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Editorial Board Member of World Journal of Gastroenterology, Gulsum Ozlem Elpek, MD, Professor, Department of Pathology, Akdeniz University Medical School, Antalya 07070, Türkiye. elpek@akdeniz.edu.tr

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EDITORIAL

## New challenges in hepatocellular carcinoma: A role for PIWIinteracting RNAs?

Domenico Tierno, Gabriele Grassi, Bruna Scaggiante

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Domenico Tierno, Gabriele Grassi, Department of Medicine, Surgery and Health Sciences, University Hospital of Cattinara, University of Trieste, Trieste 34149, Italy

Bruna Scaggiante, Department of Life Sciences, University of Trieste, Trieste 34127, Italy

Corresponding author: Bruna Scaggiante, PhD, Department of Life Sciences, University of Trieste, Via Valerio 28, Trieste 34127, Italy. bscaggiante@units.it

#### Abstract

Hepatocellular carcinoma (HCC) is the most common and deadliest subtype of liver cancer worldwide and, therefore, poses an enormous threat to global health. Understanding the molecular mechanisms underlying the development and progression of HCC is central to improving our clinical approaches. PIWIinteracting RNAs (piRNAs) are a class of small non-coding RNAs that bind to PIWI family proteins to regulate gene expression at transcriptional and posttranscriptional levels. A growing body of work shows that the dysregulation of piRNAs plays a crucial role in the progression of various human cancers. In this editorial, we report on the current knowledge of HCC-associated piRNAs and their potential clinical utility. Based on the editorial by Papadopoulos and Trifylli, on the role and clinical evaluation of exosomal circular RNAs in HCC, we highlight this other emerging class of non-coding RNAs.

**Key Words:** Biomarker; Hepatocellular carcinoma; Liquid biopsy; Non-coding RNA; PIWI-interacting RNA; Next-generation sequencing

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Core Tip: Hepatocellular carcinoma (HCC) is a worldwide clinical problem. Over the past decade, several papers have suggested that specific PIWI-interacting RNAs (piRNAs) may be useful as efficient HCC biomarkers at both tissue and serum levels. Interestingly, the piRNA expression profile changes dynamically over the course of the pathological stage from liver fibrosis to HCC development and progression. Knowledge of piRNAs may improve our understanding of HCC and open new clinical perspectives.

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#### INTRODUCTION

Hepatocellular carcinoma (HCC) accounts for 80%-90% of all liver tumors and its incidence has increased in recent decades[1,2]. The risk factors are numerous, including hepatitis B or C infection, alcoholic and non-alcoholic fatty liver disease, and diabetes. In general, these pathological conditions lead to liver fibrosis, which develops into cirrhosis and eventually HCC. This tumor is extremely heterogeneous and aggressive, has a high recurrence rate and is often resistant to chemotherapy. For this reason, survival rates for HCC are poor despite improvements in diagnostic and therapeutic approaches[3]. Understanding the molecular networks underlying the development and progression of HCC is important for new biomarkers and therapeutic goals.

Non-coding RNAs (ncRNAs) account for 97% of total transcriptional output. Advancing technologies in the enrichment and sequencing of ncRNAs shed light on their active role in the regulation of biological processes and made them increasingly attractive. The ncRNAs can be classified according to their function, length and shape[4]. The most important classes are: (1) MicroRNAs (miRNAs): Small ncRNAs with a length of about 20 nucleotides (nt). Their primary function is the inhibition of mRNA translation by binding sequences with incomplete complementarity in the 3'untranslated regions of the target mRNAs[5]; (2) PIWI-interacting RNAs (piRNAs): Small ncRNAs (mainly 24-35 nt long) [6,7], mainly found in germline cells. As the name suggests, they interact with proteins belonging to the PIWI family and regulate gene expression at transcriptional (through epigenetic chromatin modification) and post-transcriptional level (through RNA and protein interactions)[8]; (3) Long ncRNAs (lncRNAs): NcRNAs generated from pre-mRNAs. They are more than 200 nt long and are characterized by complex secondary/tertiary structures designated for interactions with their molecular targets (proteins or DNA/RNA). LncRNAs regulate many biological processes by acting as miRNA sponges, as recruiters of transcription factors to target promoters and as scaffolding meant to facilitate protein interactions[9]; and (4) Circular RNAs (circRNAs): NcRNAs which are similar in length and function to lncRNAs, except for their distinct ring-like shape. CircRNAs are formed by the circularization of exons, introns, or other ncRNAs by a noncanonical splicing event, so-called "back-splicing" [10].

Since ncRNAs play a crucial role in various biological processes, it is not surprising that evidence of their massive involvement in cancer is accumulating[11,12]. Accordingly, studying the biogenesis and functions of ncRNAs could be a valuable tool in deepening our knowledge of cancer. The editorial by Papadopoulos and Trifylli[13] in the World Journal of Gastroenterology discussed the role of exosomal circRNAs in HCC and proposed them as new therapeutic targets. In this editorial, we will extend the discussion to piRNAs, another class of ncRNAs, and their potential utility for the clinical treatment of HCC.

#### **PIRNA**

A brief note on the biology of piRNA: These ncRNAs are transcribed from specific loci, enriched with transposons, the socalled "piRNA clusters", which can be divided into single-stranded (ss) and double-stranded (ds) clusters. The ss-cluster generates a piRNA precursor with a 3'-polyadenylated tail and a 5'-methylguanosine-cap, similar to the canonical transcription of mRNAs. The ds-cluster can instead generate two non-polyadenylated piRNA precursors (sense and antisense) by means of a protein complex, including RNA polymerase II. Regardless of their origin, the piRNA precursors are exported to the cytoplasm, in particular to the outer membrane of the mitochondria, where the proteins involved in their maturation are located. The piRNA precursor is first cleaved at the 5'-end by an endonuclease called Zucchini to form a 5'-monophosphorylated pre-piRNA, which, in turn, is loaded into a PIWI protein (in a 1:1 ratio) and further cleaved at the 3'-end by the 3'-to-5' exonuclease Nibbler. The new, truncated 3'-terminal end is finally methylated at the 2'-oxygen by Hen1, a 2'-O-methyltransferase, presumably to increase the stability of the piRNA. These primary mature piRNAs can trigger a secondary piRNA maturation cycle, called "ping-pong cycle", in order to exponentially increase the mature cellular piRNA pool. Basically, a mature anti-sense piRNA is loaded into the Aub protein to cleave the sense piRNA precursor into multiple sense piRNA precursors, which are fully processed and then finally loaded into the Ago3 protein. The mature sense piRNA-Ago3 complex cleaves the anti-sense piRNA precursor into multiple anti-sense piRNA precursors, which are loaded into Aub proteins to restart the cycle[14] (Figure 1). It is important to emphasise that our knowledge of the biogenesis of piRNA is still incomplete, and comes mainly from Drosophila melanogaster and micerelated studies. Despite the high degree of evolutionary conservation of this pathway, a deeper understanding of piRNA biogenesis, especially in humans, is crucial to assessing its importance for basic and translational research.

Functionally, the main role of piRNAs is to silence gene expression at the transcriptional level by recruiting DNA methyltransferase and histone methyltransferase to the target promoter and target histone, respectively. In addition, piRNAs protect the cells from transposons and prevent their amplification and mobilisation. The piRNA-mediated regulation of gene expression can also be observed at a post-transcriptional level: piRNA can inhibit mRNA translation and trigger the degradation of pseudogenic transcripts and lncRNAs by interacting with complementary sequences at the

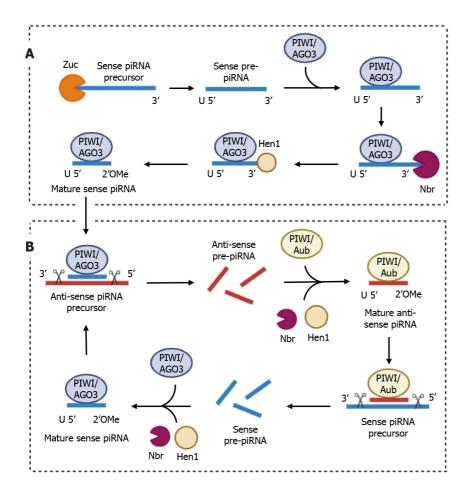


Figure 1 Schematic diagrams of the biosynthesis of PIWI-interacting RNAs. A and B: The primary (A) and secondary (B) cycles (also known as the "ping-pong cycle") are shown. Note that the PIWI-interacting RNA (piRNA) precursor that triggers primary biosynthesis can be either a sense or an anti-sense piRNA precursor. Image created by Biorender. Zuc: Zucchini; piRNA: PIWI-interacting RNA; Nbr: Nibbler.

5'-end. Finally, it has been reported that many piRNAs can act as scaffolding to facilitate interaction with multiple proteins, similar to circRNAs and lncRNAs[15]. In recent decades, increasing number of studies have reported the possible involvement of piRNA dysregulation in the development and progression of cancer. We provide a brief chronological overview of articles dealing with HCC-associated piRNAs.

#### **PIRNA IN HCC**

According to PubMed research, the paper by Law et al [16] is the first article to evaluate the clinical benefit of piRNAs in HCC. Using Illumina sequencing, they identified a novel piRNA, called piR-Hep1, which was significantly overexpressed in the tumour tissue of 73 HCC patients compared to adjacent normal tissue. Furthermore, inhibition of piR-Hep1 in HCC cell lines (HKCI-4 and HKCI-8) led to a reduction in cell viability, invasiveness, and phosphorylation levels of AKT, indicating a possible involvement of piR-Hep1 in the regulation of the AKT signalling pathway [16].

In 2016, Rizzo et al [17] used next-generation sequencing techniques to characterise the piRNA expression profile of the different pathological stages underlying the diagnosis and progression of HCC: (1) Cirrhotic nodules (CN); (2) Low-grade dysplastic nodules (LGDN); (3) High-grade dysplastic nodules (HGDN); (4) Early HCC (eHCC); and (5) Progressed HCC (pHCC). For this purpose, they obtained resection samples from 17 HCC patients with multiple nodules: (1) 17 CN; (2) 9 LGDN; (3) 6 HGDN; (4) 6 eHCC; and (5) 23 pHCC. Sequencing revealed a 125 piRNA expression signature specific for eHCC and pHCC, and a 24 piRNA expression signature specific for LDGN and HDGN[17]. Similar results were also obtained in 2018 by Koduru et al[18] through database screening and informatic analysis. The RNA-seq datasets analysed were from the NIH Short Read Archive and were from 14 CN, 9 LDGN, 6 HDGN, 6 eHCC, and 20 pHCC samples. They found a specific piRNA expression profile for each HCC-associated pathological stage (number of specific dysregulated piRNAs in round brackets): (1) CN (75); (2) LDGN (60); (3) HGDGN (49); (4) eHCC (56); and (5) pHCC (128). These results indicate dynamic changes in piRNoma during disease progression, which complicates their use as clinical tools.

In 2018, Tang et al[19] also investigated the role of piRNAs in those liver pathologies which will lead to the development of HCC in mice. They found that piR-823 was highly expressed in primary hepatic stellate cells (HSCs) and that its inhibition suppressed the activation of HSCs. In contrast, overexpression of piR-823 increased the proliferation of HSCs and the production of alfa-smooth muscle actin, collagen type I alpha 1, and transforming growth factor beta 1 (crucial genes for liver fibrosis progression). Accordingly, piR-823 could be an early therapeutic target to prevent liver

fibrosis and, subsequently, the progression of HCC.

In 2023, Wu et al [20] found that piR-017724 was significantly downregulated in 45 HCC tissues compared to adjacent normal tissues and that its downregulation was correlated with poor prognosis and advanced tumour stage. The oncosuppressive role of this piRNA was confirmed in HCC cell lines (SMMC-7721 and PLC/PRF/5), where silencing of piR-017724 resulted in the inhibition of cell proliferation and invasiveness but not apoptosis. Functional analyses suggested that piR-017724 could inhibit the expression of PLIN3, a member of the abdominal lipoprotein family involved in lipid droplet homeostasis, and plays a crucial role in the regulation of gene expression, protein degradation, signalling and membrane trafficking, particularly in the liver[20].

Rui et al[21] conducted a comprehensive characterisation of serum exosome piRNA levels in 125 HCC patients and 44 healthy controls in 2023, to assess their suitability as diagnostic biomarkers. They found 253 dysregulated piRNA in exosomes of tumour patients, compared to healthy controls. Then, Rui et al[21] selected the five most upregulated piRNA in HCC exosomes (piR-1029, piR-15254, novel-piR-35395, novel-piR-32132, and novel-piR-43597) and validated them in two different cohorts, confirming their overexpression in the tumour and, thus, their diagnostic potential.

#### CONCLUSION

As with the other classes of ncRNAs, the discovery of disease-associated piRNAs is highly dependent on building the pipeline of identification methods. Nowadays, piRNA identification methods include two approaches: Experimental and computational. The first method is based on common techniques for the isolation of protein-associated RNAs, such as RNA immunoprecipitation sequencing or cross-linking immunoprecipitation and high-throughput sequencing. However, these techniques are expensive, time-consuming, and not very sensitive for low expressed piRNAs. The computational approach to piRNA identification has attracted much attention in recent years as piRNA data has increased significantly. Essentially, it involves the application of various bioinformatics tools to specific piRNA databases [such as: (1) piRBase; (2) piRNAclusterDB; or (3) piRNAtarget] to identify novel piRNAs (e.g., 2L-piRNA), piRNA clusters (e.g. protract), piRNA targets (e.g. miRanda), and disease-associated piRNAs (e.g., WGCNA)[22]. Despite the promising performance, these computational methods have several issues that need to be addressed, such as standardising piRNA nomenclature, developing a gold standard for bioinformatics analysis pipeline and improving the quality and quantity of data in the current piRNA databases (especially regarding the data on healthy controls)[23]. Nevertheless, more and more studies show the involvement of piRNAs in the development and progression of pathological diseases in humans. It is expected that there will be a corresponding increase in high-throughput technologies for piRNA studies in the coming years.

In HCC, research into piRNAs is still in its infancy, and much still needs to be clarified about their involvement in tumour progression. The importance of piRNA mechanisms in HCC is also emphasised by the involvement of PIWI proteins. In 2015, Xie et al [24] demonstrated that Hiwi (or PIWIL1), a member of the PIWI family, is overexpressed at both mRNA and protein levels in HCC cell lines (MHCC97L and MHCC97H) compared to normal liver cell lines (L02), and in tumour tissues from 60 HCC patients compared to normal controls from 48 peritumour samples. Functional analysis showed that the silencing of Hiwi in HCC cell lines led to a reduction in cell proliferation and invasiveness. These results suggest that Hiwi is a potentially useful HCC biomarker and a valuable new therapeutic target [24]. Recently, Hammad et al[25] investigated the mRNA expression levels of PIWI family members [(1) PIWIL1; (2) PIWIL2; (3) PIWIL3; and (4) PIWIL4] in both tissues and serum of HCC patients. Their results showed significant overexpression of these mRNAs in 50 tumour tissues compared to adjacent normal tissues and in 50 HCC sera compared to those of 25 healthy controls. Furthermore, a significant correlation was found between PIWIL1-4 overexpression and advanced tumour stage, at least as far as the tissue samples are concerned[25]. With high-throughput technologies, it is now possible to extend the investigations to a large number of patients, as well as to deepen the molecular mechanisms in cellular models. All in all, these results suggest that piRNAs are attractive candidates in HCC research with the aim of expanding our knowledge of HCC development and progression, as well as opening up new potential applications in the screening and treatment of patients.

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Country of origin: Italy

ORCID number: Bruna Scaggiante 0000-0002-8662-138X.

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#### **REFERENCES**

- Foglia B, Turato C, Cannito S. Hepatocellular Carcinoma: Latest Research in Pathogenesis, Detection and Treatment. Int J Mol Sci 2023; 24 [PMID: 37569600 DOI: 10.3390/ijms241512224]
- Alawyia B, Constantinou C. Hepatocellular Carcinoma: a Narrative Review on Current Knowledge and Future Prospects. Curr Treat Options Oncol 2023; 24: 711-724 [PMID: 37103744 DOI: 10.1007/s11864-023-01098-9]
- Jesenko T, Brezar SK, Cemazar M, Biasin A, Tierno D, Scaggiante B, Grassi M, Grassi C, Dapas B, Truong NH, Abrami M, Zanconati F, Bonazza D, Rizzolio F, Parisi S, Pastorin G, Grassi G. Targeting Non-Coding RNAs for the Development of Novel Hepatocellular Carcinoma Therapeutic Approaches. Pharmaceutics 2023; 15 [PMID: 37111734 DOI: 10.3390/pharmaceutics15041249]
- Yan H, Bu P. Non-coding RNA in cancer. Essays Biochem 2021; 65: 625-639 [PMID: 33860799 DOI: 10.1042/EBC20200032] 4
- Shang R, Lee S, Senavirathne G, Lai EC. microRNAs in action: biogenesis, function and regulation. Nat Rev Genet 2023; 24: 816-833 [PMID: 37380761 DOI: 10.1038/s41576-023-00611-y]
- Hirakata S, Siomi MC. piRNA biogenesis in the germline: From transcription of piRNA genomic sources to piRNA maturation. Biochim 6 Biophys Acta 2016; **1859**: 82-92 [PMID: 26348412 DOI: 10.1016/j.bbagrm.2015.09.002]
- Sun YH, Lee B, Li XZ. The birth of piRNAs: how mammalian piRNAs are produced, originated, and evolved. Mamm Genome 2022; 33: 293-311 [PMID: 34724117 DOI: 10.1007/s00335-021-09927-8]
- Zeng Q, Wan H, Zhao S, Xu H, Tang T, Oware KA, Qu S. Role of PIWI-interacting RNAs on cell survival: Proliferation, apoptosis, and cycle. 8 IUBMB Life 2020; 72: 1870-1878 [PMID: 32697419 DOI: 10.1002/iub.2332]
- Bridges MC, Daulagala AC, Kourtidis A. LNCcation: lncRNA localization and function. J Cell Biol 2021; 220 [PMID: 33464299 DOI: 9 10.1083/jcb.202009045]
- 10 Liu CX, Chen LL. Circular RNAs: Characterization, cellular roles, and applications. Cell 2022; 185: 2016-2034 [PMID: 35584701 DOI: 10.1016/j.cell.2022.04.021]
- Liu Y, Dou M, Song X, Dong Y, Liu S, Liu H, Tao J, Li W, Yin X, Xu W. The emerging role of the piRNA/piwi complex in cancer. Mol 11 Cancer 2019; 18: 123 [PMID: 31399034 DOI: 10.1186/s12943-019-1052-9]
- Tonon F, Cemazar M, Kamensek U, Zennaro C, Pozzato G, Caserta S, Ascione F, Grassi M, Guido S, Ferrari C, Cansolino L, Trotta F, 12 Kuzmanov BG, Forte G, Martino F, Perrone F, Bomben R, Gattei V, Elvassore N, Murano E, Truong NH, Olson M, Farra R, Grassi G, Dapas B. 5-Azacytidine Downregulates the Proliferation and Migration of Hepatocellular Carcinoma Cells In Vitro and In Vivo by Targeting miR-139-5p/ROCK2 Pathway. Cancers (Basel) 2022; 14 [PMID: 35406401 DOI: 10.3390/cancers14071630]
- Papadopoulos N, Trifylli EM. Role of exosomal circular RNAs as microRNA sponges and potential targeting for suppressing hepatocellular 13 carcinoma growth and progression. World J Gastroenterol 2024; 30: 994-998 [PMID: 38577187 DOI: 10.3748/wjg.v30.i9.994]
- Zhang J, Chen S, Liu K. Structural insights into piRNA biogenesis. Biochim Biophys Acta Gene Regul Mech 2022; 1865: 194799 [PMID: 14 35182819 DOI: 10.1016/j.bbagrm.2022.194799]
- Wu X, Pan Y, Fang Y, Zhang J, Xie M, Yang F, Yu T, Ma P, Li W, Shu Y. The Biogenesis and Functions of piRNAs in Human Diseases. Mol 15 Ther Nucleic Acids 2020; 21: 108-120 [PMID: 32516734 DOI: 10.1016/j.omtn.2020.05.023]
- Law PT, Qin H, Ching AK, Lai KP, Co NN, He M, Lung RW, Chan AW, Chan TF, Wong N. Deep sequencing of small RNA transcriptome reveals novel non-coding RNAs in hepatocellular carcinoma. J Hepatol 2013; 58: 1165-1173 [PMID: 23376363 DOI: 10.1016/j.jhep.2013.01.032
- Rizzo F, Rinaldi A, Marchese G, Coviello E, Sellitto A, Cordella A, Giurato G, Nassa G, Ravo M, Tarallo R, Milanesi L, Destro A, Torzilli G, 17 Roncalli M, Di Tommaso L, Weisz A. Specific patterns of PIWI-interacting small noncoding RNA expression in dysplastic liver nodules and hepatocellular carcinoma. Oncotarget 2016; 7: 54650-54661 [PMID: 27429044 DOI: 10.18632/oncotarget.10567]
- Koduru SV, Leberfinger AN, Kawasawa YI, Mahajan M, Gusani NJ, Sanyal AJ, Ravnic DJ. Non-coding RNAs in Various Stages of Liver 18 Disease Leading to Hepatocellular Carcinoma: Differential Expression of miRNAs, piRNAs, lncRNAs, circRNAs, and sno/mt-RNAs. Sci Rep 2018; 8: 7967 [PMID: 29789629 DOI: 10.1038/s41598-018-26360-1]
- Tang X, Xie X, Wang X, Wang Y, Jiang X, Jiang H. The Combination of piR-823 and Eukaryotic Initiation Factor 3 B (EIF3B) Activates 19 Hepatic Stellate Cells via Upregulating TGF-β1 in Liver Fibrogenesis. Med Sci Monit 2018; 24: 9151-9165 [PMID: 30556540 DOI: 10.12659/MSM.914222]
- Wu YJ, Wang J, Zhang P, Yuan LX, Ju LL, Wang HX, Chen L, Cao YL, Cai WH, Ni Y, Li M. PIWIL1 interacting RNA piR-017724 inhibits 20 proliferation, invasion, and migration, and inhibits the development of HCC by silencing PLIN3. Front Oncol 2023; 13: 1203821 [PMID: 37503320 DOI: 10.3389/fonc.2023.1203821]
- Rui T, Wang K, Xiang A, Guo J, Tang N, Jin X, Lin Y, Liu J, Zhang X. Serum Exosome-Derived piRNAs Could Be Promising Biomarkers for 21 HCC Diagnosis. Int J Nanomedicine 2023; 18: 1989-2001 [PMID: 37077942 DOI: 10.2147/IJN.S398462]
- Liu Y, Li A, Xie G, Liu G, Hei X. Computational Methods and Online Resources for Identification of piRNA-Related Molecules. Interdiscip 22 Sci 2021; **13**: 176-191 [PMID: 33886096 DOI: 10.1007/s12539-021-00428-5]
- Guo C, Wang X, Ren H. Databases and computational methods for the identification of piRNA-related molecules: A survey. Comput Struct 23 Biotechnol J 2024; 23: 813-833 [PMID: 38328006 DOI: 10.1016/j.csbj.2024.01.011]



- Xie Y, Yang Y, Ji D, Zhang D, Yao X, Zhang X. Hiwi downregulation, mediated by shRNA, reduces the proliferation and migration of human hepatocellular carcinoma cells. Mol Med Rep 2015; 11: 1455-1461 [PMID: 25370791 DOI: 10.3892/mmr.2014.2847]
- Hammad G, Mamdouh S, Seoudi DM, Seleem MI, Safwat G, Mohamed RH. Elevated expression patterns of P-element Induced Wimpy Testis 25 (PIWI) transcripts are potential candidate markers for Hepatocellular Carcinoma. Cancer Biomark 2024; 39: 95-111 [PMID: 38043006 DOI: 10.3233/CBM-230134]



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