, Cui XS, Wang L	

Glucagon-like peptide-1 receptor agonists: Exploring the mechanisms from glycemic control to treatment 4036 of multisystemic diseases

Kong MW, Yu Y, Wan Y, Gao Y, Zhang CX

#### **ORIGINAL ARTICLE**

#### **Retrospective Study**

Computed tomography-based multi-organ radiomics nomogram model for predicting the risk of 4044 esophagogastric variceal bleeding in cirrhosis

Peng YJ, Liu X, Liu Y, Tang X, Zhao QP, Du Y

#### **Basic Study**

4057 Construction and validation of a pancreatic cancer prognostic model based on genes related to the hypoxic tumor microenvironment

Yang F, Jiang N, Li XY, Qi XS, Tian ZB, Guo YJ

#### **CASE REPORT**

4071 Liver transplantation following two conversions in a patient with huge hepatocellular carcinoma and portal vein invasion: A case report

Liang LC, Huang WS, Guo ZX, You HJ, Guo YJ, Cai MY, Lin LT, Wang GY, Zhu KS

#### LETTER TO THE EDITOR

4078 Beyond bacteria: Role of non-bacterial gut microbiota species in inflammatory bowel disease and colorectal cancer progression

Haque H, Zehra SW, Shahzaib M, Abbas S, Jaffar N



#### Contents

Weekly Volume 30 Number 36 September 28, 2024

#### **ABOUT COVER**

Editorial Board Member of World Journal of Gastroenterology, Olga A Sukocheva, MSc, PhD, Assistant Professor, Senior Researcher, Department of Hepatology, Royal Adelaide Hospital, Adelaide 5000, South Australia, Australia. olga.sukocheva@sa.gov.au

#### **AIMS AND SCOPE**

The primary aim of World Journal of Gastroenterology (WJG, World J Gastroenterol) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

#### **INDEXING/ABSTRACTING**

The WJG is now abstracted and indexed in Science Citation Index Expanded (SCIE), MEDLINE, PubMed, PubMed Central, Scopus, Reference Citation Analysis, China Science and Technology Journal Database, and Superstar Journals Database. The 2024 edition of Journal Citation Reports® cites the 2023 journal impact factor (JIF) for WJG as 4.3; Quartile: Q1. The WJG's CiteScore for 2023 is 7.8.

#### **RESPONSIBLE EDITORS FOR THIS ISSUE**

Production Editor: Yu-Xi Chen, Production Department Director: Xiang Li, Cover Editor: Jia-Ru Fan.

NAME OF JOURNAL World Journal of Gastroenterology	INSTRUCTIONS TO AUTHORS https://www.wjgnet.com/bpg/gerinfo/204
w orm fourna of Gustoenerougy	https://www.wjghet.com/bpg/gennio/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 1007-9327 (print) ISSN 2219-2840 (online)	https://www.wjgnet.com/bpg/GerInfo/287
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
October 1, 1995	https://www.wjgnet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Weekly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Andrzej S Tarnawski	https://www.wjgnet.com/bpg/gerinfo/208
EXECUTIVE ASSOCIATE EDITORS-IN-CHIEF	POLICY OF CO-AUTHORS
Jian-Gao Fan (Chronic Liver Disease)	https://www.wjgnet.com/bpg/GerInfo/310
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
http://www.wjgnet.com/1007-9327/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
September 28, 2024	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2024 Baishideng Publishing Group Inc	https://www.f6publishing.com
PUBLISHING PARTNER	PUBLISHING PARTNER'S OFFICIAL WEBSITE
Shanghai Pancreatic Cancer Institute and Pancreatic Cancer Institute, Fudan University Biliary Tract Disease Institute, Fudan University	https://www.shca.org.cn https://www.zs-hospital.sh.cn

© 2024 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: office@baishideng.com https://www.wjgnet.com



WŨ

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2024 September 28; 30(36): 4014-4020

DOI: 10.3748/wjg.v30.i36.4014

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

EDITORIAL

## Autophagy and its role in gastrointestinal diseases

Bo-Zong Shao, Wen-Gang Zhang, Zhen-Yu Liu, En-Qiang Linghu

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's classification Scientific Quality: Grade C, Grade С

Novelty: Grade C, Grade C Creativity or Innovation: Grade C, Grade C Scientific Significance: Grade B, Grade C

P-Reviewer: Durairajan SSK

Received: March 12, 2024 Revised: August 25, 2024 Accepted: September 9, 2024 Published online: September 28, 2024 Processing time: 192 Days and 3.4 Hours



Bo-Zong Shao, Wen-Gang Zhang, Zhen-Yu Liu, En-Qiang Linghu, Department of Gastroenterology, First Medical Center of Chinese People's Liberation Army General Hospital, Beijing 100853, China

Co-first authors: Bo-Zong Shao and Wen-Gang Zhang.

Corresponding author: En-Qiang Linghu, MD, PhD, Professor, Chief, Department of Gastroenterology, First Medical Center of Chinese People's Liberation Army General Hospital, No. 28 Fuxing Road, Beijing 100853, China. linghuenqiang@vip.sina.com

### Abstract

Gastrointestinal disorders encompass a spectrum of conditions affecting various organs within the digestive system, such as the esophagus, stomach, colon, rectum, pancreas, liver, small intestine, and bile ducts. The role of autophagy in the etiology and progression of gastrointestinal diseases has garnered significant attention. This paper seeks to evaluate the impact and mechanisms of autophagy in gastrointestinal disorders by synthesizing recent research findings. Specifically, we delve into inflammation-related gastrointestinal conditions, including ulcerative colitis, Crohn's disease, and pancreatitis, as well as gastrointestinal cancers such as esophageal, gastric, and colorectal cancers. Additionally, we provide commentary on a recent publication by Chang et al in the World Journal of Gastroenterology. Our objective is to offer fresh perspectives on the mechanisms and therapeutic approaches for these gastrointestinal ailments. This review aims to offer new perspectives on the mechanisms and therapeutic strategies for gastrointestinal disorders by critically analyzing relevant publications. As discussed, the role of autophagy in gastrointestinal diseases is complex and, at times, contentious. To harness the full therapeutic potential of autophagy in treating these conditions, more in-depth research is imperative.

Key Words: Autophagy; Inflammation; Cancer; Inflammatory bowel disease; Pancreatitis

©The Author(s) 2024. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Extensive research has implicated autophagy in the pathogenesis and advancement of diverse gastrointestinal disorders. Nevertheless, the precise role of autophagy in these ailments remains incompletely understood, and the specific underlying mechanisms remain elusive. Consequently, further investigation is warranted to address these gaps in knowledge.



WJG | https://www.wjgnet.com

**Citation:** Shao BZ, Zhang WG, Liu ZY, Linghu EQ. Autophagy and its role in gastrointestinal diseases. *World J Gastroenterol* 2024; 30(36): 4014-4020

**URL:** https://www.wjgnet.com/1007-9327/full/v30/i36/4014.htm **DOI:** https://dx.doi.org/10.3748/wjg.v30.i36.4014

#### INTRODUCTION

The term "autophagy" was coined in the 1960s[1]. Derived from the Greek roots "auto" (self) and "phagy" (eat), autophagy describes cellular metabolic processes in which cytoplasmic proteins and specific organelles are self-degraded [2]. Since its inception, the mechanisms of autophagy have been extensively explored by the scientific community. Research on autophagy won the Nobel prize in physiology or medicine in 2016[3]. Classic autophagy is classified into three types: Micro-autophagy, chaperone-mediated autophagy, and macro-autophagy[3]. Micro-autophagy represents a nonselective lysosomal process. Chaperone-mediated autophagy operates as a discerning form of autophagy that hinges on the recognition of chaperones through specific motifs within the targeted proteins, as well as lysosomal chaperones. Macro-autophagy, on the other hand, is the most extensively investigated variant of autophagy. It is a metabolic process characterized by the formation of double-membraned autophagosomes, which serve as functional units that subsequently merge with lysosomes to facilitate further degradation and recycling. In addition to traditional autophagy, specialized selective autophagy, such as pexophagy and reticulophagy, has been identified. These distinct varieties of selective autophagy embody unique roles and functions within specific organelles and under specific environmental conditions.

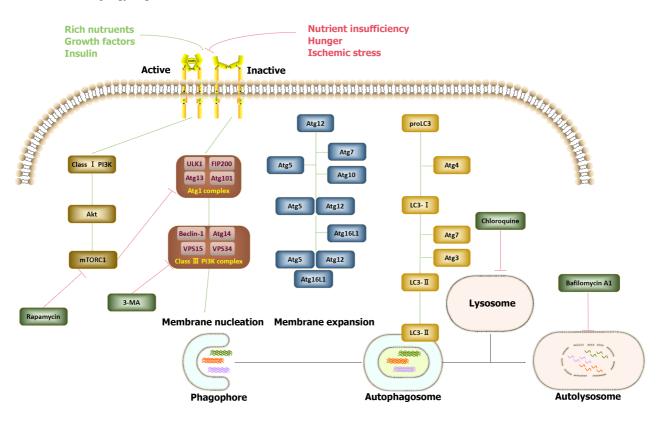
The autophagy-lysosomal system represents one of the classical pathways for protein degradation, alongside the ubiquitin-proteasome system. This intricate process is orchestrated by over 30 autophagy-related genes (Atgs), the majority of which are conserved in mammalian cells. Autophagy induction is a two-step process[4] (illustrated in Figure 1). Initially, under conditions of stress, such as hypoxia, phagophores form to envelop substrates. Phagophore formation necessitates the assembly of the Atg1 complex and the class III phosphatidylinositol 3-kinase (PI3K) complex. The Atg1 complex includes Unc-51-like kinase, focal adhesion kinase family interacting protein of 200 kD, ATG13, and ATG101. Following initiation, the bilayer membrane undergoes a series of processes, including expansion, elongation, and nucleation, leading to the formation of double-membrane, sphere-shaped autophagosomes. This process relies on the assembly of the ATG16L1 complex, which is composed of ATG5, ATG12, and ATG16L1, as well as the participation of two ubiquitin-like proteins, ATG12 and Atg8 (light chain 3). In the second step, autophagosomes subsequently shed their "coat proteins" [light chain 3-II (LC3-II)] and merge with lysosomes to form functional autolysosomes. This process is facilitated by ATG3 and ATG7. The class I PI3K-mammalian target of rapamycin (mTOR) pathway acts as an inhibitory regulator of autophagy, whereas the class III PI3K pathway serves as an autophagy inducer. Various autophagy inducers and inhibitors have been utilized in experimental studies and clinical practice to modulate autophagy levels<sup>[5]</sup>. For example, rapamycin is commonly employed to increase autophagy levels by inhibiting mTORC1 activation. Conversely, the inhibition of autophagy is achieved through mechanisms such as 3-methyladenine. Additionally, chloroquine disrupts autophagy through its influence on the acidic environment of lysosomes, whereas bafilomycin A1 disturbs the formation of autolysosomes.

On the basis of previous research both within our laboratory and from other sources[4], autophagy has been demonstrated to actively participate in and regulate numerous diseases across various domains. In the context of gastrointestinal diseases, the role of autophagy in influencing the pathogenesis and progression of these conditions has been extensively investigated. The influence of autophagy is exerted through the regulation of inflammation and immune responses, as well as the modulation of cellular functions and biological status, including gastrointestinal epithelial cells, inflammatory cells, and even cancer cells. A recent publication by Chang *et al*[6] in the *World Journal of Gastroenterology* comprehensively reviewed the impact of autophagy on the functions of gastrointestinal cells, including digestive cells, secretory cells, regenerative cells, and physical barriers. The authors illustrate how autophagy plays a pivotal role in influencing several gastrointestinal diseases by modulating inflammatory reactions. As a result, autophagy is a crucial determinant in maintaining homeostasis within the digestive system. In the following sections, we comprehensively examine the role of autophagy in various well-characterized gastrointestinal diseases, including inflammation-associated conditions such as ulcerative colitis (UC), Crohn's disease (CD), and pancreatitis, as well as gastrointestinal malignancies such as esophageal, gastric, and colorectal cancers.

#### AUTOPHAGY IN INFLAMMATION-RELATED GASTROINTESTINAL DISEASES

UC is a type of inflammatory bowel disease (IBD) characterized by chronic inflammation affecting the entire length of the colon and rectum. The pathogenesis of UC involves a complex interplay of factors, including abnormalities in the immune system, environmental influences, disturbances in the gut microbiota, exogenous infections, and specific genetic mutations. However, the precise mechanisms underlying its pathogenesis remain incompletely understood. UC typically manifests as a pattern of inflammation spreading from the rectum to the distal colon, eventually encompassing the entire large intestine. In recent years, considerable attention has been given to exploring the role of autophagy in UC. Studies have revealed a potential association between decreased autophagic activity and the onset of UC[7]. Recent findings have highlighted the potential of activating the intestinal nuclear receptor vitamin D receptor (VDR) *via* autophagy. This has

Shao BZ et al. Autophagy in gastrointestinal diseases



**Figure 1 Illustrative diagram elucidating the signaling pathways governing the initiation and regulation of autophagy.** Autophagy, a cellular process, is stimulated under conditions such as nutrient deficiency, starvation, or ischemic stress. The initiation of autophagy involves distinct groups of autophagy-related gene (Atg) proteins. The formation of the Atg1 complex, comprising *ULK1*, focal adhesion kinase family kinase-interacting protein of 200 kDa, *ATG13*, and *ATG101*, triggers the assembly of the class III phosphatidylinositol 3-kinase (PI3K) complex, encompassing beclin-1, *ATG14*, *VSP15*, and *VSP34*, thereby initiating membrane nucleation and phagophore formation. Subsequent membrane expansion and fusion, facilitated by *ATG5-ATG12-ATG16L1* and light chain 3-II, give rise to the autophagosome, which subsequently fuses with a lysosome, resulting in the establishment of a functional autophagy unit. Conversely, under conditions characterized by ample nutrients, growth factors, and insulin, activation of the class I PI3K-Akt- mechanistic target of rapamycin complex 1 (mTORC1) signaling pathway inhibits the formation of the Atg1 complex. While rapamycin serves as an autophagy inducer by inhibiting mTORC1, 3-MA, bafilomycin A1, and chloroquine obstruct the autophagy process through varied mechanisms. PI3K: Phosphatidylinositol 3-kinase.

been demonstrated in several studies[8-10]. Notably, decreased expression of VDR and dysregulated vitamin D/VDR signaling pathways have been observed in patients with UC. Moreover, Zhou *et al*[11] reported mTOR-dependent deficiency in autophagy flux in human intestinal epithelial cells derived from active UC patients. In dextran sulfate sodium (DSS)-induced UC models, Wu *et al*[12] reported the contribution of autophagy to gut microbiota homeostasis in UC models. In line with these findings, recent investigations conducted by our research team have demonstrated that inducing autophagy *via* the adenosine 5′-monophosphate-activated protein kinase-mTOR-p70S6K pathway through the activation of specific receptors alleviates UC symptoms and suppresses intestinal inflammation in DSS-induced mouse models[13]. Collectively, these studies underscore the therapeutic potential of autophagy in ameliorating the onset and progression of UC. Notably, polymorphisms in specific Atgs, including *CASP1*, *SERPINA1*, and *CCL2*, have been linked to an increased risk of UC. These findings identify novel molecular targets for the development of autophagy-modulating therapies for the management of active UC[14].

CD is a distinct entity within the spectrum of IBD. CD manifests as localized or regional enteritis of uncertain etiology and is influenced by a multitude of pathogenetic factors, including compromised immunity, environmental stressors, familial predisposition, and specific genetic mutations. In contrast to UC, CD is more susceptible to genetic influences. Extensive genome-wide association studies conducted in 2007 highlighted the significant association between mutations in certain Atgs, notably ATG16L1 and immunity-related GTPase M (IRGM), and the pathogenesis of CD[15,16]. Moreover, investigations by Rioux et al [17] revealed that polymorphisms in Atgs are related to CD-associated adherentinvasive Escherichia coli replication. Nucleotide binding oligomerization domain-containing protein 2 (NOD2) has emerged as a pivotal inducer of autophagy, orchestrating the recruitment of ATG16L1 to the cell membrane[18]. Notably, mutations in NOD2 observed in CD patients have been linked to disruptions in intestinal microbiota homeostasis and heightened gut inflammatory responses [19,20]. In addition to NOD2, IRGM, ULK-1, and XBP-1, mutations or deletions in other Atgs are highly related to the pathogenesis of IBD, particularly CD, according to recent reviews[21]. Studies utilizing 2,4,6-trinitrobenzenesulfonic acid-induced models have reported that inhibiting autophagy exacerbates CD symptoms and intestinal inflammation, highlighting the protective role of autophagy in CD[22]. Collectively, these findings underscore the specific involvement of autophagy in the onset and progression of CD. On the basis of our insights into the role of autophagy in the pathogenesis and progression of CD, targeting impaired autophagy, particularly that caused by polymorphisms in Atgs, may represent a promising therapeutic approach for CD[23].

Zaishidena® WJG https://www.wjgnet.com

In addition to UC and CD, pancreatitis has emerged as another prevalent inflammation-related gastrointestinal disorder. Pancreatitis, characterized by pancreatic self-digestion primarily mediated by trypsin, manifests as damage to and necrosis of pancreatic tissue due to the overactivation of self-protective inflammatory and immune responses. Extensive evidence underscores the close association between autophagy and pancreatitis. As previously documented, the onset of acute pancreatitis triggers a significant increase in autophagosome formation within pancreatic acinar cells [24]. However, recent findings have revealed elevated levels of both LC3-II and sequestosome 1 in mouse models of pancreatitis[25], suggesting impaired autophagy. Furthermore, multiple investigations have demonstrated a decrease in lysosome formation in cerulein-treated mouse pancreatic acinar cells, as evidenced by the downregulation of lysosomeassociated membrane proteins (LAMP1 and LAMP2), which play crucial roles in stabilizing lysosomal membranes and safeguarding the cytoplasm from acid hydrolases [26,27]. These studies underscore the importance of restoring autophagic function in the management of pancreatitis.

#### AUTOPHAGY IN GASTROINTESTINAL CANCERS

Gastrointestinal cancers encompass a spectrum of malignancies affecting tissues within the gastrointestinal tract, such as the esophagus, stomach, colon, and rectum[28]. Esophageal cancer has been extensively linked with autophagy. A recent study revealed that 22 autophagic long-chain noncoding ribonucleic acids are strongly associated with the survival rate of patients with esophageal adenocarcinoma<sup>[29]</sup>. Research by Fang *et al*<sup>[30]</sup> revealed that the induction of autophagy protected esophageal cancer cells exposed to diketopyrrolopyrrole treatment. Moreover, autophagy is related to microRNA-193b-related chemoresistance to 5-fluorouracil[31]. Furthermore, microRNA-498 has been identified as a suppressor of esophageal cancer through the inhibition of autophagy[32]. These findings underscore the intricate interplay between autophagy and esophageal cancer progression, suggesting potential avenues for therapeutic intervention in this malignancy. In contrast, several studies have presented conflicting findings. A recent investigation demonstrated that silencing autophagy exacerbates radiotherapy resistance in esophageal squamous cell carcinoma[33]. Moreover, inducing autophagy can elicit autophagy-mediated cell cycle arrest[34,35]. We hypothesize that the conflicting outcomes of autophagy in esophageal cancer may stem from variations in the induction methods used and the extent of autophagy activation. Therefore, further research is urgently needed to clarify the exact function of autophagy in esophageal cancer.

Gastric cancer, also known as stomach cancer, is one of the most formidable human malignancies globally and poses a significant health burden [36,37]. Recent investigations have extensively probed the role of autophagy in gastric cancer. Numerous studies suggest that modulating autophagy levels could be a promising therapeutic strategy for gastric cancer treatment. Notably, the interplay between autophagy and programmed cell death (PD)-1 has garnered considerable attention. Wang et al[38] demonstrated that inhibiting autophagy could augment PD-L1 expression, thereby increasing sensitivity to PD-L1-targeted immunotherapy. Moreover, studies have revealed the involvement of unfolded protein response-induced autophagy activation mediated by SEC62, an endoplasmic reticulum membrane protein implicated in protein transport, in promoting gastric cancer metastasis[39]. However, the role of autophagy in the context of Helicobacter pylori (H. pylori) infection is complex. Autophagy has been shown to confer protection against H. pylori infection [40], thereby influencing the incidence of gastric cancer [40,41]. Prolonged exposure to *H. pylori* has been demonstrated to suppress autophagy in gastric epithelial cells[42]. Furthermore, the degradative functions of autophagy induced by vitamin D3 are effective in protecting gastric epithelial cells against *H. pylori* infection[43]. These findings underscore the need to carefully consider the multifaceted and occasionally contradictory effects of autophagy in the treatment of gastric cancer, particularly its influence on *H. pylori* infection.

Colorectal cancer is a prevalent malignant tumor affecting the digestive tract<sup>[21]</sup>. It is the fourth most prevalent malignancy globally and accounts for approximately 11% of all diagnosed malignant tumors worldwide[44]. Extensive research, including investigations conducted by our team and other researchers, has highlighted the role of autophagy in regulating inflammation in diseases[45-47]. Similarly, in colorectal cancer, autophagy has emerged as a significant player in modulating inflammatory and immune processes. Notably, autophagy has metabolic homeostatic roles centered on preserving regulatory Treg cells[48]. Additionally, induction of mitophagy has been shown to stimulate antitumor adaptive immunity[49]. Conversely, a deficiency in autophagy has been associated with tumor progression in colorectal cancer[50,51]. Intriguingly, however, a separate study revealed that the Thr300Ala variation of ATG16L1, a pivotal Atg, was correlated with improved overall survival in human patients with colorectal cancer[52]. These data suggest that the inflammatory and immunoregulatory functions of autophagy may play crucial roles in mitigating tumorigenesis. Additionally, specific mutations in Atgs could offer therapeutic advantages in colorectal cancer.

#### CONCLUSION

This paper provides an in-depth exploration of the role of autophagy in several prominent gastrointestinal disorders, including inflammation-related conditions such as UC, CD, and pancreatitis, as well as gastrointestinal cancers, including esophageal, gastric, and colorectal cancers. Our discussion underscores the intricate and occasionally competitive effects of autophagy on gastrointestinal ailments. The complex and sometimes contradictory roles of autophagy in gastrointestinal diseases may be attributed to its varying effects at different stages of disease development, the differential induction of autophagy, or the intricate nature of gastrointestinal pathogenesis and progression. Despite extensive research in this area, the precise mechanisms through which autophagy operates in gastrointestinal diseases remain



WJG https://www.wjgnet.com

Shao BZ et al. Autophagy in gastrointestinal diseases

elusive. The complexity of the role of autophagy in gastrointestinal diseases has hindered the successful clinical application of autophagy-modulating therapies. Consequently, further studies are essential to fully elucidate the role of autophagy in these diseases and to develop appropriate modulation strategies tailored to different stages of disease progression.

#### ACKNOWLEDGEMENTS

The authors would like to thank Professor Li X for his help in language editing.

#### FOOTNOTES

Author contributions: Shao BZ and Zhang WG retrieved concerned literatures and wrote the manuscript; Liu ZY designed the figure; Linghu EQ revised the manuscript.

Supported by the National Natural Science Foundation of China, No. 82204483.

Conflict-of-interest statement: The authors declare that they have no conflict of interest.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

Country of origin: China

ORCID number: Bo-Zong Shao 0000-0002-2621-0669; Wen-Gang Zhang 0000-0002-3314-0283; En-Qiang Linghu 0000-0003-4506-7877.

S-Editor: Fan M L-Editor: A P-Editor: Cai YX

#### REFERENCES

- Sakai Y, Oku M. ATG and ESCRT control multiple modes of microautophagy. FEBS Lett 2024; 598: 48-58 [PMID: 3785750] DOI: 1 10.1002/1873-3468.14760
- 2 Kumar V, Jurkunas UV. Mitochondrial Dysfunction and Mitophagy in Fuchs Endothelial Corneal Dystrophy. Cells 2021; 10 [PMID: 34440658 DOI: 10.3390/cells10081888]
- Zhu Y, Liu F, Jian F, Rong Y. Recent progresses in the late stages of autophagy. Cell Insight 2024; 3: 100152 [PMID: 38435435 DOI: 3 10.1016/j.cellin.2024.100152]
- Shao BZ, Wang P, Bai Y. Editorial: Autophagy in Inflammation Related Diseases. Front Pharmacol 2022; 13: 912487 [PMID: 35600850 4 DOI: 10.3389/fphar.2022.912487]
- 5 Wang P, Shao BZ, Deng Z, Chen S, Yue Z, Miao CY. Autophagy in ischemic stroke. Prog Neurobiol 2018; 163-164: 98-117 [PMID: 29331396 DOI: 10.1016/j.pneurobio.2018.01.001]
- Chang YF, Li JJ, Liu T, Wei CQ, Ma LW, Nikolenko VN, Chang WL. Morphological and biochemical characteristics associated with 6 autophagy in gastrointestinal diseases. World J Gastroenterol 2024; 30: 1524-1532 [PMID: 38617452 DOI: 10.3748/wjg.v30.i11.1524]
- 7 Hu X, Deng J, Yu T, Chen S, Ge Y, Zhou Z, Guo Y, Ying H, Zhai Q, Chen Y, Yuan F, Niu Y, Shu W, Chen H, Ma C, Liu Z, Guo F. ATF4 Deficiency Promotes Intestinal Inflammation in Mice by Reducing Uptake of Glutamine and Expression of Antimicrobial Peptides. Gastroenterology 2019; 156: 1098-1111 [PMID: 30452920 DOI: 10.1053/j.gastro.2018.11.033]
- 8 Bakke D, Sun J. Ancient Nuclear Receptor VDR With New Functions: Microbiome and Inflammation. Inflamm Bowel Dis 2018; 24: 1149-1154 [PMID: 29718408 DOI: 10.1093/ibd/izy092]
- Law AD, Dutta U, Kochhar R, Vaishnavi C, Kumar S, Noor T, Bhadada S, Singh K. Vitamin D deficiency in adult patients with ulcerative 9 colitis: Prevalence and relationship with disease severity, extent, and duration. Indian J Gastroenterol 2019; 38: 6-14 [PMID: 30864011 DOI: 10.1007/s12664-019-00932-z]
- Karimi S, Tabataba-Vakili S, Yari Z, Alborzi F, Hedayati M, Ebrahimi-Daryani N, Hekmatdoost A. The effects of two vitamin D regimens on 10 ulcerative colitis activity index, quality of life and oxidant/anti-oxidant status. Nutr J 2019; 18: 16 [PMID: 30871542 DOI: 10.1186/s12937-019-0441-71
- Zhou M, Xu W, Wang J, Yan J, Shi Y, Zhang C, Ge W, Wu J, Du P, Chen Y. Boosting mTOR-dependent autophagy via upstream TLR4-11 MyD88-MAPK signalling and downstream NF-kB pathway quenches intestinal inflammation and oxidative stress injury. EBioMedicine 2018; 35: 345-360 [PMID: 30170968 DOI: 10.1016/j.ebiom.2018.08.035]
- Wu MY, Liu L, Wang EJ, Xiao HT, Cai CZ, Wang J, Su H, Wang Y, Tan J, Zhang Z, Wang J, Yao M, Ouyang DF, Yue Z, Li M, Chen Y, 12 Bian ZX, Lu JH. PI3KC3 complex subunit NRBF2 is required for apoptotic cell clearance to restrict intestinal inflammation. Autophagy 2021; 17: 1096-1111 [PMID: 32160108 DOI: 10.1080/15548627.2020.1741332]
- Ke P, Shao BZ, Xu ZQ, Wei W, Han BZ, Chen XW, Su DF, Liu C. Activation of Cannabinoid Receptor 2 Ameliorates DSS-Induced Colitis 13



WJG https://www.wjgnet.com

through Inhibiting NLRP3 Inflammasome in Macrophages. PLoS One 2016; 11: e0155076 [PMID: 27611972 DOI: 10.1371/journal.pone.0155076]

- Qiu P, Liu L, Fang J, Zhang M, Wang H, Peng Y, Chen M, Liu J, Wang F, Zhao Q. Identification of Pharmacological Autophagy Regulators 14 of Active Ulcerative Colitis. Front Pharmacol 2021; 12: 769718 [PMID: 34925026 DOI: 10.3389/fphar.2021.769718]
- Tysk C, Lindberg E, Järnerot G, Flodérus-Myrhed B. Ulcerative colitis and Crohn's disease in an unselected population of monozygotic and 15 dizygotic twins. A study of heritability and the influence of smoking. Gut 1988; 29: 990-996 [PMID: 3396969 DOI: 10.1136/gut.29.7.990]
- Hampe J, Franke A, Rosenstiel P, Till A, Teuber M, Huse K, Albrecht M, Mayr G, De La Vega FM, Briggs J, Günther S, Prescott NJ, Onnie 16 CM, Häsler R, Sipos B, Fölsch UR, Lengauer T, Platzer M, Mathew CG, Krawczak M, Schreiber S. A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1. Nat Genet 2007; 39: 207-211 [PMID: 17200669 DOI: 10.1038/ng1954]
- Rioux JD, Xavier RJ, Taylor KD, Silverberg MS, Goyette P, Huett A, Green T, Kuballa P, Barmada MM, Datta LW, Shugart YY, Griffiths 17 AM, Targan SR, Ippoliti AF, Bernard EJ, Mei L, Nicolae DL, Regueiro M, Schumm LP, Steinhart AH, Rotter JI, Duerr RH, Cho JH, Daly MJ, Brant SR. Genome-wide association study identifies new susceptibility loci for Crohn disease and implicates autophagy in disease pathogenesis. Nat Genet 2007; 39: 596-604 [PMID: 17435756 DOI: 10.1038/ng2032]
- Park SC, Jeen YT. Genetic Studies of Inflammatory Bowel Disease-Focusing on Asian Patients. Cells 2019; 8 [PMID: 31052430 DOI: 18 10.3390/cells8050404]
- 19 Belyayev L, Hawksworth J, Khan K, Kaufman S, Subramanian S, Kroemer A, Loh K, Girlanda R, Fishbein TM, Matsumoto CS. Immunologic Complications and Graft Survival in Crohn's Disease and NOD2 Mutant Non-Crohn's Disease Adult Recipients Following Intestine Transplantation. Transplant Direct 2020; 6: e556 [PMID: 32607422 DOI: 10.1097/TXD.000000000001006]
- Frade-Proud'Hon-Clerc S, Smol T, Frenois F, Sand O, Vaillant E, Dhennin V, Bonnefond A, Froguel P, Fumery M, Guillon-Dellac N, 20 Gower-Rousseau C, Vasseur F. A Novel Rare Missense Variation of the NOD2 Gene: Evidences of Implication in Crohn's Disease. Int J Mol Sci 2019; 20 [PMID: 30769939 DOI: 10.3390/ijms20040835]
- 21 Wang SL, Shao BZ, Zhao SB, Fang J, Gu L, Miao CY, Li ZS, Bai Y. Impact of Paneth Cell Autophagy on Inflammatory Bowel Disease. Front Immunol 2018; 9: 693 [PMID: 29675025 DOI: 10.3389/fimmu.2018.00693]
- Macias-Ceja DC, Cosín-Roger J, Ortiz-Masiá D, Salvador P, Hernández C, Esplugues JV, Calatayud S, Barrachina MD. Stimulation of 22 autophagy prevents intestinal mucosal inflammation and ameliorates murine colitis. Br J Pharmacol 2017; 174: 2501-2511 [PMID: 28500644 DOI: 10.1111/bph.13860]
- 23 Alula KM, Theiss AL. Autophagy in Crohn's Disease: Converging on Dysfunctional Innate Immunity. Cells 2023; 12 [PMID: 37443813 DOI: 10.3390/cells12131779]
- 24 Gukovskaya AS, Gukovsky I, Algül H, Habtezion A. Autophagy, Inflammation, and Immune Dysfunction in the Pathogenesis of Pancreatitis. Gastroenterology 2017; 153: 1212-1226 [PMID: 28918190 DOI: 10.1053/j.gastro.2017.08.071]
- Wang S, Chao X, Jiang X, Wang T, Rodriguez Y, Yang L, Pacher P, Ni HM, Ding WX. Loss of acinar cell VMP1 triggers spontaneous 25 pancreatitis in mice. Autophagy 2022; 18: 1572-1582 [PMID: 34709991 DOI: 10.1080/15548627.2021.1990672]
- Wang S, Ni HM, Chao X, Wang H, Bridges B, Kumer S, Schmitt T, Mareninova O, Gukovskaya A, De Lisle RC, Ballabio A, Pacher P, Ding 26 WX. Impaired TFEB-mediated lysosomal biogenesis promotes the development of pancreatitis in mice and is associated with human pancreatitis. Autophagy 2019; 15: 1954-1969 [PMID: 30894069 DOI: 10.1080/15548627.2019.1596486]
- Zhang T, Gan Y, Zhu S. Association between autophagy and acute pancreatitis. Front Genet 2023; 14: 998035 [PMID: 36793898 DOI: 27 10.3389/fgene.2023.998035]
- Griffin-Sobel JP. Gastrointestinal Cancers: Screening and Early Detection. Semin Oncol Nurs 2017; 33: 165-171 [PMID: 28343837 DOI: 28 10.1016/j.soncn.2017.02.004]
- 29 Wu L, Zheng Y, Ruan X, Wu D, Xu P, Liu J, Wu D, Li X. Long-chain noncoding ribonucleic acids affect the survival and prognosis of patients with esophageal adenocarcinoma through the autophagy pathway: construction of a prognostic model. Anticancer Drugs 2022; 33: e590-e603 [PMID: 34338240 DOI: 10.1097/CAD.00000000001189]
- Fang F, Li Y, Chang L. Mechanism of autophagy regulating chemoresistance in esophageal cancer cells. Exp Mol Pathol 2020; 117: 104564 30 [PMID: 33137292 DOI: 10.1016/j.yexmp.2020.104564]
- Nyhan MJ, O'Donovan TR, Boersma AW, Wiemer EA, McKenna SL. MiR-193b promotes autophagy and non-apoptotic cell death in 31 oesophageal cancer cells. BMC Cancer 2016; 16: 101 [PMID: 26878873 DOI: 10.1186/s12885-016-2123-6]
- 32 Li D, Yan M, Sun F, Song J, Hu X, Yu S, Tang L, Deng S. miR-498 inhibits autophagy and M2-like polarization of tumor-associated macrophages in esophageal cancer via MDM2/ATF3. Epigenomics 2021; 13: 1013-1030 [PMID: 34114479 DOI: 10.2217/epi-2020-0341]
- Yao X, Chen H, Xu B, Lu J, Gu J, Chen F, Ju M, Sun X. The ATPase subunit of ATP6V1C1 inhibits autophagy and enhances radiotherapy 33 resistance in esophageal squamous cell carcinoma. Gene 2021; 768: 145261 [PMID: 33183740 DOI: 10.1016/j.gene.2020.145261]
- 34 Ma Q, Liao H, Xu L, Li Q, Zou J, Sun R, Xiao D, Liu C, Pu W, Cheng J, Zhou X, Huang G, Yao L, Zhong X, Guo X. Autophagy-dependent cell cycle arrest in esophageal cancer cells exposed to dihydroartemisinin. Chin Med 2020; 15: 37 [PMID: 32351616 DOI: 10.1186/s13020-020-00318-w]
- Chen X, He LY, Lai S, He Y. Dihydroartemisinin inhibits the migration of esophageal cancer cells by inducing autophagy. Oncol Lett 2020; 35 20: 94 [PMID: 32831913 DOI: 10.3892/o1.2020.11955]
- 36 Yeoh KG, Tan P. Mapping the genomic diaspora of gastric cancer. Nat Rev Cancer 2022; 22: 71-84 [PMID: 34702982 DOI: 10.1038/s41568-021-00412-7]
- 37 Ajani JA, D'Amico TA, Bentrem DJ, Chao J, Cooke D, Corvera C, Das P, Enzinger PC, Enzler T, Fanta P, Farjah F, Gerdes H, Gibson MK, Hochwald S, Hofstetter WL, Ilson DH, Keswani RN, Kim S, Kleinberg LR, Klempner SJ, Lacy J, Ly QP, Matkowskyj KA, McNamara M, Mulcahy MF, Outlaw D, Park H, Perry KA, Pimiento J, Poultsides GA, Reznik S, Roses RE, Strong VE, Su S, Wang HL, Wiesner G, Willett CG, Yakoub D, Yoon H, McMillian N, Pluchino LA. Gastric Cancer, Version 2.2022, NCCN Clinical Practice Guidelines in Oncology. J Natl Compr Canc Netw 2022; 20: 167-192 [PMID: 35130500 DOI: 10.6004/jnccn.2022.0008]
- Wang X, Wu WKK, Gao J, Li Z, Dong B, Lin X, Li Y, Li Y, Gong J, Qi C, Peng Z, Yu J, Shen L. Autophagy inhibition enhances PD-L1 38 expression in gastric cancer. J Exp Clin Cancer Res 2019; 38: 140 [PMID: 30925913 DOI: 10.1186/s13046-019-1148-5]
- 39 Su S, Shi YT, Chu Y, Jiang MZ, Wu N, Xu B, Zhou H, Lin JC, Jin YR, Li XF, Liang J. Sec62 promotes gastric cancer metastasis through mediating UPR-induced autophagy activation. Cell Mol Life Sci 2022; 79: 133 [PMID: 35165763 DOI: 10.1007/s00018-022-04143-2]
- Raju D, Hussey S, Ang M, Terebiznik MR, Sibony M, Galindo-Mata E, Gupta V, Blanke SR, Delgado A, Romero-Gallo J, Ramjeet MS, 40 Mascarenhas H, Peek RM, Correa P, Streutker C, Hold G, Kunstmann E, Yoshimori T, Silverberg MS, Girardin SE, Philpott DJ, El Omar E,



Jones NL. Vacuolating cytotoxin and variants in Atg16L1 that disrupt autophagy promote Helicobacter pylori infection in humans. Gastroenterology 2012; 142: 1160-1171 [PMID: 22333951 DOI: 10.1053/j.gastro.2012.01.043]

- 41 Muhammad JS, Nanjo S, Ando T, Yamashita S, Maekita T, Ushijima T, Tabuchi Y, Sugiyama T. Autophagy impairment by Helicobacter pylori-induced methylation silencing of MAP1LC3Av1 promotes gastric carcinogenesis. Int J Cancer 2017; 140: 2272-2283 [PMID: 28214334 DOI: 10.1002/ijc.30657]
- He Y, Wang C, Zhang X, Lu X, Xing J, Lv J, Guo M, Huo X, Liu X, Lu J, Du X, Li C, Chen Z. Sustained Exposure to Helicobacter pylori 42 Lysate Inhibits Apoptosis and Autophagy of Gastric Epithelial Cells. Front Oncol 2020; 10: 581364 [PMID: 33194715 DOI: 10.3389/fonc.2020.581364]
- Hu W, Zhang L, Li MX, Shen J, Liu XD, Xiao ZG, Wu DL, Ho IHT, Wu JCY, Cheung CKY, Zhang YC, Lau AHY, Ashktorab H, Smoot DT, 43 Fang EF, Chan MTV, Gin T, Gong W, Wu WKK, Cho CH. Vitamin D3 activates the autolysosomal degradation function against Helicobacter pylori through the PDIA3 receptor in gastric epithelial cells. Autophagy 2019; 15: 707-725 [PMID: 30612517 DOI: 10.1080/15548627.2018.1557835]
- Zhang B, Wang HE, Bai YM, Tsai SJ, Su TP, Chen TJ, Wang YP, Chen MH. Inflammatory bowel disease is associated with higher dementia 44 risk: a nationwide longitudinal study. Gut 2021; 70: 85-91 [PMID: 32576641 DOI: 10.1136/gutjnl-2020-320789]
- Shao BZ, Wang SL, Fang J, Li ZS, Bai Y, Wu K. Alpha7 Nicotinic Acetylcholine Receptor Alleviates Inflammatory Bowel Disease Through 45 Induction of AMPK-mTOR-p7086K-Mediated Autophagy. Inflammation 2019; 42: 1666-1679 [PMID: 31236857 DOI: 10.1007/s10753-019-01027-9
- Xu W, Hua Z, Wang Y, Tang W, Ou W, Liu F, Yang Y, Ding W, Wang Z, Cui L, Ge W, Gu Y, Wang X, Chen Y, Liu CY, Du P. AMBRA1 46 promotes intestinal inflammation by antagonizing PP4R1/PP4c mediated IKK dephosphorylation in an autophagy-independent manner. Cell Death Differ 2024; 31: 618-634 [PMID: 38424148 DOI: 10.1038/s41418-024-01275-9]
- Wu MY, Wang EJ, Ye RD, Lu JH. Enhancement of LC3-associated efferocytosis for the alleviation of intestinal inflammation. Autophagy 47 2024; 20: 1442-1443 [PMID: 38311819 DOI: 10.1080/15548627.2024.2311548]
- Wei J, Long L, Yang K, Guy C, Shrestha S, Chen Z, Wu C, Vogel P, Neale G, Green DR, Chi H. Autophagy enforces functional integrity of 48 regulatory T cells by coupling environmental cues and metabolic homeostasis. Nat Immunol 2016; 17: 277-285 [PMID: 26808230 DOI: 10.1038/ni.3365
- Ziegler PK, Bollrath J, Pallangyo CK, Matsutani T, Canli Ö, De Oliveira T, Diamanti MA, Müller N, Gamrekelashvili J, Putoczki T, Horst D, 49 Mankan AK, Öner MG, Müller S, Müller-Höcker J, Kirchner T, Slotta-Huspenina J, Taketo MM, Reinheckel T, Dröse S, Larner AC, Wels WS, Ernst M, Greten TF, Arkan MC, Korn T, Wirth D, Greten FR. Mitophagy in Intestinal Epithelial Cells Triggers Adaptive Immunity during Tumorigenesis. Cell 2018; 174: 88-101.e16 [PMID: 29909986 DOI: 10.1016/j.cell.2018.05.028]
- 50 Lucas C, Salesse L, Hoang MHT, Bonnet M, Sauvanet P, Larabi A, Godfraind C, Gagnière J, Pezet D, Rosenstiel P, Barnich N, Bonnet R, Dalmasso G, Nguyen HTT. Autophagy of Intestinal Epithelial Cells Inhibits Colorectal Carcinogenesis Induced by Colibactin-Producing Escherichia coli in Apc(Min/+) Mice. Gastroenterology 2020; 158: 1373-1388 [PMID: 31917256 DOI: 10.1053/j.gastro.2019.12.026]
- Liu X, Chen J, Long X, Lan J, Liu X, Zhou M, Zhang S, Zhou J. RSL1D1 promotes the progression of colorectal cancer through RAN-51 mediated autophagy suppression. Cell Death Dis 2022; 13: 43 [PMID: 35013134 DOI: 10.1038/s41419-021-04492-z]
- Grimm WA, Messer JS, Murphy SF, Nero T, Lodolce JP, Weber CR, Logsdon MF, Bartulis S, Sylvester BE, Springer A, Dougherty U, 52 Niewold TB, Kupfer SS, Ellis N, Huo D, Bissonnette M, Boone DL. The Thr300Ala variant in ATG16L1 is associated with improved survival in human colorectal cancer and enhanced production of type I interferon. Gut 2016; 65: 456-464 [PMID: 25645662 DOI: 10.1136/gutjnl-2014-308735]



WJG https://www.wjgnet.com



### Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: office@baishideng.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

