World Journal of *Gastroenterology*

World J Gastroenterol 2020 July 28; 26(28): 3998-4181





Published by Baishideng Publishing Group Inc

WJG

World Journal of VVoria jon. Gastroenterology

Contents

Weekly Volume 26 Number 28 July 28, 2020

REVIEW

3998	Secondary causes of inflammatory bowel diseases	
	Ghouri YA, Tahan V, Shen B	
4018	Clinical considerations in the management of non-alcoholic steatohepatitis cirrhosis pre- and post- transplant: A multi-system challenge	
	Steggerda JA, Mahendraraj K, Todo T, Noureddin M	
4036	Pancreatic neuroendocrine tumors: Therapeutic challenges and research limitations	
	Mpilla GB, Philip PA, El-Rayes B, Azmi AS	

4055 Differential regulation of JAK/STAT-signaling in patients with ulcerative colitis and Crohn's disease Cordes F, Foell D, Ding JN, Varga G, Bettenworth D

MINIREVIEWS

4076 Helicobacter pylori infection: Beyond gastric manifestations

> Santos MLC, de Brito BB, da Silva FAF, Sampaio MM, Marques HS, Oliveira e Silva N, de Magalhães Queiroz DM, de Melo FF

ORIGINAL ARTICLE

Basic Study

4094 Celecoxib attenuates hepatocyte apoptosis by inhibiting endoplasmic reticulum stress in thioacetamideinduced cirrhotic rats

Su W, Tai Y, Tang SH, Ye YT, Zhao C, Gao JH, Tuo BG, Tang CW

Case Control Study

4108 Food groups, diet quality and colorectal cancer risk in the Basque Country

> Alegria-Lertxundi I, Aguirre C, Bujanda L, Fernández FJ, Polo F, Ordovás JM, Etxezarraga MC, Zabalza I, Larzabal M, Portillo I, de Pancorbo MM, Garcia-Etxebarria K, Rocandio AM, Arroyo-Izaga M

Retrospective Study

4126 Primary sclerosing cholangitis associated colitis: Characterization of clinical, histologic features, and their associations with liver transplantation

Aranake-Chrisinger J, Dassopoulos T, Yan Y, Nalbantoglu I

4140 Insulin receptor substrate 1 may play divergent roles in human colorectal cancer development and progression

Lomperta K, Jakubowska K, Grudzinska M, Kanczuga-Koda L, Wincewicz A, Surmacz E, Sulkowski S, Koda M



Contents

World Journal of Gastroenterology

Weekly Volume 26 Number 28 July 28, 2020

4151 Enhancement parameters of contrast-enhanced computed tomography for pancreatic ductal adenocarcinoma: Correlation with pathologic grading

Seo W, Kim YC, Min SJ, Lee SM

Observational Study

4159 Detection of reflux-symptom association in children with esophageal atresia by video-pH-impedance study

Maholarnkij S, Sanpavat A, Decharun K, Dumrisilp T, Tubjareon C, Kanghom B, Patcharatrakul T, Chaijitraruch N, Chongsrisawat V, Sintusek P

Randomized Controlled Trial

4170 Epigastric pain syndrome: What can traditional Chinese medicine do? A randomized controlled trial of **Biling Weitong Granules**

Wen YD, Lu F, Zhao YP, Wang P, Yang Q, Li JX, Li HZ, Chi LL, Zhou ZH, Tang YP, Xu JK, Zhao Y, Tang XD



Contents

Weekly Volume 26 Number 28 July 28, 2020

ABOUT COVER

Editorial board member of World Journal of Gastroenterology, Dr. Osamu Toyoshima is a Director of Toyoshima Endoscopy Clinic in Tokyo, Japan. Dr. Toyoshima graduated from the University of Tokyo with his master's degree in Medicine. After graduating, he joined the Department of Gastroenterology and Surgical Oncology at the University of Tokyo Hospital and engaged in clinical practice and medical research. After that, he established the Toyoshima Endoscopy Clinic with his father, Dr. Hiroshi Toyoshima. Toyoshima Endoscopy Clinic is an endoscopy-specialized clinic, which performs 10000 endoscopies annually. Dr. Osamu Toyoshima mainly conducts research using clinical data from Toyoshima Endoscopy Clinic. He is an expert in the field of gastroenterology, especially of gastric cancer risk evaluation based on the endoscopic gastritis and of quality indicators of colonoscopy such as colorectal polyp detection.

AIMS AND SCOPE

The primary aim of World Journal of Gastroenterology (WJG, World J Gastroenterol) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

INDEXING/ABSTRACTING

The WJG is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2020 edition of Journal Citation Report® cites the 2019 impact factor (IF) for WJG as 3.665; IF without journal self cites: 3.534; 5-year IF: 4.048; Ranking: 35 among 88 journals in gastroenterology and hepatology; and Quartile category: Q2.

RESPONSIBLE EDITORS FOR THIS ISSUE

Electronic Editor: Yan-Liang Zhang, Production Department Director: Yun-Xiaojian Wu; Editorial Office Director: Ze-Mao Gong,

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS
World Journal of Gastroenterology	https://www.wjgnet.com/bpg/gerinfo/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 1007-9327 (print) ISSN 2219-2840 (online)	https://www.wjgnet.com/bpg/GerInfo/287
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
October 1, 1995	https://www.wjgnet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Weekly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Andrzej S Tarnawski, Subrata Ghosh	https://www.wjgnet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
http://www.wjgnet.com/1007-9327/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
July 28, 2020	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2020 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2020 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



WJG

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2020 July 28; 26(28): 3998-4017

DOI: 10.3748/wjg.v26.i28.3998

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

REVIEW

Secondary causes of inflammatory bowel diseases

Yezaz A Ghouri, Veysel Tahan, Bo Shen

ORCID number: Yezaz A Ghouri 0000-0002-8677-1871; Veysel Tahan 0000-0001-6796-9359; Bo Shen 0000-0002-7229-4840.

Author contributions: Ghouri YA review of scientific literature, writing of the manuscript and designing the table; Tahan V review of scientific literature and editing of the manuscript; Shen B review of scientific literature and editing of the manuscript.

Conflict-of-interest statement:

Authors declare no conflict of interests for this article.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/licenses /by-nc/4.0/

Manuscript source: Invited manuscript

Received: April 8, 2020 Peer-review started: April 8, 2020 First decision: April 30, 2020

Yezaz A Ghouri, Veysel Tahan, Department of Internal Medicine, Division of Gastroenterology and Hepatology, University of Missouri- School of Medicine, Columbia, MO 65201, United States

Bo Shen, Department of Medicine and Surgery, Interventional IBD Center, Columbia University Irving Medical Center/New York Presbyterian Hospital, New York, NY 10032, United States

Corresponding author: Bo Shen, MD, Professor of the Edelman-Jarislowsky Surgical Sciences, Medicine and Surgical Sciences, Columbia University Irving Medical Center/New York Presbyterian Hospital, 161 Ft Washington Avenue, Herbert Irving Pavilion Rm 843, New York, NY 10032, United States. bs3270@.columbia.edu

Abstract

Inflammatory bowel diseases (IBD), conventionally consist of Crohn's disease (CD) and ulcerative colitis. They occur in individuals with high risk genotype for the disease in the setting of appropriate environmental factors. The pathogenesis of IBD involves a dysregulated autoimmune response to gut dysbiosis, which in turn is triggered due to exposure to various inciting environmental factors. But there is no clearly defined etiology of IBD and this type of disease is termed as "idiopathic IBD", "classic IBD", or "primary IBD". We reviewed the current medical literature and found that certain etiological factors may be responsible for the development of IBD or IBD-like conditions, and we consider this form of de novo IBD as "secondary IBD". Currently known factors that are potentially responsible for giving rise to secondary IBD are medications; bowel altering surgeries and transplantation of organs, stem cells or fecal microbiome. Medications associated with the development of secondary IBD include; immunomodulators, anti-tumor necrosis factor alpha agents, anti-interleukin agents, interferons, immune stimulating agents and checkpoint inhibitors. Colectomy can in some cases give rise to de novo CD, pouchitis of the ileal pouch, or postcolectomy enteritis syndrome. After solid organ transplantation or hematopoietic stem cell transplantation, the recipient may develop de novo IBD or IBD flare. Fecal microbiota transplantation has been widely used to treat patients suffering from recurrent Clostridium difficile infection but can also causes IBD flares.

Key words: *De novo* inflammatory bowel disease; Secondary inflammatory bowel disease; Inflammatory bowel disease; Crohn's disease; Ulcerative colitis



WJG | https://www.wjgnet.com

Revised: May 15, 2020 Accepted: July 16, 2020 Article in press: July 16, 2020 Published online: July 28, 2020

P-Reviewer: Maehata Y S-Editor: Zhang L L-Editor: A E-Editor: Zhang YL



©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Inflammatory bowel diseases (IBD) are chronic illnesses of the gastrointestinal tract with no clearly defined etiology and are traditionally termed as primary IBD. It is generally believed that IBD results from abnormal immune response to dysbiosis of gut microbiota in a genetically susceptible individual. IBD or IBD-like conditions may also be caused by well-defined etiologies; such as medical, surgical, and organ transplantation. These conditions are coined as secondary IBD. In this review we attempted to highlight some etiological factors, pathogenetic pathways, and clinical features of secondary IBD.

Citation: Ghouri YA, Tahan V, Shen B. Secondary causes of inflammatory bowel diseases. World J Gastroenterol 2020; 26(28): 3998-4017 URL: https://www.wjgnet.com/1007-9327/full/v26/i28/3998.htm DOI: https://dx.doi.org/10.3748/wjg.v26.i28.3998

INTRODUCTION

Inflammatory bowel diseases (IBD) are idiopathic chronic diseases of the gastrointestinal (GI) tract that are traditionally divided into ulcerative colitis (UC) and Crohn's disease (CD) based on their respective phenotypic presentation. Sometimes there is an overlap in clinical presentation, colonoscopic findings and histopathological features between UC and CD; which is termed as indeterminate colitis^[1,2]. UC is the most predominant type of IBD with a prevalence of 7.6 to 246.0 cases per 100000 per year, followed by CD which has a prevalence of 3.6 to 214.0 cases per 100000 per year^[3]. The worldwide distribution of IBD is skewed towards being more prominent in North America and Europe, although in the past two decades its prevalence has risen in developing countries like China and India^[4,5]. This change in trends has paralleled with changes in dietary habits like inclusion of processed foods, increased intake of sugars and fats, overutilization of antibiotics and an overall improvement in hygiene.

The diagnosis of IBD is made by correlating clinical presentation, endoscopic findings and histopathological features of diseased tissue specimens. There is no single test to diagnose IBD or to distinguish between the CD and UC, although use of perinuclear anti-neutrophil cytoplasmic antibody and anti-saccharomyces cerevisiae antibody titers can sometimes be helpful in distinguishing the two^[6]. Gut inflammation in UC is limited to the mucosal layer (epithelium, lamina propria and muscularis mucosa) and may extend up to the superficial submucosa. On the other hand, CD is characterized by the presence of non-caseating granulomas, transmural inflammation of the gut and formation of strictures and fistulas. In rare circumstances, CD can manifest solely as a perianal disease without bowel involvement^[7]. The main differentiating features distinguishing CD from UC are the presence of granulomas, transmural disease, rectal sparing, and formation of strictures and/or fistulas. Although UC can occasionally manifest with strictures and perianal abscess or fistulas^[8], Classic UC is expressed as a contiguous inflammation almost always involving the rectum and extending proximally to the left colon or entire colon, *i.e.*, extensive colitis or pancolitis. Sometimes the disease can extend into distal 10-15 cm segment of the terminal ileum, which is termed as backwash ileitis. CD on the other hand can arise in any part of the GI tract with a segmental distribution. It usually involves the ileocecal region, sparing the rectum. Smoking is considered as a risk factor for CD, but a protective factor for UC. These features of UC and CD suggest that their etiopathogenesis may not completely overlap. In addition, there are extraintestinal manifestations such as erythema nodosum, pyoderma gangrenosum, and primary sclerosing cholangitis (PSC).

The etiology and pathogenesis of IBD remain unclear with several speculations suggesting the role of genetic factors, gut microbiome, and immune dysregulation. The interaction between these aforementioned factors gives rise to various immunogeneic types and clinical phenotypes of disease state. We term this type of traditional disease state as "idiopathic inflammatory bowel disease", "classic inflammatory bowel disease" or "primary inflammatory bowel disease" due to its unclear or idiopathic etiology and pathogenesis. However, certain identifiable factors like medications, bowel-alternating surgery, and transplantation of organs, stem cells or fecal microbiota appear to induce IBD or IBD-like changes in the GI tract. The affected individuals



essentially present with the fitting clinical symptoms and signs supported by endoscopic, histopathologic, and radiologic findings of IBD. We have termed this specific type of disease as "secondary inflammatory bowel disease (SIBD)" (Table 1).

Pathogenesis of conventional IBD

Genetic mutations or acquiring variants of certain genes have been proposed to be a pre-disposing risk factor for developing IBD. Genetic alteration in the gene coding for nucleotide-binding oligomerization domain 2 has been found to be associated with CD in about 20% of cases^[9]. This mutation is associated with reduced response to bacterial lipopolysaccharides leading to increased survivability of certain gram negative bacteria that translocate into the bowel epithelium and induce inflammation. A loss of function mutation in the alleles coding for fucosyltransferase 2 and thereby absence of secretion of this enzyme in the intestinal tract is associated with an increased risk of alterations in the microbiome^[10]. Genetic defects leading to abnormal T-cell function and macrophage activity can induce immune-mediated gut injury^[11]. These cellular alterations induce dysregulated cytokine production and release, which recruits more inflammatory cells and continues the process of immune-mediated inflammatory response. Genetic linkage analysis has shown that mutations in the IL-10/IL-10R signaling pathway has been associated with infancy or early childhood onset IBD^[12,13].

The GI tract along with the mesentery consist of a vast number of immune cells making it a highly immunogenic organ. The gut microbiome lives in harmony with the host defense system that protects the host from invasive GI pathogens. This tolerance to the gut microbiome is mediated by the homeostasis of intestinal microbiota, gut epithelial cell, stromal cells of the intestines, antigen presenting cells (dendritic cells, tissue macrophages) and inflammatory cells (neutrophils, lymphocytes)^[14]. Alteration in the gut microbiome and a dysregulated response by any of these cells can shift the delicate balance of host defense and immune tolerance leading to development of IBD^[15]. There is decreased biodiversity in the microbiome of individuals diagnosed with IBD^[16]. Although a study in monozygotic twins showed the opposite results among individuals diagnosed with CD^[17].

Dysbiosis plays an important role in the pathogenesis of classic IBD. The Phyla Firmicutes and specifically the family of Gram negative enteric organisms, i.e., Enterobacteriaceae, have been found to be abundant in the diseased state of IBD^[18,19]. Short-term antibiotic therapy has shown to improve gut inflammation, likely by affecting gut bacteria and regulating dysbiosis^[20]. Another mechanism of microbial effect on the GI tract is the ability of the gut bacteria to adhere to mucosal surface and invade the deeper submucosal layers inducing an inflammatory reaction^[21,22]. Subsequently mucosal breakdown occurs due to inflammatory cell-mediated tissue injury and the damaged mucosa further exposes the sub-epithelium to more colonies of bacteria leading to a vicious cycle of antigenic exposure and mucosal injury.

In short, the pathogenesis of IBD involves a dysregulated autoimmune response to gut dysbiosis which is precipitated by exposure to environmental factors among individuals who have a pre-existing high-risk genotype. Proinflammatory factors like tumor necrosis factor (TNF) alpha, interleukins-12/23 and cell adhesion molecules (integrins/intercellular adhesion molecules) play a key role in immune activation and recruiting immune cells^[23]. Modern biological therapy has been designed to block these mechanisms or pathways, with TNFa blockers like infliximab and adalimumab being examples. These agents have also been effective in treating other autoimmune or rheumatological conditions. Interestingly, IBD often coexist with some of the systemic autoimmune disorders, and some of them are classified as extra-intestinal manifestations of IBD. In addition, genetic linkage analyses have shown an overlap of mutations in gene loci for IBD and these other autoimmune conditions^[24]. These conditions include ankylosing spondylitis, lupus and rheumatoid arthritis. There could be a common pathogenic mechanism or a relationship at genomic level existing between IBD and other immune-mediated conditions.

DRUG-INDUCED SECONDARY IBD

Environmental factors influence the disease course in IBD patients. Two such wellstudied factors are smoking and appendectomy, which are associated with an increased risk of developing CD, and a decreased risk for UC^[25]. However, there are several other factors highlighted in this article that appear to induce IBD (Table 1). The pathogenesis of IBD is centered on dysregulated immunity, as described earlier. Hence it is not surprising to find that medications which alter the host immunity can



Table 1 Classification of secondary inflammatory bowel diseases based on etiology			
Classification			
Drug-induced secondary IBD	Immunomodulators: Azathioprine, 6-mercaptopurine, tacrolimus, mycophenolic acid, cyclosporine; Anti-TNF agents: Infliximab, adalimumab, etanercept; Anti-interleukin agents: Secukinumab, tocilizumab; Interferons: Interferon α; Immune stimulating agents: GM-CSF (sargramostim), G-CSF (filgrastim); Checkpoint inhibitors: Ipilimumab, nivolumab, pembrolizumab		
Post-surgical secondary IBD	Post-colectomy enteritis syndrome; Post-colectomy ileal pouchitis; Post-colectomy <i>de novo</i> Crohn's Disease of the Pouch; Post-bariatric surgery: Roux-en-Y gastric bypass		
Post-transplant secondary IBD	Fecal microbiota transplantation related IBD; Post- hematopoietic stem cell transplant IBD: cord colitis; Post-solid organ transplant IBD: liver, kidney		

IBD: Inflammatory bowel disease; GM-CSF: Granulocyte monocyte-colony stimulating factor; G-CSF: Granulocyte-colony stimulating factor.

sometimes lead to the development of *de novo* IBD, which we have termed as "druginduced secondary IBD"^[26]. These medications mainly include immunomodulators and biological agents. A newer class of drugs called the checkpoint inhibitors used in treating melanoma and other malignancies have also been implicated in precipitating IBD. There is a similar but weak association with the use of immune stimulating agents as well.

Immunomodulators

Immunomodulators alter the immune system mainly by inhibiting lymphocyte function. These medications consist of, but are not limited to azathioprine, 6mercaptopurine (6-MP), tacrolimus, cyclosporine A, and mycophenolate mofetil (MMF). They are commonly used to prevent graft rejection after kidney and liver transplantation. Azathioprine and 6-MP have a well-established role in achieving longterm disease remission in the management of IBD. Cyclosporine A has also been used for the treatment of refractory IBD, including during acute UC flares^[27].

Immunomodulators may induce IBD or IBD-like conditions. Studies have shown that the post-organ transplant use of immunomodulators causes a down-regulation of regulatory T-cells in the colonic mucosa^[28]. This may create a propensity to develop immune-mediated inflammation in the colon, as regulatory T-cells prevent activation of B and cytotoxic T-lymphocytes. Tacrolimus is a routinely used immunosuppressant for organ transplantation due to its predictable side-effect profile and the availability of tests for monitoring its serum level. The use of tacrolimus has been reported to induce a flare of pre-existing IBD among individuals with solid organ transplantation^[29]. Its use has also been shown to be associated with the development of de novo IBD^[29,30]. In an observational study of 53 patients without a diagnosis of IBD who underwent liver transplantation, 6 (11%) of them developed de novo IBD during a median follow up of 3.9 years^[29].

The use of MMF as an immunosuppressant is mainly seen in kidney or liver transplant recipients. One of its common side effects is diarrhea, and in about 9% of cases it causes "MMF-induced colitis"^[31,32]. This type of colitis presents with IBD-like features of endoscopic and histologic changes in the colon^[31]. The colonoscopic appearance of MMF-induced colitis is similar to that seen in classic IBD or graftversus-host disease. But it exhibits mucosal eosinophilic predominance with the lack of apoptotic microabscesses and endocrine cell aggregates in lamina propria on histology^[33]. This disease can sometimes persist even after discontinuation of MMF, taking up to 4-6 m for endoscopic resolution^[34]. There have been multiple reported cases of individuals who developed *de novo* CD after exposure to MMF, followed by improvement of disease after stopping the medication^[32,35,36]. There was a reported case of rapid resolution of MMF-induced colitis with a single dose of infliximab, suggesting the role of TNF in its pathogenesis^[37]. One can speculate that MMF-induced colitis represents a variant of IBD. Paradoxically, there is some evidence to support the use of MMF for treating active IBD^[38]. In a study of 25 patients with steroid-dependent disease unresponsive to biologics, MMF therapy achieved a clinical response in nearly 50% of cases^[39]. Two small studies have shown that MMF induced steroid-free clinical remission in about 25% of cases^[39,40]. Its therapeutic effect may help achieve long-term disease remission^[41]. Therefore it appears to be effective in treating IBD while it can induce colitis resembling IBD or lead to SIBD^[42]. This shows that interference with our complex immune system may be beneficial or detrimental for either treatment or induction of disease.

WJG | https://www.wjgnet.com

Anti-TNFa agents

Anti-TNFa inhibitors are molecules directed against the proinflammatory TNFa and they alter TNFa-mediated immune signaling in inflammatory pathways. Some examples of these agents are infliximab, adalimumab, and etanercept. They have been used for treating various immune-mediated rheumatological disorders, IBD, or IBDrelated extra-intestinal manifestations, such as ankylosing spondylitis, uveitis, erythema nodosum, and pyoderma gangrenosum^[43]. Treating one autoimmune condition can sometimes precipitate another due to alteration in the immunological homeostasis. Interestingly, individuals who are exposed to an anti-TNF agent, may develop de novo IBD or a form of drug-induced SIBD[44,45]. Along the same lines, newonset psoriasis or eczema has been reported after exposure to the anti-TNF agents^[44,46]. For example, the administration of infliximab for the treatment of IBD-related and non-IBD related inflammatory arthropathies has shown to induce autoimmune conditions like drug-induced lupus, autoimmune hepatitis and multiple sclerosis^[47-49]. In some cases a paradoxical response is encountered after its administration, where previously arthritis-free individuals with IBD develop new-onset IBD-related arthropathy^[50].

Etanercept, an anti-TNFa agent, has been extensively used to treat rheumatoid arthritis, while being ineffective in treating IBD. In fact, the use of this agent has shown to be associated with development of de novo IBD in multiple clinical observations^[45,51-53]. In a large case series of patients who received etanercept for various rheumatological disorders, 49 individuals developed *de novo* IBD^[52]. The average duration of therapy before the onset of symptoms was 3.58 mo. Its use has also been associated with precipitating pre-existing UC^[51]. In a French study, the average frequency of etanercept-related de novo IBD was approximately 0.15%. In the same study, two patients who were treated with infliximab developed IBD^[45]. In another large study of 17018 patients with auto immune disorders, several patients were on etanercept, infliximab, or adalimumab. The ones on etanercept showed a significantly increased risk of developing de novo UC or CD, yet no such effect was seen with infliximab or adalimumab^[53]. In a case report, a 56-year-old male with psoriasis who was treated with adalimumab developed de novo UC^[54]. His UC responded to the anti-IL-12/ 23 agent, ustekinumab.

The phenomenon of paradoxical response to agents like infliximab, adalimumab, or etanercept leading to the development of *de novo* IBD needs to be further explored. We hope that ongoing research may be able to identify individualized pathogenetic pathways for each patients which will aid in selecting their appropriate therapeutic agent, and pave the way to personalized medicine in IBD care. Another emerging aspect of IBD therapy is the use of biosimilars, which needs to be further studied in regard to their ability to induce de novo IBD resulting in SIBD.

Anti-interleukin agents

Anti-TNFa agents like infliximab and adalimumab have become the preferred therapies for treating IBD. Recently an anti-interleukin agent, ustekinumab (anti-IL-12/23) has been approved for the treatment of CD and UC^[55,56]. The agent has been successfully used for treating psoriasis and other rheumatological conditions for the past decade. Other anti-interleukin agents have been created for their antiinflammatory effects; and these include, but are not limited to; secukinumab, an anti-IL-17A agent used for treating psoriasis, and tocilizumab an anti-IL-6R inhibitor used for treating rheumatoid arthritis^[57,58]. In a separate study, both these agents have been found to be associated with exacerbation of pre-existing IBD^[59,60]. These agents may even induce de novo IBD in individuals at risk. Therefore we recommend that new agents, which could have the potential for therapy in immune-mediated disorders like IBD, need to be thoroughly studied for the immunogenicity and their association with the development of *de novo* IBD.

Interferon

Interferons (IFNs) are glycoprotein molecules which are secreted by host cells in response to viral infections and act as cytokines in inflammatory cascades. They have a role in inhibiting viral replication by inducing an immunological response through the activation of antigen-presenting cells, natural killer cells, neutrophils, and lymphocytes. IFNs have been investigated as potential immunological agents for treating IBD, but the studies showed no considerable clinical benefits^[61-63]. IFNa has been used for treating hepatitis B or hepatitis C infection. With the current use of highly effective direct acting antiviral agents, the use of IFNs has been largely out of practice. Nevertheless, IFNs are still used for treating multiple sclerosis, certain types



WJG https://www.wjgnet.com

of lymphomas, and leukemias.

There have been multiple reported cases of chronic hepatitis C-infected individuals who developed *de novo* UC after receiving IFNa-based therap^[64-66]. There was one reported case in which CD developed after IFN therapy for an HCV and HIV coinfected individual^[67]. All of these reported cases responded to 5-aminosalicylic acid compounds or corticosteroids. There have been several reported cases of UC flare during IFN therapy for chronic HCV infection^[68,69]. IFNa has a tendency to stimulate T helper-1 cells which play a key role in the pathogenesis of several immune-mediated disorders^[70]. This immune response in the gut may be responsible for the development of de novo IBD.

Immune stimulating agents

It has been proposed that CD in some individuals is related to a state of immune deficiency rather than over-activity. The immunodeficiency hypothesis can be used to explain the higher propensity of developing IBD among individuals with inherited immune disorders like Wiskott-Aldrich syndrome and glycogen storage disorders^[71]. Cellular cytokines like granulocyte monocyte-colony stimulating factor (GM-CSF) and granulocyte-colony stimulating factor stimulate the hematopoietic stem cells in the bone marrow to induce production and maturation of granulocytes and monocytes, which play a role in innate immunity. Animal studies have shown that mice with an increased level of anti-GM-CSF antibodies had a reduced neutrophilic phagocytic capacity. The defect in the innate mucosal defense mechanism of the gut can lead to an increased risk of developing bowel inflammation^[72]. The deficiency in GM-CSF along with nucleotide-binding oligomerization domain 2 mutations, impairs the innate immune response resulting in bacterial invasion of the lamina propria, and these bacteria in turn stimulate gut-specific T-lymphocytes^[9]. These activated T-cells produce several inflammatory markers which may lead to development of IBD among predisposed individuals.

Several lines of clinical evidence suggest an association between GM-CSF and IBD. An Australian study of patients with IBD found that the level of antibody to GM-CSF was higher in the CD cohort than that in the UC cohort^[73]. The higher levels were associated with more penetrating or stricturing disease, which in turn is associated with an increased risk of bowel surgery. Overall, the antibody level was significantly higher in the IBD group than the control group. The presence of anti-GM-CSF antibody suggests a disease state and the antibody titer could be used as a prognostic marker for disease activity, especially in case of CD^[74]. A familial association with this antibody production has been described among those who have a family history of UC or CD^[75]. The anti-GM-CSF antibody was found to be associated with an increased intestinal permeability and bacterial translocation, which was in turn correlated with a higher likelihood of developing IBD flares^[76]. The use of immune stimulating therapy has been proposed for the treatment IBD. The use of recombinant GM-CSF (sargramostim) and granulocyte-colony stimulating factor (filgrastim) for treating CD has shown some therapeutic effect^[77,78]. The studies demonstrated an improved steroidfree survival and reduction in the Crohn's disease Activity Index score^[78,79].

Among individuals with CD, low immunoglobulin levels have been associated with an increased risk for undergoing surgeries^[80]. Based on immunodeficiency hypothesis, individuals with lower immunoglobulin levels are at risk for developing severe CD. We can also speculate that such individuals could be susceptible for developing de *novo* IBD or IBD-like conditions in the presence of underlying high-risk genotype. Intravenous immunoglobulin therapy is used for various autoimmune disorders like myasthenia gravis, systemic sclerosis, and multiple sclerosis^[81-83]. Studies have also demonstrated therapeutic benefits of IVIG in treating refractory IBD^[84,85].

Checkpoint inhibitors

Checkpoint inhibitors include, but are not limited to, ipilimumab, nivolumab, and pembrolizumab. These medications are used for treating multiple malignant conditions including melanoma^[86]. Ipilimumab is a monoclonal antibody directed against the cytotoxic T-lymphocyte antigen-4 which helps in downregulating the T-cell function^[87]. Nivolumab and pembrolizumab are monoclonal antibodies which act on the programmed cell death-1 receptor. Studies have shown that approximately 30% of individuals treated with ipilimumab develop IBD-like gut inflammation^[88]. This phenomenon has been observed since its early human trials in 2009^[86,89]. Ipilimumab alters the immunological homeostasis, which could lead to the development of autoimmune conditions. It can precipitate an IBD-like disease state and also worsen pre-existing autoimmune disorders like rheumatoid arthritis, psoriasis and lupus^[90,91]. It may even permanently alter the immune system which could give rise to IBD^[92].



WJG | https://www.wjgnet.com

Ipilimumab-associated colitis shares many endoscopic and histologic features of classic IBD. The affected individuals have increased CD4 + T-lymphocytes and plasma cells in the gut mucosa, resembling that seen in IBD^[93]. There are upregulated inflammatory pathways with an increased number of inflammatory cells which in turn stimulate other surrounding inflammatory cells causing further release of cytokines and inducing mucosal injury. Increased inflammatory markers or cells in the affected colonic mucosa include granzyme B, FoxP3, and CD8 + lymphocytes^[88]. It commonly presents in the form of an inflammatory colitis, followed by an enteritis^[94]. Severe forms of the disease are susceptible to develop bowel perforation. Extreme precautions should be taken if diagnostic colonoscopy is performed in these patients with severe disease^[95,96].

The principle of treating such inflammatory conditions of the bowel is, like in case of classic IBD, to inhibit immune-mediated injury. Suppressing immunity with the use of steroids is the first-line therapy. Severe cases that do not respond to steroids, have shown to be effectively treated with infliximab, especially those with deep colonic ulcerations^[97,98]. Vedolizumab (anti-integrin molecule) has also shown to induce remission among patients with ipilimumab-induced colitis^[96,99]. With the advent of programmed cell death-1 inhibitors, the use of ipilimumab could be avoided especially due to its strongly associated adverse events and toxicities[100]. Some experts also suggest the prophylactic use of budesonide along with ipilimumab. In a published double-blinded randomized controlled trial (RCT), prophylactic use of budesonide was most beneficial if the severity of diarrhea was grade 3 or 4, and the authors recommended its use for grade 2 diarrhea or higher^[101]. Discontinuation of the medication followed by steroid therapy seems to be an effective strategy for treating ipilimumab-induced colitis. Once the symptoms resolve it is reasonable to restart therapy, since the recurrence rate of colitis is about 6% and this is independent of the duration of steroid treatment^[102].

The use of ipilimumab among patients with pre-existing IBD could induce a disease flare. As a general rule, it may be best to avoid it among patients who have an existing diagnosis of IBD^[103]. But one could make an argument for treating those patients in long-term disease remission, especially when administered in combination with corticosteroids. In this situation, we recommend to approach it on a case-by-case basis. Combination therapy of nivolumab and ipilimumab is more effective in treating melanoma than ipilimumab alone but it can increase the risk of developing colitis. In a head-to-head analysis of the two treatment options, combination therapy was associated with a higher incidence of adverse events (54%) than ipilimumab monotherapy (20%)^[104]. The most common severe adverse event was colitis, seen in 13% of overall combination treatment group compared to no cases of colitis in ipilimumab monotherapy group. Cases of nivolimumab monotherapy-induced colitis have also been reported resembling characteristics like those of UC^[105].

Currently there are no randomized clinical trials in the evaluation of therapies for checkpoint inhibitor induced colitis, although a majority of them are treated with various steroid formulations^[106]. Another approach for its treatment is the use of fecal microbiome transplantation, where stool from healthy donors is transplanted into the gut of individuals suffering from checkpoint inhibitor-induced colitis^[107]. A small case series has demonstrated the benefit in this therapy. This may also suggest the role of fecal microbiome in pathogenic mechanisms of de novo IBD^[108].

POST-SURGICAL SECONDARY IBD

Abdominal and pelvic surgeries are commonly performed in IBD patients, especially among those with CD. Studies have shown that bowel altering surgeries can have an effect on the microbiome in the GI tract, creating an environment for IBD remission or flare^[109]. For example, surgical ileocolonic resection in patients with isolated CD resulted in 10-year disease remission in 50% of cases^[110]. But some patients developed postoperative recurrence of bowel inflammation following the same surgery. A murine study showed that ileocolonic resection can alter the microbiota not just in the colon but also in the jejunum, which could precipitate IBD in the large and small bowel^[111]. Animal studies have shown that surgical changes alter the gut microbiome, which can make the commensals virulent and cause anastomotic leaks^[112,113]. These chronic mucosal lesions may represent an IBD-like phenomenon in the post-surgical bowel segments. Individuals with IBD with an abundance of bacterial species like Bacteroides vulgatus, Clostridium perfringens and Ruminococcus gnavus in the gut are at an increased risk for the development of CD of the pouch if they undergo



proctocolectomy with ileal pouch-anal anastomosis (IPAA)[114].

Post-colectomy enteritis syndrome

Patients with UC that undergo total colectomy or total proctocolectomy can develop a chronic inflammatory state of the small bowel called post-colectomy enteritis syndrome^[115,116]. It is characterized by diffuse chronic enteritis which usually develops several months after the surgery. When suspected, an upper GI endoscopy, ileoscopy *via* stoma, or enteroscopy should be performed^[117]. It appears to be distinct from CD in terms of its non-segmental involvement, superficial mucosal inflammation, absence of fistulas/strictures and the absence of granulomas. This condition can present with severe ulcerations and may even lead to a fatal outcome^[117,118]. This has been speculated to be a form of UC of the small bowel due to its lack of typical features of CD. It is usually treated with immunosuppressive therapy like corticosteroids especially during the initial presentation or a flare, followed by a long-term use of immunosuppressants like azathioprine or infliximab^[117].

Post-colectomy ileal pouchitis

Surgical resection of diseased bowel can be curative among individuals with severe forms of IBD which is unresponsive to medical treatment. Total proctocolectomy with IPAA is considered a definitive treatment for those with UC, since the entire colon with almost complete rectum is removed, leaving no available organ to manifest the disease. CD on the other hand is a segmental disease that can involve any part of the GI tract, hence resection of the diseased segments can be curative in many cases but disease recurrence is not uncommon^[119,120]. Patients with refractory UC or UC with colitis-associated neoplasia require colectomy. Restorative proctocolectomy with IPAA has become the surgical treatment of choice in those who require total colectomy. This standard bowel reconstruction surgery can be associated with pouchitis or CD of the pouch. Pouchitis, a chronic inflammation of the ileal pouch after IPAA is speculated to occur due to fecal stasis, dysbiosis, altered mucosal immunity, and surgery-associated ischemia. It is commonly treated with antibiotics but some cases do not respond to long-term antibiotic therapy and is termed as chronic antibiotic-refractory pouchitis. Therefore, pouchitis represents a disease spectrum ranging from acute antibioticresponsive phenotype to chronic antibiotic-refractory entity. The latter form of pouchitis resembles classic IBD in clinical, endoscopic, and histologic features, often requiring immunosuppressive therapy, including biologics^[121,122].

In cases that are refractory to medical therapy, fecal diversion, away from the diseased segment of the bowel is an effective modality of treatment^[123]. Experts consider chronic antibiotic-refractory pouchitis as an independent entity of IBD. Another type of pouchitis is called "diversion pouchitis". Diversion pouchitis responds to short chain fatty acids therapy^[124]. It is important to differentiate this type of inflammation from ischemic changes which are frequently seen near the stoma and is related to the surgical technique.

Fecal diversion with the construction of a stoma can be performed to treat downstream bowel or perianal diseases in IBD. In some cases, fecal diversion may induce *de novo* IBD in an uninvolved segment of the bowel^[125]; *i.e.*, postcolectomy enteritis syndrome as outlined above. This suggests that surgery is a trigger which influences the immune function in the uninvolved segment, leading to the development of inflammation.

Post-colectomy de novo CD of the pouch

Proctocolectomy with IPAA in patients with a preoperative diagnosis of UC has been shown to induce *de novo* CD among 2.7%-13% of patients who undergo the surgery for an initial diagnosis of UC or indeterminate colitis^[126]. The new disease can develop weeks to even years after the surgery. It is believed that the bowel reconstructive surgery for UC creates a CD-friendly environment. After colectomy the GI transit of consumed food is quick, altering the gut microbiome and creating a CD inducing environment. This along with areas of ischemia due to surgical alteration can give rise to *de novo* CD or induce a disease flare^[126]. The development of new disease may occur in those individuals who already have an underlying high-risk genotype for developing IBD and surgery only acts as a trigger that precipitates its phenotypic expression. The role of other environmental factors like peri-operative use of antibiotics and non-steroidal anti-inflammatory drugs, ischemia, obesity, and anxiety related to the surgical intervention may have a significant contributing effect in giving rise to this disease state.

De novo IBD after bariatric surgery

Morbid obesity is a chronic illness which can increase the risk of developing other comorbidities like diabetes, hyperlipidemia, cardiovascular diseases and several types of cancers including colon cancer. Interventions to treat obesity are mainly directed towards lifestyle changes like adopting a low-calorie diet and adequate exercise. Following these interventions can be a challenge to many and a growing number of obese individuals instead choose to undergo bariatric surgery^[127]. There are several types of bariatric procedures performed in the United States, the commonly performed ones are Roux-en-Y gastric bypass (RYGB), sleeve gastrectomy, and laparoscopic gastric band placement^[128,129].

One of the most effective surgeries for morbid obesity is RYGB and it has been shown to induce de novo CD^[130,131]. There have been several reported cases of individuals who developed new-onset IBD after undergoing a bariatric surgery^[132-135]. A recently published case series of 44 patients who developed *de novo* IBD after surgery, demonstrated that there seems to be a higher incidence of developing the disease after RYGB^[133]. The majority of cases were females and CD was the most common type of disease (n = 31), followed by UC (n = 12) and one case of indeterminate colitis. The median time to develop the disease after undergoing surgery was 7 years. Similar findings were observed in another series of 15 patients, suggesting a possible role of bariatric surgery in the development of SIBD^[136].

It is suspected that the alteration in microbiota or nutritional status could have a role in precipitating the disease in these individuals. Similarly, development of celiac disease after undergoing pancreaticodudenectomy^[137], Billroth II procedure^[138], pyloroplasty^[138] or IPAA^[139] has been reported. This could suggest an immunogenic response to the surgery leading to immune-mediated gluten sensitivity and therefore giving rise to celiac disease. Individuals who undergo bariatric surgery have shown to develop an alteration in their gut microbiota^[140,141]. This change could lead to colonic dysbiosis which plays an important role in pathogenesis of IBD^[15,16]. In addition, there is alteration in the nutritional parameters in these individuals due to artificially induced malabsorption which could play a role in the disease process, for example vitamin D deficiency and hypoglobulinemia have been linked to pathogenesis of IBD^[84,142]. Interestingly, when the surgery is performed in individuals with well controlled IBD, it has shown to be safe and effective, with an acceptable risk of postoperative complications^[143-145]. Our understanding of these mechanisms is limited^[146].

POST-TRANSPLANT SECONDARY IBD

The immune system plays a central role in the pathogenesis of IBD. Hematopoietic stem cell transplantation (HSCT) or solid organ transplantation (SOT) is performed with the use of steroids and immunomodulators. These agents suppress the immunogenic response of the recipients lymphocytes against transplanted human leukocyte antigens, to prevent immune-mediated rejection^[147]. Therefore appropriate cross-matching is carried out prior to transplantation to prevent human leukocyte antigens-mismatch^[148]. The alteration in the immune system with the use of immunosuppressant's or introduction of foreign antigens could lead to a dysregulated immune response and subsequent development of autoimmune disorders. One such autoimmune conditions is IBD that can develop after HSCT or SOT. Another type of transplantation is that of a healthy donor's fecal material, which is used to treat individuals with recurrent Clostridium difficile infection (CDI)^[149]. If the donor suffers from IBD then their gut microbiome may carry the microbiota that is proinflammatory and could lead to development of *de novo* IBD in the recipient^[150,151]. Additionally, the alteration in the gut microbiome may induce an exaggerated immune response to the newly introduced bacteria which can precipitate immune-mediated gut inflammation.

Post-hematopoietic stem cell transplant IBD

HSCT is an effective treatment in several hematopoietic disorders including leukemia. Recipients of stem cell therapy are immunosuppressed and are at risk for developing various GI infections, like infectious enterocolitis. Umbilical cord blood of newborns is a good source of harvesting stem cells which are used for HSCT^[152]. The recipients can develop a unique type of colitis, which is clinically and histologically distinct from the typical infectious colitis or colitis associated with graft-versus-host disease^[153,154]. This condition is termed as "cord colitis" and the affected individuals usually present with non-bloody diarrhea several months after the transplantation^[153]. Colonic biopsies demonstrate chronic active colitis with non-caseating granulomas and the disease can



involve the upper or lower GI tract, a pattern resembling that of CD^[154,155]. They also tend to have high loads of the bacteria Bradyrhizobium enterica in their gut. Some speculate that the alteration in gut microbiome has a role in the pathogenesis of this disease^[156]. This is also supported by the fact that antibiotic therapy is effective in the treatment. Cord colitis could be a variant of IBD which develops in the setting of altered immunity due to the use of cord blood stem cells or the use of immunomodulators.

Post-solid organ transplant IBD

Individuals who undergo SOT are immunosuppressed during the peri-transplant and post-transplantation periods. There is increased risk of developing diarrhea after transplantation, commonly due to infections or due to side effects of medications, but there are other conditions that could cause diarrhea in these patients. Two such conditions are the onset of an IBD flare in those with a pre-existing diagnosis of IBD or the development of *de novo* IBD^[157]. This phenomenon is more common after orthotopic liver transplantation than other SOT, like kidney or heart^[158]. IBD in general and UC in particular are associated with concomitant PSC, for which the mainstay of treatment is OLT. This is probably the reason for encountering higher number of OLT than SOT of other organs in the IBD population^[159,160]. The incidence risk of developing *de novo* IBD in SOT recipients versus general population is 206 and 20 respectively per 100000 person-years^[161]. In a study of post-renal transplant recipients, it was found that the incidence risk of developing IBD was twice than that for general population^[162]. In a retrospective chart review of 6800 liver and/or kidney transplant recipients that received some form of immunosuppression, it was found that 14 individuals developed de novo IBD^[161]. Post-OLT patients who develop de novo IBD had a tendency to have underlying PSC or develop PSC in the future^[160,163].

The etiopathogenesis of *de novo* IBD after SOT is likely related to the use of immunosuppressants like steroids, immunomodulators, and anti-thymocyte globulin; that alter the "immune thermostat" leading to a dysregulated immune response to gut microbiome^[157]. This in turn can lead to development of *de novo* IBD among at risk individuals. This is a paradoxical response to immunosuppression, a key principle in treating IBD with agents like azathioprine and 6-MP. Details regarding effects of various immunomodulators in precipitating IBD have been described in the previous section of this review. In a recently published retrospective study of 373 patients suffering from PSC, with or without concurrent IBD, the 10-year cumulative risk of developing a disease flare or *de novo* IBD was about 25%^[42]. These risks were higher with the use of MMF and lower with azathioprine in the post-operative period. The use of azathioprine after SOT seemed to be protective against the development of IBD flare or *de novo* IBD^[29,42].

Interestingly, there is evidence suggesting that immunosuppression with organ transplantation may be beneficial in patients with IBD. In a study of 41 IBD patients that underwent OLT the rate of clinical remission was higher than 42 IBD patients who did not undergo OLT (54% vs 33%, P = 0.03)^[164]. Transplant recipients who received MMF had better outcomes than those with other immunosuppressants. This variable response to immunotherapy by different individuals illustrates the complex nature of the immunological processes at play where alteration in immunity causes a disease flare in some while in others it could be protective against flaring. One could argue that the transplant recipients were monitored closely and were more compliant with medications, therefore had lesser incidence of disease flare.

Fecal microbiota transplantation-associated IBD

Gut microbiome consists of trillions of organisms which include bacteria, fungi and viruses. In patients with IBD there is a state of gut dysbiosis which plays a role in its pathogenesis^[150,151,165]. Individuals with recurrent CDI have a significant alteration of their colonic microbiota leading to gut dysbiosis, which can be successfully treated with fecal microbiome transplantation (FMT) from healthy donors^[149]. The fecal material is introduced via upper delivery methods (oral capsules or nasogastric/jejunal tube) or lower delivery methods (colonoscopy or enema). Among individuals with IBD who develop CDI, FMT has shown to be successful in treating 87%-91% of cases after a single transplantation^[166,167]. Interestingly, this rate of success is slightly lower when compared to individuals without IBD^[168]. Since gut dysbiosis is seen in IBD as well, investigators have utilized FMT for treating IBD flares with good response, especially in case of UC^[169,170]. There are few reported cases of CD flare that responded successfully to FMT^[171,172]. It may also have a role among patients with active disease that is not responding to biologic therapy^[172]. Fecal transplantation has also shown to modestly improve disease severity in case of chronic pouchitis^[173].



WJG | https://www.wjgnet.com

It is interesting to note that individuals treated with FMT for CDI appear to have an alteration of gut microbiome mirroring the donor's microbiome and with a higher microbial biodiversity^[174]. But in patients with IBD the microbiome after FMT resists this change and rater tends to retain the pre-transplant type of gut microbiome. This may suggest that the microbiome by itself may not be an influential pathogenic mechanism in IBD but rather a source to stimulate the immune system that drives the disease process and which in turn influences the gut microbiome. This may also explain the lack of consistent response of FMT in treating IBD. A meta-analysis of nine cohort studies, eight case studies and one RCT showed that FMT appears to be safe in IBD but its efficacy was not consistent^[169]. In a more recent meta-analysis of four RCT's comparing FMT versus placebo for the treatment of active UC, showed that clinical remission was achieved in 28% of patients in FMT group when compared to only 9% in placebo group^[170]. Rates of clinical response were 49% and 28% in the FMT and placebo groups, respectively. Well-designed RCT's are needed to further study the role of FMT in the treatment of IBD^[175].

Use of FMT is not a completely safe procedure and is associated with few adverse events, especially infections and in rare cases death^[176,177]. There have been multiple reported cases of individuals with IBD and CDI who developed an IBD flare after treatment with FMT^[176,178,179]. Studies have shown that the rate of developing a disease flare after FMT is about 15%-25%, higher among those who were transplanted via lower delivery method than peroral method^[168,180]. Based on these observations, we can speculate that FMT could also induce development of de novo IBD among individuals who carry the preferential genotype for the disease. This may be more likely if the donors themselves have active IBD. As a part of screening process for donors, several experts that FMT continue to exclude those who have IBD, even though we have not had any direct evidence that shows that such donors with IBD could cause adverse effects in the recipient^[181-183]. Perhaps the alteration of gut microbiome may be more harmful in certain individuals and further studies are needed to recognize the factors that influence the development of FMT-induced IBD flare up.

CONCLUSION

IBD have a complex pathogenesis which consists of an interaction between immune system, gut microbiome and genetic factors. These three components can be influenced by several environmental factors which could tip the balance towards an immune mediated proinflammatory state in the gut and in certain extra-intestinal tissues. In a normal state there is immune tolerance towards host microbiome. External influences can alter this balance by inducing a hyper-immune response, leading to mucosal injury of the GI tract. This type of *de novo* IBD due to specific external causes is termed as secondary inflammatory bowel disease (SIBD). These external factors have been categorized into three main groups: Drugs, surgery and organ/fecal transplantation. Drugs that can influence immunity and potentially alter it, have been implicated with the development of drug-induced SIBD; these include immunomodulators, biologics, interferons, immune stimulators, and immune checkpoint inhibitors. Bowel altering surgery can influence the microbiome and lead to malabsorption, especially in case of bowel resection and bariatric surgery. Surgeries that are used to treat one type of disease, like UC with proctocolectomy followed by ileal pouch-anal anastomosis, have shown to precipitate new forms of diseases like chronic pouchitis or CD of the pouch. This type of *de novo* chronic gut inflammation after surgery is termed as post-surgical SIBD. The role of immunosuppressants in organ transplant recipients has been key in preventing immune-mediated rejection. These immunosuppressants can paradoxically induce an autoimmune mediated gut inflammation. The donor's foreign organs or stem cells could also induce an immune reaction causing immune mediated tissue injury. Both these factors can lead to post-solid organ/stem cell transplant related. The interaction between microbiota and host immune system is complex process, and factors altering bowel anatomy and gut homeostasis may reset host's immune thermostat, triggering the development of IBD or SIBD.

ACKNOWLEDGEMENTS

Dr. Bo Shen is supported by the Edelman-Jarislowsky endowed professorship in surgical sciences.



WJG https://www.wjgnet.com

REFERENCES

- Tremaine WJ. Diagnosis and treatment of indeterminate colitis. Gastroenterol Hepatol (NY) 2011; 7: 826-828 [PMID: 22347823]
- 2 Burakoff R. Indeterminate colitis: clinical spectrum of disease. J Clin Gastroenterol 2004; 38: S41-S43 [PMID: 15115931 DOI: 10.1097/01.mcg.0000123991.13937.7e]
- Loftus EV Jr. Clinical epidemiology of inflammatory bowel disease: Incidence, prevalence, and 3 environmental influences. Gastroenterology 2004; 126: 1504-1517 [PMID: 15168363 DOI: 10.1053/j.gastro.2004.01.063]
- Ng SC, Tang W, Ching JY, Wong M, Chow CM, Hui AJ, Wong TC, Leung VK, Tsang SW, Yu HH, Li MF, Ng KK, Kamm MA, Studd C, Bell S, Leong R, de Silva HJ, Kasturiratne A, Mufeena MNF, Ling KL, Ooi CJ, Tan PS, Ong D, Goh KL, Hilmi I, Pisespongsa P, Manatsathit S, Rerknimitr R, Aniwan S, Wang YF, Ouyang Q, Zeng Z, Zhu Z, Chen MH, Hu PJ, Wu K, Wang X, Simadibrata M, Abdullah M, Wu JC, Sung JJY, Chan FKL; Asia-Pacific Crohn's and Colitis Epidemiologic Study (ACCESS) Study Group. Incidence and phenotype of inflammatory bowel disease based on results from the Asia-pacific Crohn's and colitis epidemiology study. Gastroenterology 2013; 145: 158-165.e2 [PMID: 23583432 DOI: 10.1053/j.gastro.2013.04.007]
- Ray G. Inflammatory bowel disease in India Past, present and future. World J Gastroenterol 2016; 22: 8123-8136 [PMID: 27688654 DOI: 10.3748/wjg.v22.i36.8123]
- 6 Smids C, Horjus Talabur Horje CS, Groenen MJM, van Koolwijk EHM, Wahab PJ, van Lochem EG. The value of serum antibodies in differentiating inflammatory bowel disease, predicting disease activity and disease course in the newly diagnosed patient. Scand J Gastroenterol 2017; 52: 1104-1112 [PMID: 28661185 DOI: 10.1080/00365521.2017.1344875]
- Schwartz DA, Ghazi LJ, Regueiro M, Fichera A, Zoccali M, Ong EM, Mortelé KJ; Crohn's & Colitis 7 Foundation of America, Inc. Guidelines for the multidisciplinary management of Crohn's perianal fistulas: summary statement. Inflamm Bowel Dis 2015; 21: 723-730 [PMID: 25751066 DOI: 10.1097/MIB.000000000000315
- 8 de Dombal FT, Watts JM, Watkinson G, Goligher JC. Incidence and management of anorectal abscess, fistula and fissure, in patients with ulcerative colitis. Dis Colon Rectum 1966; 9: 201-206 [PMID: 5949200 DOI: 10.1007/BF02616981]
- Ogura Y, Bonen DK, Inohara N, Nicolae DL, Chen FF, Ramos R, Britton H, Moran T, Karaliuskas R, Duerr RH, Achkar JP, Brant SR, Bayless TM, Kirschner BS, Hanauer SB, Nuñez G, Cho JH. A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. Nature 2001; 411: 603-606 [PMID: 11385577 DOI: 10.1038/35079114]
- Rausch P, Rehman A, Künzel S, Häsler R, Ott SJ, Schreiber S, Rosenstiel P, Franke A, Baines JF. Colonic 10 mucosa-associated microbiota is influenced by an interaction of Crohn disease and FUT2 (Secretor) genotype. Proc Natl Acad Sci USA 2011; 108: 19030-19035 [PMID: 22068912 DOI: 10.1073/pnas.1106408108
- 11 Geremia A, Biancheri P, Allan P, Corazza GR, Di Sabatino A. Innate and adaptive immunity in inflammatory bowel disease. Autoimmun Rev 2014; 13: 3-10 [PMID: 23774107 DOI: 10.1016/j.autrev.2013.06.004]
- 12 Lee CH, Hsu P, Nanan B, Nanan R, Wong M, Gaskin KJ, Leong RW, Murchie R, Muise AM, Stormon MO. Novel de novo mutations of the interleukin-10 receptor gene lead to infantile onset inflammatory bowel disease. J Crohns Colitis 2014; 8: 1551-1556 [PMID: 24813381 DOI: 10.1016/j.crohns.2014.04.004]
- Glocker EO, Kotlarz D, Boztug K, Gertz EM, Schäffer AA, Noyan F, Perro M, Diestelhorst J, Allroth A, Murugan D, Hätscher N, Pfeifer D, Sykora KW, Sauer M, Kreipe H, Lacher M, Nustede R, Woellner C, Baumann U, Salzer U, Koletzko S, Shah N, Segal AW, Sauerbrey A, Buderus S, Snapper SB, Grimbacher B, Klein C. Inflammatory bowel disease and mutations affecting the interleukin-10 receptor. N Engl J Med 2009; 361: 2033-2045 [PMID: 19890111 DOI: 10.1056/NEJMoa0907206]
- Abraham C, Medzhitov R. Interactions between the host innate immune system and microbes in 14 inflammatory bowel disease. Gastroenterology 2011; 140: 1729-1737 [PMID: 21530739 DOI: 10.1053/j.gastro.2011.02.012
- 15 Mizoguchi E, Low D, Ezaki Y, Okada T. Recent updates on the basic mechanisms and pathogenesis of inflammatory bowel diseases in experimental animal models. Intest Res 2020; 18: 151-167 [PMID: 32326669 DOI: 10.5217/ir.2019.09154]
- Kostic AD, Xavier RJ, Gevers D. The microbiome in inflammatory bowel disease: current status and the 16 future ahead. Gastroenterology 2014; 146: 1489-1499 [PMID: 24560869 DOI: 10.1053/j.gastro.2014.02.009
- 17 Dicksved J, Halfvarson J, Rosenquist M, Järnerot G, Tysk C, Apajalahti J, Engstrand L, Jansson JK. Molecular analysis of the gut microbiota of identical twins with Crohn's disease. ISME J 2008; 2: 716-727 [PMID: 18401439 DOI: 10.1038/ismej.2008.37]
- 18 Kang S, Denman SE, Morrison M, Yu Z, Dore J, Leclerc M, McSweeney CS. Dysbiosis of fecal microbiota in Crohn's disease patients as revealed by a custom phylogenetic microarray. Inflamm Bowel Dis 2010; 16: 2034-2042 [PMID: 20848492 DOI: 10.1002/ibd.21319]
- Lupp C, Robertson ML, Wickham ME, Sekirov I, Champion OL, Gaynor EC, Finlay BB. Host-mediated 19 inflammation disrupts the intestinal microbiota and promotes the overgrowth of Enterobacteriaceae. Cell Host Microbe 2007; 2: 119-129 [PMID: 18005726 DOI: 10.1016/j.chom.2007.06.010]
- Casellas F, Borruel N, Papo M, Guarner F, Antolín M, Videla S, Malagelada JR. Antiinflammatory effects of enterically coated amoxicillin-clavulanic acid in active ulcerative colitis. Inflamm Bowel Dis 1998; 4: 1-5 [PMID: 9552221 DOI: 10.1097/00054725-199802000-00001]
- 21 Swidsinski A, Weber J, Loening-Baucke V, Hale LP, Lochs H. Spatial organization and composition of the mucosal flora in patients with inflammatory bowel disease. J Clin Microbiol 2005; 43: 3380-3389 [PMID: 16000463 DOI: 10.1128/JCM.43.7.3380-3389.2005]
- 22 Meconi S, Vercellone A, Levillain F, Payré B, Al Saati T, Capilla F, Desreumaux P, Darfeuille-Michaud A,



Altare F. Adherent-invasive Escherichia coli isolated from Crohn's disease patients induce granulomas in vitro. Cell Microbiol 2007; 9: 1252-1261 [PMID: 17223928 DOI: 10.1111/j.1462-5822.2006.00868.x]

- Danese S, Fiocchi C. Ulcerative colitis. N Engl J Med 2011; 365: 1713-1725 [PMID: 22047562 DOI: 23 10.1056/NEJMra1102942]
- Lees CW, Barrett JC, Parkes M, Satsangi J. New IBD genetics: common pathways with other diseases. Gut 24 2011; 60: 1739-1753 [PMID: 21300624 DOI: 10.1136/gut.2009.199679]
- Burke KE, Boumitri C, Ananthakrishnan AN, Modifiable Environmental Factors in Inflammatory Bowel 25 Disease. Curr Gastroenterol Rep 2017; 19: 21 [PMID: 28397132 DOI: 10.1007/s11894-017-0562-0]
- 26 Dubeau MF, Iacucci M, Beck PL, Moran GW, Kaplan GG, Ghosh S, Panaccione R. Drug-induced inflammatory bowel disease and IBD-like conditions. Inflamm Bowel Dis 2013; 19: 445-456 [PMID: 22573536 DOI: 10.1002/ibd.22990]
- Williams JG, Alam MF, Alrubaiy L, Arnott I, Clement C, Cohen D, Gordon JN, Hawthorne AB, Hilton M, 27 Hutchings HA, Jawhari AU, Longo M, Mansfield J, Morgan JM, Rapport F, Seagrove AC, Sebastian S, Shaw I. Travis SP. Watkins A. Infliximab versus ciclosporin for steroid-resistant acute severe ulcerative colitis (CONSTRUCT): a mixed methods, open-label, pragmatic randomised trial. Lancet Gastroenterol Hepatol 2016; 1: 15-24 [PMID: 27595142 DOI: 10.1016/S2468-1253(16)30003-6]
- Verdonk RC, Haagsma EB, Jonker MR, Bok LI, Zandvoort JH, Kleibeuker JH, Faber KN, Dijkstra G. Effects of different immunosuppressive regimens on regulatory T-cells in noninflamed colon of liver transplant recipients. Inflamm Bowel Dis 2007; 13: 703-709 [PMID: 17230494 DOI: 10.1002/ibd.20087]
- 29 Haagsma EB, Van Den Berg AP, Kleibeuker JH, Slooff MJ, Dijkstra G. Inflammatory bowel disease after liver transplantation: the effect of different immunosuppressive regimens. Aliment Pharmacol Ther 2003; 18: 33-44 [PMID: 12848624 DOI: 10.1046/j.1365-2036.2003.01613.x]
- Kurnatowska I, Banasiak M, Daniel P, Wagrowska-Danilewicz M, Nowicki M. Two cases of severe de 30 novo colitis in kidney transplant recipients after conversion to prolonged-release tacrolimus. Transpl Int 2010; 23: 553-558 [PMID: 19951264 DOI: 10.1111/j.1432-2277.2009.01009.x]
- 31 Calmet FH, Yarur AJ, Pukazhendhi G, Ahmad J, Bhamidimarri KR. Endoscopic and histological features of mycophenolate mofetil colitis in patients after solid organ transplantation. Ann Gastroenterol 2015; 28: 366-373 [PMID: 26126799]
- 32 Farooqi R, Kamal A, Burke C. Mycophenolate-induced Colitis: A Case Report with Focused Review of Literature. Cureus 2020; 12: e6774 [PMID: 32117661 DOI: 10.7759/cureus.6774]
- 33 Star KV, Ho VT, Wang HH, Odze RD. Histologic features in colon biopsies can discriminate mycophenolate from GVHD-induced colitis. Am J Surg Pathol 2013; 37: 1319-1328 [PMID: 24076772 DOI: 10.1097/PAS.0b013e31829ab1ef]
- Al-Absi AI, Cooke CR, Wall BM, Sylvestre P, Ismail MK, Mya M. Patterns of injury in mycophenolate 34 mofetil-related colitis. Transplant Proc 2010; 42: 3591-3593 [PMID: 21094821 DOI: 10.1016/j.transproceed.2010.08.066]
- 35 Jakes AD, Roy A, Veerasamy M, Bhandari S. Case report: Crohn's-like mycophenolate-induced colitis, a fallout in steroid-free regimens. Transplant Proc 2013; 45: 842-844 [PMID: 23498833 DOI: 10.1016/i transproceed 2012.11.003
- Moroncini G, Benfaremo D, Mandolesi A, Gabrielli A. Mycophenolate mofetil-induced colitis in a patient with systemic sclerosis. BMJ Case Rep 2018; 2018 [PMID: 29776943 DOI: 10.1136/bcr-2018-224829]
- Bouhbouh S, Rookmaaker MB. Rapid resolution of persistent mycophenolate mofetil-induced diarrhoea 37 with a single dose of infliximab. Nephrol Dial Transplant 2010; 25: 3437-3438 [PMID: 20615909 DOI: 10.1093/ndt/gfq379]
- Smith MR, Cooper SC. Mycophenolate mofetil therapy in the management of inflammatory bowel disease-38 -a retrospective case series and review. J Crohns Colitis 2014; 8: 890-897 [PMID: 24507162 DOI: 10.1016/j.crohns.2014.01.014
- Macaluso FS, Maida M, Renna S, Orlando E, Affronti M, Sapienza C, Dimarco M, Orlando R, Rizzuto G, 39 Cottone M, Orlando A. Mycophenolate mofetil is a valid option in patients with inflammatory bowel disease resistant to TNF-a inhibitors and conventional immunosuppressants. Dig Liver Dis 2017; 49: 157-162 [PMID: 27876682 DOI: 10.1016/j.dld.2016.10.001]
- Palaniappan S, Ford AC, Greer D, Everett SM, Chalmers DM, Axon AT, Hamlin PJ. Mycophenolate 40 mofetil therapy for refractory inflammatory bowel disease. Inflamm Bowel Dis 2007; 13: 1488-1492 [PMID: 17924566 DOI: 10.1002/ibd.20258]
- 41 Tan T, Lawrance IC. Use of mycophenolate mofetil in inflammatory bowel disease. World J Gastroenterol 2009; 15: 1594-1599 [PMID: 19340901 DOI: 10.3748/wjg.15.1594]
- 42 Mouchli MA, Singh S, Boardman L, Bruining DH, Lightner AL, Rosen CB, Heimbach JK, Hasan B, Poterucha JJ, Watt KD, Kane SV, Raffals LE, Loftus EV Jr. Natural History of Established and De Novo Inflammatory Bowel Disease After Liver Transplantation for Primary Sclerosing Cholangitis. Inflamm Bowel Dis 2018; 24: 1074-1081 [PMID: 29522202 DOI: 10.1093/ibd/izx096]
- 43 Peyrin-Biroulet L, Van Assche G, Gómez-Ulloa D, García-Álvarez L, Lara N, Black CM, Kachroo S. Systematic Review of Tumor Necrosis Factor Antagonists in Extraintestinal Manifestations in Inflammatory Bowel Disease. Clin Gastroenterol Hepatol 2017; 15: 25-36.e27 [PMID: 27392760 DOI: 10.1016/j.cgh.2016.06.025]
- 44 Fouache D, Goëb V, Massy-Guillemant N, Avenel G, Bacquet-Deschryver H, Kozyreff-Meurice M, Ménard JF, Muraine M, Savoye G, Le Loët X, Tharasse C, Vittecoq O. Paradoxical adverse events of antitumour necrosis factor therapy for spondyloarthropathies: a retrospective study. Rheumatology (Oxford) 2009; 48: 761-764 [PMID: 19395543 DOI: 10.1093/rheumatology/kep083]
- Toussirot É, Houvenagel É, Goëb V, Fouache D, Martin A, Le Dantec P, Dernis E, Wendling D, 45 Ansemant T, Berthelot JM, Bader-Meunier B, Kantelip B, Le CRI. Development of inflammatory bowel disease during anti-TNF- α therapy for inflammatory rheumatic disease: a nationwide series. Joint Bone Spine 2012; 79: 457-463 [PMID: 22088934 DOI: 10.1016/j.jbspin.2011.10.001]
- Rahier JF, Buche S, Peyrin-Biroulet L, Bouhnik Y, Duclos B, Louis E, Papay P, Allez M, Cosnes J, Cortot A, Laharie D, Reimund JM, Lémann M, Delaporte E, Colombel JF; Groupe d'Etude Thérapeutique des Affections Inflammatoires du Tube Digestif (GETAID). Severe skin lesions cause patients with



inflammatory bowel disease to discontinue anti-tumor necrosis factor therapy. Clin Gastroenterol Hepatol 2010; 8: 1048-1055 [PMID: 20728573 DOI: 10.1016/j.cgh.2010.07.022]

- 47 Yilmaz B, Roach EC, Koklu S. Infliximab leading to autoimmune hepatitis: an increasingly recognized side effect. Dig Dis Sci 2014; 59: 2602-2603 [PMID: 25146841 DOI: 10.1007/s10620-014-3323-z]
- 48 Costa MF, Said NR, Zimmermann B. Drug-induced lupus due to anti-tumor necrosis factor alpha agents. Semin Arthritis Rheum 2008; 37: 381-387 [PMID: 17977585 DOI: 10.1016/j.semarthrit.2007.08.003]
- Robinson WH, Genovese MC, Moreland LW. Demyelinating and neurologic events reported in association 49 with tumor necrosis factor alpha antagonism: by what mechanisms could tumor necrosis factor alpha antagonists improve rheumatoid arthritis but exacerbate multiple sclerosis? Arthritis Rheum 2001; 44: 1977-1983 [PMID: 11592357 DOI: 10.1002/1529-0131(200109)44:9<1977::AID-ART345>3.0.CO;2-6]
- Thiebault H, Boyard-Lasselin P, Guignant C, Guillaume N, Wacrenier A, Sabbagh C, Rebibo L, Brazier F, Meynier J, Nguyen-Khac E, Dupas JL, Goëb V, Fumery M. Paradoxical articular manifestations in patients with inflammatory bowel diseases treated with infliximab. Eur J Gastroenterol Hepatol 2016; 28: 876-881 [PMID: 27101404 DOI: 10.1097/MEG.000000000000643]
- Prescott K, Costner M, Cohen S, Kazi S. Tumor necrosis factor-alpha inhibitor associated ulcerative colitis. 51 Am J Med Sci 2007; 333: 137-139 [PMID: 17496730 DOI: 10.1097/MAJ.0b013e3180312362]
- 52 O'Toole A. Lucci M. Korzenik J. Inflammatory Bowel Disease Provoked by Etanercept: Report of 443 Possible Cases Combined from an IBD Referral Center and the FDA. Dig Dis Sci 2016; 61: 1772-1774 [PMID: 26728477 DOI: 10.1007/s10620-015-4007-z]
- 53 Korzenik J, Larsen MD, Nielsen J, Kjeldsen J, Nørgård BM. Increased risk of developing Crohn's disease or ulcerative colitis in 17 018 patients while under treatment with anti-TNF α agents, particularly etanercept, for autoimmune diseases other than inflammatory bowel disease. Aliment Pharmacol Ther 2019; 50: 289-294 [PMID: 31267570 DOI: 10.1111/apt.15370]
- 54 Kolios AGA, Biedermann L, Weber A, Navarini AA, Meier J, Cozzio A, French LE. Paradoxical ulcerative colitis during adalimumab treatment of psoriasis resolved by switch to ustekinumab. Br J Dermatol 2018; 178: 551-555 [PMID: 28477389 DOI: 10.1111/bjd.15631]
- Kawalec P, Holko P, Moćko P, Pilc A. Comparative effectiveness of abatacept, apremilast, secukinumab and ustekinumab treatment of psoriatic arthritis: a systematic review and network meta-analysis, Rheumatol Int 2018; 38: 189-201 [PMID: 29285605 DOI: 10.1007/s00296-017-3919-7]
- 56 Wils P, Bouhnik Y, Michetti P, Flourie B, Brixi H, Bourrier A, Allez M, Duclos B, Serrero M, Buisson A, Amiot A, Fumery M, Roblin X, Peyrin-Biroulet L, Filippi J, Bouguen G, Abitbol V, Coffin B, Simon M, Laharie D, Pariente B; Groupe d'Etude Thérapeutique des Affections Inflammatoires du Tube Digestif (GETAID). Long-term efficacy and safety of ustekinumab in 122 refractory Crohn's disease patients: a multicentre experience. Aliment Pharmacol Ther 2018; 47: 588-595 [PMID: 29315694 DOI: 10.1111/apt.14487]
- 57 Puig L. Paradoxical Reactions: Anti-Tumor Necrosis Factor Alpha Agents, Ustekinumab, Secukinumab, Ixekizumab, and Others. Curr Probl Dermatol 2018; 53: 49-63 [PMID: 29131037 DOI: 10.1159/000479475]
- Dige A, Støy S, Rasmussen TK, Kelsen J, Hvas CL, Sandahl TD, Dahlerup JF, Deleuran B, Agnholt J. 58 Increased levels of circulating Th17 cells in quiescent versus active Crohn's disease. J Crohns Colitis 2013; 7: 248-255 [PMID: 22784949 DOI: 10.1016/j.crohns.2012.06.015]
- Atreva R, Billmeier U, Rath T, Mudter J, Vieth M, Neumann H, Neurath MF. First case report of 59 exacerbated ulcerative colitis after anti-interleukin-6R salvage therapy. World J Gastroenterol 2015; 21: 12963-12969 [PMID: 26668517 DOI: 10.3748/wjg.v21.i45.12963]
- Wang J, Bhatia A, Krugliak Cleveland N, Gupta N, Dalal S, Rubin DT, Sakuraba A. Rapid Onset of Inflammatory Bowel Disease after Receiving Secukinumab Infusion. ACG Case Rep J 2018; 5: e56 [PMID: 30105273 DOI: 10.14309/crj.2018.56]
- 61 Tilg H, Vogelsang H, Ludwiczek O, Lochs H, Kaser A, Colombel JF, Ulmer H, Rutgeerts P, Krüger S, Cortot A, D'Haens G, Harrer M, Gasche C, Wrba F, Kuhn I, Reinisch W. A randomised placebo controlled trial of pegylated interferon alpha in active ulcerative colitis. Gut 2003; 52: 1728-1733 [PMID: 14633951 DOI: 10.1136/gut.52.12.1728]
- 62 Wang Y, MacDonald JK, Benchimol EI, Griffiths AM, Steinhart AH, Panaccione R, Seow CH. Type I interferons for induction of remission in ulcerative colitis. Cochrane Database Syst Rev 2015: CD006790 [PMID: 26368001 DOI: 10.1002/14651858.CD006790.pub3]
- 63 Musch E, Andus T, Kruis W, Raedler A, Spehlmann M, Schreiber S, Krakamp B, Malek M, Malchow H, Zavada F, Engelberg Feurle G. Interferon-beta-1a for the treatment of steroid-refractory ulcerative colitis: a randomized, double-blind, placebo-controlled trial. Clin Gastroenterol Hepatol 2005; 3: 581-586 [PMID: 15952100 DOI: 10.1016/s1542-3565(05)00208-9]
- Villa F, Rumi MG, Signorelli C, Saibeni S, Del Ninno E, Ferrero Bogetto S, de Franchis R, Vecchi M. Onset of inflammatory bowel diseases during combined alpha-interferon and ribayirin therapy for chronic hepatitis C: report of two cases. Eur J Gastroenterol Hepatol 2005; 17: 1243-1245 [PMID: 16215439 DOI: 10.1097/00042737-200511000-00015
- Mavrogiannis C, Papanikolaou IS, Elefsiniotis IS, Psilopoulos DI, Karameris A, Karvountzis G. Ulcerative 65 colitis associated with interferon treatment for chronic hepatitis C. J Hepatol 2001; 34: 964-965 [PMID: 11451187 DOI: 10.1016/s0168-8278(01)00022-8]
- Sprenger R, Sagmeister M, Offner F. Acute ulcerative colitis during successful interferon/ribavirin 66 treatment for chronic hepatitis. Gut 2005; 54: 438-9; author reply 439 [PMID: 15710996 DOI: 10.1136/gut.2004.049940]
- 67 Bongiovanni M, Ranieri R, Ferrero S, Casanova F, Monforte Ad. Crohn's disease onset in an HIV/hepatitis C virus co-infected woman taking pegylated interferon alpha-2b plus ribavirin. AIDS 2006; 20: 1989-1990 [PMID: 16988527 DOI: 10.1097/01.aids.0000247127.19882.f6]
- 68 Watanabe T, Inoue M, Harada K, Homma N, Uchida M, Ogata N, Funada R, Hasegawa K, Soga K, Shibasaki K. A case of exacerbation of ulcerative colitis induced by combination therapy with PEGinterferon alpha-2b and ribavirin. Gut 2006; 55: 1682-1683 [PMID: 17047132 DOI: 10.1136/gut.2006.105197]



- 69 Mitoro A, Yoshikawa M, Yamamoto K, Mimura M, Yoshikawa Y, Shiroi A, Mochi T, Sakamoto T, Yamao J, Kikuchi E. Exacerbation of ulcerative colitis during alpha-interferon therapy for chronic hepatitis C. Intern Med 1993; 32: 327-331 [PMID: 8102914 DOI: 10.2169/internalmedicine.32.327]
- Tilg H. New insights into the mechanisms of interferon alfa: an immunoregulatory and anti-inflammatory cytokine. Gastroenterology 1997; 112: 1017-1021 [PMID: 9041265 DOI: 10.1053/gast.1997.v112.pm9041265]
- Doe WF. Immunodeficiency and the gastrointestinal tract. Clin Gastroenterol 1983; 12: 839-853 [PMID: 71 6604600
- 72 Han X, Uchida K, Jurickova I, Koch D, Willson T, Samson C, Bonkowski E, Trauernicht A, Kim MO, Tomer G, Dubinsky M, Plevy S, Kugathsan S, Trapnell BC, Denson LA. Granulocyte-macrophage colonystimulating factor autoantibodies in murine ileitis and progressive ileal Crohn's disease. Gastroenterology 2009; 136: 1261-1271, e1-e3 [PMID: 19230854 DOI: 10.1053/j.gastro.2008.12.046]
- 73 Gathungu G, Kim MO, Ferguson JP, Sharma Y, Zhang W, Ng SM, Bonkowski E, Ning K, Simms LA, Croft AR, Stempak JM, Walker N, Huang N, Xiao Y, Silverberg MS, Trapnell B, Cho JH, Radford-Smith GL, Denson LA. Granulocyte-macrophage colony-stimulating factor autoantibodies: a marker of aggressive Crohn's disease. Inflamm Bowel Dis 2013; 19: 1671-1680 [PMID: 23749272 DOI: 10.1097/MIB.0b013e318281f506]
- 74 Bonneau J, Dumestre-Perard C, Rinaudo-Gaujous M, Genin C, Sparrow M, Roblin X, Paul S. Systematic review: new serological markers (anti-glycan, anti-GP2, anti-GM-CSF Ab) in the prediction of IBD patient outcomes. Autoimmun Rev 2015; 14: 231-245 [PMID: 25462578 DOI: 10.1016/j.autrev.2014.11.004]
- Wright SS, Trauernicht A, Bonkowski E, McCall CA, Maier EA, Bezold R, Lake K, Chalk C, Trapnell BC, 75 Kim MO, Kugathasan S, Denson LA. Familial Association of Granulocyte-Macrophage Colony-Stimulating Factor Autoantibodies in Inflammatory Bowel Disease. J Pediatr Gastroenterol Nutr 2018; 66: 767-772 [PMID: 29216019 DOI: 10.1097/MPG.00000000001851]
- Däbritz J, Bonkowski E, Chalk C, Trapnell BC, Langhorst J, Denson LA, Foell D. Granulocyte 76 macrophage colony-stimulating factor auto-antibodies and disease relapse in inflammatory bowel disease. Am J Gastroenterol 2013; 108: 1901-1910 [PMID: 24145675 DOI: 10.1038/ajg.2013.360]
- Korzenik JR, Dieckgraefe BK, Valentine JF, Hausman DF, Gilbert MJ: Sargramostim in Crohn's Disease 77 Study Group. Sargramostim for active Crohn's disease. N Engl J Med 2005; 352: 2193-2201 [PMID: 15917384 DOI: 10.1056/NEJMoa0411091
- Barahona-Garrido J, Yamamoto-Furusho JK. New treatment options in the management of IBD focus 78 on colony stimulating factors. Biologics 2008; 2: 501-504 [PMID: 19707380 DOI: 10.2147/btt.s3543]
- Takazoe M, Matsui T, Motoya S, Matsumoto T, Hibi T, Watanabe M. Sargramostim in patients with Crohn's disease: results of a phase 1-2 study. J Gastroenterol 2009; 44: 535-543 [PMID: 19352588 DOI: 10.1007/s00535-009-0029-7
- Horton N, Wu X, Philpott J, Garber A, Achkar JP, Brzezinski A, Lashner BA, Shen B. Impact of Low 80 Immunoglobulin G Levels on Disease Outcomes in Patients with Inflammatory Bowel Diseases. Dig Dis Sci 2016; 61: 3270-3277 [PMID: 27619393 DOI: 10.1007/s10620-016-4294-z]
- Barnett C, Wilson G, Barth D, Katzberg HD, Bril V. Changes in quality of life scores with intravenous immunoglobulin or plasmapheresis in patients with myasthenia gravis. J Neurol Neurosurg Psychiatry 2013; 84: 94-97 [PMID: 23154126 DOI: 10.1136/jnnp-2011-301449]
- 82 Poelman CL, Hummers LK, Wigley FM, Anderson C, Boin F, Shah AA. Intravenous immunoglobulin may be an effective therapy for refractory, active diffuse cutaneous systemic sclerosis. J Rheumatol 2015; 42: 236-242 [PMID: 25433527 DOI: 10.3899/jrheum.140833]
- Pöhlau D, Przuntek H, Sailer M, Bethke F, Koehler J, König N, Heesen C, Späth P, Andresen I. 83 Intravenous immunoglobulin in primary and secondary chronic progressive multiple sclerosis: a randomized placebo controlled multicentre study. Mult Scler 2007; 13: 1107-1117 [PMID: 17623736 DOI: 10.1177/1352458507078400
- Horton N, Kochhar G, Patel K, Lopez R, Shen B. Efficacy and Factors Associated with Treatment 84 Response of Intravenous Immunoglobulin in Inpatients with Refractory Inflammatory Bowel Diseases. Inflamm Bowel Dis 2017; 23: 1080-1087 [PMID: 28452863 DOI: 10.1097/MIB.000000000001116]
- Merkley SA, Beaulieu DB, Horst S, Duley C, Annis K, Nohl A, Schwartz DA. Use of Intravenous 85 Immunoglobulin for Patients with Inflammatory Bowel Disease with Contraindications or Who Are Unresponsive to Conventional Treatments. Inflamm Bowel Dis 2015; 21: 1854-1859 [PMID: 25993689 DOI: 10.1097/MIB.0000000000004561
- Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, Akerley W, van den Eertwegh AJ, Lutzky J, Lorigan P, Vaubel JM, Linette GP, Hogg D, Ottensmeier CH, Lebbé C, Peschel C, Quirt I, Clark JI, Wolchok JD, Weber JS, Tian J, Yellin MJ, Nichol GM, Hoos A, Urba WJ. Improved survival with ipilimumab in patients with metastatic melanoma. N Engl J Med 2010; 363: 711-723 [PMID: 20525992 DOI: 10.1056/NEJMoa1003466]
- 87 Friedman CF, Wolchok JD. Checkpoint inhibition and melanoma: Considerations in treating the older adult. J Geriatr Oncol 2017; 8: 237-241 [PMID: 28506536 DOI: 10.1016/j.jgo.2017.04.003]
- Arriola E, Wheater M, Lopez MA, Thomas G, Ottensmeier C. Evaluation of immune infiltration in the 88 colonic mucosa of patients with ipilimumab-related colitis. Oncoimmunology 2016; 5: e1209615 [PMID: 27757302 DOI: 10.1080/2162402X.2016.1209615]
- 89 Downey SG, Klapper JA, Smith FO, Yang JC, Sherry RM, Royal RE, Kammula US, Hughes MS, Allen TE, Levy CL, Yellin M, Nichol G, White DE, Steinberg SM, Rosenberg SA. Prognostic factors related to clinical response in patients with metastatic melanoma treated by CTL-associated antigen-4 blockade. Clin Cancer Res 2007; 13: 6681-6688 [PMID: 17982122 DOI: 10.1158/1078-0432.CCR-07-0187]
- 90 Johnson DB, Sullivan RJ, Ott PA, Carlino MS, Khushalani NI, Ye F, Guminski A, Puzanov I, Lawrence DP, Buchbinder EI, Mudigonda T, Spencer K, Bender C, Lee J, Kaufman HL, Menzies AM, Hassel JC, Mehnert JM, Sosman JA, Long GV, Clark JI. Ipilimumab Therapy in Patients With Advanced Melanoma and Preexisting Autoimmune Disorders. JAMA Oncol 2016; 2: 234-240 [PMID: 26633184 DOI: 10.1001/jamaoncol.2015.4368
- Tison A, Quéré G, Misery L, Funck-Brentano E, Danlos FX, Routier E, Robert C, Loriot Y, Lambotte O,



Bonniaud B, Scalbert C, Maanaoui S, Lesimple T, Martinez S, Marcq M, Chouaid C, Dubos C, Brunet-Possenti F, Stavris C, Chiche L, Beneton N, Mansard S, Guisier F, Doubre H, Skowron F, Aubin F, Zehou O, Roge C, Lambert M, Pham-Ledard A, Beylot-Barry M, Veillon R, Kramkimel N, Giacchero D, De Quatrebarbes J, Michel C, Auliac JB, Gonzales G, Decroisette C, Le Garff G, Carpiuc I, Vallerand H, Nowak E, Cornec D, Kostine M; Groupe de Cancérologie Cutanée, Groupe Français de Pneumo-Cancérologie, and Club Rhumatismes et Inflammations, Safety and Efficacy of Immune Checkpoint Inhibitors in Patients With Cancer and Preexisting Autoimmune Disease: A Nationwide, Multicenter Cohort Study. Arthritis Rheumatol 2019; 71: 2100-2111 [PMID: 31379105 DOI: 10.1002/art.41068]

- Akel R, Anouti B, Tfayli A. Late-Onset Inflammatory Bowel Disease-Like Syndrome after Ipilimumab 92 Therapy: A Case Report. Case Rep Oncol 2017; 10: 456-461 [PMID: 28626406 DOI: 10.1159/000475709]
- Bamias G, Delladetsima I, Perdiki M, Siakavellas SI, Goukos D, Papatheodoridis GV, Daikos GL, Gogas H. Immunological Characteristics of Colitis Associated with Anti-CTLA-4 Antibody Therapy. Cancer Invest 2017; 35: 443-455 [PMID: 28548891 DOI: 10.1080/07357907.2017.1324032]
- 94 Messmer M, Upreti S, Tarabishy Y, Mazumder N, Chowdhury R, Yarchoan M, Holdhoff M. Ipilimumab-Induced Enteritis without Colitis: A New Challenge. Case Rep Oncol 2016; 9: 705-713 [PMID: 27920706 DOI: 10.1159/000452403]
- Shah R, Witt D, Asif T, Mir FF. Ipilimumab as a Cause of Severe Pan-Colitis and Colonic Perforation. 95 Cureus 2017; 9: e1182 [PMID: 28533998 DOI: 10.7759/cureus.1182]
- Abu-Sbeih H, Wang Y. Management Considerations for Immune Checkpoint Inhibitor-Induced 96 Enterocolitis Based on Management of Inflammatory Bowel Disease. Inflamm Bowel Dis 2020; 26: 662-668 [PMID: 31560045 DOI: 10.1093/ibd/izz212]
- Fukumoto T, Fujiwara S, Tajima S, Tamesada Y, Sakaguchi M, Oka M, Nishigori C. Infliximab for severe colitis associated with nivolumab followed by ipilimumab. J Dermatol 2018; 45: e1-e2 [PMID: 2889124] DOI: 10.1111/1346-8138.140341
- Jain A, Lipson EJ, Sharfman WH, Brant SR, Lazarev MG. Colonic ulcerations may predict steroid-98 refractory course in patients with ipilimumab-mediated enterocolitis. World J Gastroenterol 2017; 23: 2023-2028 [PMID: 28373768 DOI: 10.3748/wjg.v23.i11.2023]
- 99 Hsieh AH, Ferman M, Brown MP, Andrews JM. Vedolizumab: a novel treatment for ipilimumab-induced colitis. BMJ Case Rep 2016; 2016 [PMID: 27539137 DOI: 10.1136/bcr-2016-216641]
- 100 Ribas A, Puzanov I, Dummer R, Schadendorf D, Hamid O, Robert C, Hodi FS, Schachter J, Pavlick AC, Lewis KD, Cranmer LD, Blank CU, O'Day SJ, Ascierto PA, Salama AK, Margolin KA, Loquai C, Eigentler TK, Gangadhar TC, Carlino MS, Agarwala SS, Moschos SJ, Sosman JA, Goldinger SM, Shapira-Frommer R, Gonzalez R, Kirkwood JM, Wolchok JD, Eggermont A, Li XN, Zhou W, Zernhelt AM, Lis J, Ebbinghaus S, Kang SP, Daud A. Pembrolizumab versus investigator-choice chemotherapy for ipilimumabrefractory melanoma (KEYNOTE-002): a randomised, controlled, phase 2 trial. Lancet Oncol 2015; 16: 908-918 [PMID: 26115796 DOI: 10.1016/S1470-2045(15)00083-2]
- 101 Weber J, Thompson JA, Hamid O, Minor D, Amin A, Ron I, Ridolfi R, Assi H, Maraveyas A, Berman D, Siegel J, O'Day SJ. A randomized, double-blind, placebo-controlled, phase II study comparing the tolerability and efficacy of ipilimumab administered with or without prophylactic budesonide in patients with unresectable stage III or IV melanoma. Clin Cancer Res 2009; 15: 5591-5598 [PMID: 19671877 DOI: 10.1158/1078-0432.CCR-09-1024]
- 102 Pollack MH, Betof A, Dearden H, Rapazzo K, Valentine I, Brohl AS, Ancell KK, Long GV, Menzies AM, Eroglu Z, Johnson DB, Shoushtari AN. Safety of resuming anti-PD-1 in patients with immune-related adverse events (irAEs) during combined anti-CTLA-4 and anti-PD1 in metastatic melanoma. Ann Oncol 2018; 29: 250-255 [PMID: 29045547 DOI: 10.1093/annonc/mdx642]
- Gielisse EA, de Boer NK. Ipilimumab in a patient with known Crohn's disease: to give or not to give? J 103 Crohns Colitis 2014; 8: 1742 [PMID: 25154682 DOI: 10.1016/j.crohns.2014.08.002]
- 104 Hodi FS, Chesney J, Pavlick AC, Robert C, Grossmann KF, McDermott DF, Linette GP, Meyer N, Giguere JK, Agarwala SS, Shaheen M, Ernstoff MS, Minor DR, Salama AK, Taylor MH, Ott PA, Horak C, Gagnier P, Jiang J, Wolchok JD, Postow MA. Combined nivolumab and ipilimumab versus ipilimumab alone in patients with advanced melanoma: 2-year overall survival outcomes in a multicentre, randomised, controlled, phase 2 trial. Lancet Oncol 2016; 17: 1558-1568 [PMID: 27622997 DOI: 10.1016/S1470-2045(16)30366-71
- Yamauchi R, Araki T, Mitsuyama K, Tokito T, Ishii H, Yoshioka S, Kuwaki K, Mori A, Yoshimura T, 105 Tsuruta O, Torimura T. The characteristics of nivolumab-induced colitis: an evaluation of three cases and a literature review. BMC Gastroenterol 2018; 18: 135 [PMID: 30170560 DOI: 10.1186/s12876-018-0864-1]
- 106 Bellaguarda E, Hanauer S. Checkpoint Inhibitor-Induced Colitis. Am J Gastroenterol 2020; 115: 202-210 [PMID: 31922959 DOI: 10.14309/ajg.00000000000497]
- 107 Wang Y, Wiesnoski DH, Helmink BA, Gopalakrishnan V, Choi K, DuPont HL, Jiang ZD, Abu-Sbeih H, Sanchez CA, Chang CC, Parra ER, Francisco-Cruz A, Raiu GS, Stroehlein JR, Campbell MT, Gao J. Subudhi SK, Maru DM, Blando JM, Lazar AJ, Allison JP, Sharma P, Tetzlaff MT, Wargo JA, Jenq RR. Fecal microbiota transplantation for refractory immune checkpoint inhibitor-associated colitis. Nat Med 2018; 24: 1804-1808 [PMID: 30420754 DOI: 10.1038/s41591-018-0238-9]
- 108 Abu-Sbeih H. Wang Y. Gut Microbiome and Immune Checkpoint Inhibitor-Induced Enterocolitis. Dig Dis Sci 2020; 65: 797-799 [PMID: 32040664 DOI: 10.1007/s10620-020-06103-x]
- 109 Feng X, Su Y, Jiang J, Li N, Ding W, Wang Z, Hu X, Zhu W, Li J. Changes in fecal and colonic mucosal microbiota of patients with refractory constipation after a subtotal colectomy. Am Surg 2015; 81: 198-206 [PMID: 25642885]
- 110 Margagnoni G, Aratari A, Mangone M, Moretti A, Spagnolo A, Fascì Spurio F, Luchetti R, Papi C. Natural history of ileo-caecal Crohn's disease after surgical resection. A long term study. Minerva Gastroenterol Dietol 2011; 57: 335-344 [PMID: 22105722]
- Devine AA, Gonzalez A, Speck KE, Knight R, Helmrath M, Lund PK, Azcarate-Peril MA. Impact of 111 ileocecal resection and concomitant antibiotics on the microbiome of the murine jejunum and colon. PLoS One 2013; 8: e73140 [PMID: 24015295 DOI: 10.1371/journal.pone.0073140]



- Shogan BD, Belogortseva N, Luong PM, Zaborin A, Lax S, Bethel C, Ward M, Muldoon JP, Singer M, An 112 G, Umanskiy K, Konda V, Shakhsheer B, Luo J, Klabbers R, Hancock LE, Gilbert J, Zaborina O, Alverdy JC. Collagen degradation and MMP9 activation by Enterococcus faecalis contribute to intestinal anastomotic leak. Sci Transl Med 2015; 7: 286ra68 [PMID: 25947163 DOI: 10.1126/scitranslmed.3010658]
- 113 Guyton K, Alverdy JC. The gut microbiota and gastrointestinal surgery. Nat Rev Gastroenterol Hepatol 2017; 14: 43-54 [PMID: 27729657 DOI: 10.1038/nrgastro.2016.139]
- Machiels K, Sabino J, Vandermosten L, Joossens M, Arijs I, de Bruyn M, Eeckhaut V, Van Assche G, 114 Ferrante M, Verhaegen J, Van Steen K, Van Immerseel F, Huys G, Verbeke K, Wolthuis A, de Buck Van Overstraeten A, D'Hoore A, Rutgeerts P, Vermeire S. Specific members of the predominant gut microbiota predict pouchitis following colectomy and IPAA in UC. Gut 2017; 66: 79-88 [PMID: 26423113 DOI: 10.1136/gutjnl-2015-309398]
- Rubenstein J, Sherif A, Appelman H, Chey WD. Ulcerative colitis associated enteritis: is ulcerative colitis 115 always confined to the colon? J Clin Gastroenterol 2004; 38: 46-51 [PMID: 14679327 DOI: 10.1097/00004836-200401000-00011
- Rush B, Berger L, Rosenfeld G, Bressler B. Tacrolimus Therapy for Ulcerative Colitis-Associated Post-116 Colectomy Enteritis. ACG Case Rep J 2014; 2: 33-35 [PMID: 26157899 DOI: 10.14309/crj.2014.76]
- 117 Yang Y, Liu Y, Zheng W, Zhou W, Wu B, Sun X, Chen W, Guo T, Li X, Yang H, Qian J, Li Y. A literature review and case report of severe and refractory post-colectomy enteritis. BMC Gastroenterol 2019; 19: 61 [PMID: 31023233 DOI: 10.1186/s12876-019-0974-4]
- 118 Annese V, Caruso N, Bisceglia M, Lombardi G, Clemente R, Modola G, Tardio B, Villani MR, Andriulli A. Fatal ulcerative panenteritis following colectomy in a patient with ulcerative colitis. Dig Dis Sci 1999; 44: 1189-1195 [PMID: 10389695 DOI: 10.1023/a:1026688526551]
- Reese GE, Lovegrove RE, Tilney HS, Yamamoto T, Heriot AG, Fazio VW, Tekkis PP. The effect of 119 Crohn's disease on outcomes after restorative proctocolectomy. Dis Colon Rectum 2007; 50: 239-250 [PMID: 17180251 DOI: 10.1007/s10350-006-0777-x]
- 120 Shen B, Patel S, Lian L. Natural history of Crohn's disease in patients who underwent intentional restorative proctocolectomy with ileal pouch-anal anastomosis. Aliment Pharmacol Ther 2010; 31: 745-753 [PMID: 20047579 DOI: 10.1111/j.1365-2036.2009.04227.x]
- 121 Armuzzi A, Felice C, Papa A, Marzo M, Pugliese D, Andrisani G, Federico F, De Vitis I, Rapaccini GL, Guidi L. Prevention of postoperative recurrence with azathioprine or infliximab in patients with Crohn's disease: an open-label pilot study. J Crohns Colitis 2013; 7: e623-e629 [PMID: 23810678 DOI: 10.1016/j.crohns.2013.04.020
- Philpott J, Ashburn J, Shen B. Efficacy of Vedolizumab in Patients with Antibiotic and Anti-tumor Necrosis Alpha Refractory Pouchitis. Inflamm Bowel Dis 2017; 23: E5-E6 [PMID: 27930413 DOI: 10.1097/MIB.000000000000992
- Singh S, Ding NS, Mathis KL, Dulai PS, Farrell AM, Pemberton JH, Hart AL, Sandborn WJ, Loftus EV Jr. 123 Systematic review with meta-analysis: faecal diversion for management of perianal Crohn's disease. Aliment Pharmacol Ther 2015; 42: 783-792 [PMID: 26264359 DOI: 10.1111/apt.13356]
- 124 Wu XR, Liu XL, Katz S, Shen B. Pathogenesis, diagnosis, and management of ulcerative proctitis, chronic radiation proctopathy, and diversion proctitis. Inflamm Bowel Dis 2015; 21: 703-715 [PMID: 25687266 DOI: 10.1097/MIB.00000000000227]
- 125 Du P, Sun C, Ashburn J, Wu X, Philpott J, Remzi FH, Shen B. Risk factors for Crohn's disease of the neosmall intestine in ulcerative colitis patients with total proctocolectomy and primary or secondary ileostomies. J Crohns Colitis 2015; 9: 170-176 [PMID: 25518056 DOI: 10.1093/ecco-jcc/jju014]
- Shen B. Crohn's disease of the ileal pouch: reality, diagnosis, and management. Inflamm Bowel Dis 2009; 126 15: 284-294 [PMID: 18816633 DOI: 10.1002/ibd.20661]
- Pareek M, Schauer PR, Kaplan LM, Leiter LA, Rubino F, Bhatt DL. Metabolic Surgery: Weight Loss, 127 Diabetes, and Beyond. J Am Coll Cardiol 2018; 71: 670-687 [PMID: 29420964 DOI: 10.1016/j.jacc.2017.12.014
- Flum DR, Kwon S, MacLeod K, Wang B, Alfonso-Cristancho R, Garrison LP, Sullivan SD; Bariatric 128 Obesity Outcome Modeling Collaborative. The use, safety and cost of bariatric surgery before and after Medicare's national coverage decision. Ann Surg 2011; 254: 860-865 [PMID: 21975317 DOI: 10.1097/SLA.0b013e31822f2101]
- 129 Nguyen NT, Vu S, Kim E, Bodunova N, Phelan MJ. Trends in utilization of bariatric surgery, 2009-2012. Surg Endosc 2016; 30: 2723-2727 [PMID: 26659240 DOI: 10.1007/s00464-015-4535-9]
- Ahn LB, Huang CS, Forse RA, Hess DT, Andrews C, Farraye FA. Crohn's disease after gastric bypass 130 surgery for morbid obesity: is there an association? Inflamm Bowel Dis 2005; 11: 622-624 [PMID: 15905716 DOI: 10.1097/01.mib.0000165113.33557.3a]
- 131 **Dodell GB** Albu JB. Attia L. McGinty J. Pi-Sunver FX. Laferrère B. The bariatric surgery patient: lost to follow-up; from morbid obesity to severe malnutrition. Endocr Pract 2012; 18: e21-e25 [PMID: 22138075 DOI: 10.4158/EP11200.CR]
- Korelitz BI, Sonpal N, Schneider J, Swaminath A, Felder J, Roslin M, Aronoff J. Obesity/Bariatric Surgery 132 and Crohn's Disease. J Clin Gastroenterol 2018; 52: 50-54 [PMID: 28489647 DOI: 10.1097/MCG.00000000000765]
- Braga Neto MB, Gregory M, Ramos GP, Loftus EV Jr, Ciorba MA, Bruining DH, Bazerbachi F, Abu 133 Dayyeh BK, Kushnir VM, Shah M, Collazo-Clavell ML, Raffals LE, Deepak P. De-novo Inflammatory Bowel Disease After Bariatric Surgery: A Large Case Series. J Crohns Colitis 2018; 12: 452-457 [PMID: 29272375 DOI: 10.1093/ecco-jcc/jjx177]
- 134 Bernstein GR. Pickett-Blakely O. De Novo Inflammatory Bowel Disease After Bariatric Surgery: A Case Series and Literature Review. Dig Dis Sci 2017; 62: 817-820 [PMID: 28012102 DOI: 10.1007/s10620-016-4412-y
- 135 Cañete F, Mañosa M, Clos A, Cabré E, Domènech E. Review article: the relationship between obesity, bariatric surgery, and inflammatory bowel disease. Aliment Pharmacol Ther 2018; 48: 807-816 [PMID: 30178869 DOI: 10.1111/apt.14956]
- Ungaro R, Fausel R, Chang HL, Chang S, Chen LA, Nakad A, El Nawar A, Prytz Berset I, Axelrad J, 136



Lawlor G, Atreja A, Roque Ramos L, Torres J, Colombel JF. Bariatric surgery is associated with increased risk of new-onset inflammatory bowel disease: case series and national database study. Aliment Pharmacol Ther 2018; 47: 1126-1134 [PMID: 29512187 DOI: 10.1111/apt.14569]

- 137 Maple JT, Pearson RK, Murray JA, Kelly DG, Lara LF, Fan AC. Silent celiac disease activated by pancreaticoduodenectomy. Dig Dis Sci 2007; 52: 2140-2144 [PMID: 17373587 DOI: 10.1007/s10620-006-9598-y]
- Bai J, Moran C, Martinez C, Niveloni S, Crosetti E, Sambuelli A, Boerr L. Celiac sprue after surgery of the 138 upper gastrointestinal tract. Report of 10 patients with special attention to diagnosis, clinical behavior, and follow-up. J Clin Gastroenterol 1991; 13: 521-524 [PMID: 1744387 DOI: 10.1097/00004836-199110000-00009]
- Shen L, Lian L, Goldblum JR, Remzi FH. Development of de novo celiac disease after restorative 139 proctocolectomy and ileal pouch-anal anastomosis. Inflamm Bowel Dis 2009; 15: 1131-1132 [PMID: 19067415 DOI: 10.1002/ibd.207911
- Guo Y, Huang ZP, Liu CQ, Qi L, Sheng Y, Zou DJ. Modulation of the gut microbiome: a systematic 140 review of the effect of bariatric surgery. Eur J Endocrinol 2018; 178: 43-56 [PMID: 28916564 DOI: 10.1530/EJE-17-0403
- 141 Jahansouz C, Staley C, Bernlohr DA, Sadowsky MJ, Khoruts A, Ikramuddin S. Sleeve gastrectomy drives persistent shifts in the gut microbiome. Surg Obes Relat Dis 2017; 13: 916-924 [PMID: 28279578 DOI: 10.1016/j.soard.2017.01.003]
- Margulies SL, Kurian D, Elliott MS, Han Z. Vitamin D deficiency in patients with intestinal malabsorption 142 syndromes--think in and outside the gut. J Dig Dis 2015; 16: 617-633 [PMID: 26316334 DOI: 10.1111/1751-2980.122831
- Aminian A, Andalib A, Ver MR, Corcelles R, Schauer PR, Brethauer SA. Outcomes of Bariatric Surgery in 143 Patients with Inflammatory Bowel Disease. Obes Surg 2016; 26: 1186-1190 [PMID: 26420765 DOI: 10.1007/s11695-015-1909-y]
- Aelfers S, Janssen IMC, Aarts EO, Smids C, Groenen MJ, Berends FJ. Inflammatory Bowel Disease Is Not 144 a Contraindication for Bariatric Surgery. Obes Surg 2018; 28: 1681-1687 [PMID: 29282629 DOI: 10.1007/s11695-017-3076-9
- 145 Bazerbachi F, Sawas T, Vargas EJ, Haffar S, Deepak P, Kisiel JB, Loftus EV Jr, Abu Dayyeh BK. Bariatric Surgery Is Acceptably Safe in Obese Inflammatory Bowel Disease Patients: Analysis of the Nationwide Inpatient Sample. Obes Surg 2018; 28: 1007-1014 [PMID: 29019151 DOI: 10.1007/s11695-017-2955-4
- Shoar S, Shahabuddin Hoseini S, Naderan M, Mahmoodzadeh H, Ying Man F, Shoar N, Hosseini M, 146 Bagheri-Hariri S. Bariatric surgery in morbidly obese patients with inflammatory bowel disease: A systematic review. Surg Obes Relat Dis 2017; 13: 652-659 [PMID: 27986584 DOI: 10.1016/j.soard.2016.10.017]
- Yang JY, Sarwal MM. Transplant genetics and genomics. Nat Rev Genet 2017; 18: 309-326 [PMID: 147 28286337 DOI: 10.1038/nrg.2017.12]
- 148 Shi X, Han W, Ding J. The impact of human leukocyte antigen mismatching on graft survival and mortality in adult renal transplantation: A protocol for a systematic review and meta-analysis. Medicine (Baltimore) 2017; 96: e8899 [PMID: 29245253 DOI: 10.1097/MD.00000000008899]
- 149 Seekatz AM, Aas J, Gessert CE, Rubin TA, Saman DM, Bakken JS, Young VB. Recovery of the gut microbiome following fecal microbiota transplantation. mBio 2014; 5: e00893-e00814 [PMID: 24939885 DOI: 10.1128/mBio.00893-141
- 150 Forbes JD, Van Domselaar G, Bernstein CN. Microbiome Survey of the Inflamed and Noninflamed Gut at Different Compartments Within the Gastrointestinal Tract of Inflammatory Bowel Disease Patients. Inflamm Bowel Dis 2016; 22: 817-825 [PMID: 26937623 DOI: 10.1097/MIB.00000000000684]
- 151 Shah R, Cope JL, Nagy-Szakal D, Dowd S, Versalovic J, Hollister EB, Kellermayer R. Composition and function of the pediatric colonic mucosal microbiome in untreated patients with ulcerative colitis. Gut Microbes 2016: 7: 384-396 [PMID: 27217061 DOI: 10.1080/19490976.2016.1190073]
- 152 Klein AK, Patel DD, Gooding ME, Sempowski GD, Chen BJ, Liu C, Kurtzberg J, Haynes BF, Chao NJ. T-Cell recovery in adults and children following umbilical cord blood transplantation. Biol Blood Marrow Transplant 2001; 7: 454-466 [PMID: 11569891 DOI: 10.1016/s1083-8791(01)80013-6]
- 153 Herrera AF, Soriano G, Bellizzi AM, Hornick JL, Ho VT, Ballen KK, Baden LR, Cutler CS, Antin JH, Soiffer RJ, Marty FM. Cord colitis syndrome in cord-blood stem-cell transplantation. N Engl J Med 2011; 365: 815-824 [PMID: 21879899 DOI: 10.1056/NEJMoa1104959]
- Shimoji S, Kato K, Eriguchi Y, Takenaka K, Iwasaki H, Miyamoto T, Oda Y, Akashi K, Teshima T. 154 Evaluating the association between histological manifestations of cord colitis syndrome with GVHD. Bone Marrow Transplant 2013; 48: 1249-1252 [PMID: 23749110 DOI: 10.1038/bmt.2013.44]
- Gupta NK, Masia R. Cord colitis syndrome: a cause of granulomatous inflammation in the upper and lower 155 gastrointestinal tract. Am J Surg Pathol 2013; 37: 1109-1113 [PMID: 23715165 DOI: 10.1097/PAS.0b013e31828a827a]
- Bhatt AS, Freeman SS, Herrera AF, Pedamallu CS, Gevers D, Duke F, Jung J, Michaud M, Walker BJ, 156 Young S, Earl AM, Kostic AD, Ojesina AI, Hasserjian R, Ballen KK, Chen YB, Hobbs G, Antin JH, Soiffer RJ, Baden LR, Garrett WS, Hornick JL, Marty FM, Meyerson M. Sequence-based discovery of Bradyrhizobium enterica in cord colitis syndrome. N Engl J Med 2013; 369: 517-528 [PMID: 23924002 DOI: 10.1056/NEJMoa12111151
- Nepal S, Navaneethan U, Bennett AE, Shen B. De novo inflammatory bowel disease and its mimics after 157 organ transplantation. Inflamm Bowel Dis 2013; 19: 1518-1527 [PMID: 23656896 DOI: 10.1097/MIB.0b013e3182813365]
- Wörns MA, Lohse AW, Neurath MF, Croxford A, Otto G, Kreft A, Galle PR, Kanzler S. Five cases of de 158 novo inflammatory bowel disease after orthotopic liver transplantation. Am J Gastroenterol 2006; 101: 1931-1937 [PMID: 16790037 DOI: 10.1111/j.1572-0241.2006.00624.x]
- Singh S, Loftus EV Jr, Talwalkar JA. Inflammatory bowel disease after liver transplantation for primary 159 sclerosing cholangitis. Am J Gastroenterol 2013; 108: 1417-1425 [PMID: 23896954 DOI:



10.1038/ajg.2013.163]

- 160 Bajer L, Slavcev A, Macinga P, Sticova E, Brezina J, Roder M, Janousek R, Trunecka P, Spicak J, Drastich P. Risk of recurrence of primary sclerosing cholangitis after liver transplantation is associated with de novo inflammatory bowel disease. World J Gastroenterol 2018; 24: 4939-4949 [PMID: 30487703 DOI: 10.3748/wjg.v24.i43.4939]
- **Riley TR.** Schoen RE. Lee RG. Rakela J. A case series of transplant recipients who despite 161 immunosuppression developed inflammatory bowel disease. Am J Gastroenterol 1997; 92: 279-282 [PMID: 90402061
- Pittman ME, Jessurun J, Yantiss RK. Differentiating Posttransplant Inflammatory Bowel Disease and 162 Other Colitides in Renal Transplant Patients. Am J Surg Pathol 2017; 41: 1666-1674 [PMID: 28786879 DOI: 10.1097/PAS.000000000000921]
- 163 Bajer L, Wohl P, Drastich P. PSC-IBD: specific phenotype of inflammatory bowel disease associated with primary sclerosing cholangitis. Vnitr Lek Summer; 64: 659-664 [PMID: 30223664]
- 164 Mogl MT, Baumgart DC, Fischer A, Pratschke J, Pascher A. Immunosuppression following liver transplantation and the course of inflammatory bowel disease - a case control study. Z Gastroenterol 2018; 56: 117-127 [PMID: 29212098 DOI: 10.1055/s-0043-117183]
- 165 Karolewska-Bochenek K, Grzesiowski P, Banaszkiewicz A, Gawronska A, Kotowska M, Dziekiewicz M, Albrecht P, Radzikowski A, Lazowska-Przeorek I. A Two-Week Fecal Microbiota Transplantation Course in Pediatric Patients with Inflammatory Bowel Disease. Adv Exp Med Biol 2018; 1047: 81-87 [PMID: 29151253 DOI: 10.1007/5584_2017_123]
- Brandt LJ, Aroniadis OC, Mellow M, Kanatzar A, Kelly C, Park T, Stollman N, Rohlke F, Surawicz C. 166 Long-term follow-up of colonoscopic fecal microbiota transplant for recurrent Clostridium difficile infection. Am J Gastroenterol 2012; 107: 1079-1087 [PMID: 22450732 DOI: 10.1038/ajg.2012.60]
- Jiang ZD, Ajami NJ, Petrosino JF, Jun G, Hanis CL, Shah M, Hochman L, Ankoma-Sey V, DuPont AW, 167 Wong MC, Alexander A, Ke S, DuPont HL. Randomised clinical trial: faecal microbiota transplantation for recurrent Clostridum difficile infection - fresh, or frozen, or lyophilised microbiota from a small pool of healthy donors delivered by colonoscopy. Aliment Pharmacol Ther 2017; 45: 899-908 [PMID: 28220514 DOI: 10.1111/apt.139691
- 168 Khoruts A, Rank KM, Newman KM, Viskocil K, Vaughn BP, Hamilton MJ, Sadowsky MJ. Inflammatory Bowel Disease Affects the Outcome of Fecal Microbiota Transplantation for Recurrent Clostridium difficile Infection. Clin Gastroenterol Hepatol 2016; 14: 1433-1438 [PMID: 26905904 DOI: 10.1016/j.cgh.2016.02.018
- Colman RJ, Rubin DT. Fecal microbiota transplantation as therapy for inflammatory bowel disease: a 169 systematic review and meta-analysis. J Crohns Colitis 2014; 8: 1569-1581 [PMID: 25223604 DOI: 10.1016/j.crohns.2014.08.006
- 170 Costello SP, Soo W, Bryant RV, Jairath V, Hart AL, Andrews JM. Systematic review with meta-analysis: faecal microbiota transplantation for the induction of remission for active ulcerative colitis. Aliment Pharmacol Ther 2017; 46: 213-224 [PMID: 28612983 DOI: 10.1111/apt.14173]
- 171 Izquierdo Romero M, Varela Trastoy P, Mancebo Mata A. Fecal transplantation as a treatment for Clostridium difficile infection in patients with ulcerative colitis. Rev Esp Enferm Dig 2017; 109: 670 [PMID: 28747056 DOI: 10.17235/reed.2017.4941/2017]
- 172 Bak SH. Choi HH. Lee J. Kim MH. Lee YH. Kim JS. Cho YS. Fecal microbiota transplantation for refractory Crohn's disease. Intest Res 2017; 15: 244-248 [PMID: 28522956 DOI: 10.5217/ir.2017.15.2.244]
- Lan N, Ashburn J, Shen B. Fecal microbiota transplantation for Clostridium difficile infection in patients 173 with ileal pouches. Gastroenterol Rep (Oxf) 2017; 5: 200-207 [PMID: 28852524 DOI: 10.1093/gastro/gox018]
- 174 Khanna S, Vazquez-Baeza Y, González A, Weiss S, Schmidt B, Muñiz-Pedrogo DA, Rainey JF 3rd, Kammer P. Nelson H. Sadowsky M. Khoruts A. Farrugia SL, Knight R. Pardi DS, Kashyap PC, Changes in microbial ecology after fecal microbiota transplantation for recurrent C. difficile infection affected by underlying inflammatory bowel disease. Microbiome 2017; 5: 55 [PMID: 28506317 DOI: 10.1186/s40168-017-0269-3
- 175 Pai N, Popov J. Protocol for a randomised, placebo-controlled pilot study for assessing feasibility and efficacy of faecal microbiota transplantation in a paediatric ulcerative colitis population: PediFETCh trial. BMJ Open 2017; 7: e016698 [PMID: 28827258 DOI: 10.1136/bmjopen-2017-016698]
- 176 Hohmann EL, Ananthakrishnan AN, Deshpande V. Case Records of the Massachusetts General Hospital. Case 25-2014. A 37-year-old man with ulcerative colitis and bloody diarrhea. N Engl J Med 2014; 371: 668-675 [PMID: 25119613 DOI: 10.1056/NEJMcpc1400842]
- 177 Wang S, Xu M, Wang W, Cao X, Piao M, Khan S, Yan F, Cao H, Wang B. Systematic Review: Adverse Events of Fecal Microbiota Transplantation. PLoS One 2016; 11: e0161174 [PMID: 27529553 DOI: 10.1371/journal.pone.0161174]
- 178 De Leon LM, Watson JB, Kelly CR. Transient flare of ulcerative colitis after fecal microbiota transplantation for recurrent Clostridium difficile infection. Clin Gastroenterol Hepatol 2013; 11: 1036-1038 [PMID: 23669309 DOI: 10.1016/j.cgh.2013.04.045]
- Kelly CR, Ihunnah C, Fischer M, Khoruts A, Surawicz C, Afzali A, Aroniadis O, Barto A, Borody T, 179 Giovanelli A, Gordon S, Gluck M, Hohmann EL, Kao D, Kao JY, McQuillen DP, Mellow M, Rank KM, Rao K, Ray A, Schwartz MA, Singh N, Stollman N, Suskind DL, Vindigni SM, Youngster I, Brandt L. Fecal microbiota transplant for treatment of Clostridium difficile infection in immunocompromised patients. Am J Gastroenterol 2014; 109: 1065-1071 [PMID: 24890442 DOI: 10.1038/ajg.2014.133]
- 180 Qazi T, Amaratunga T, Barnes EL, Fischer M, Kassam Z, Allegretti JR. The risk of inflammatory bowel disease flares after fecal microbiota transplantation: Systematic review and meta-analysis. Gut Microbes 2017; 8: 574-588 [PMID: 28723262 DOI: 10.1080/19490976.2017.1353848]
- 181 Woodworth MH, Carpentieri C, Sitchenko KL, Kraft CS. Challenges in fecal donor selection and screening for fecal microbiota transplantation: A review. Gut Microbes 2017; 8: 225-237 [PMID: 28129018 DOI: 10.1080/19490976.2017.1286006]
- Paramsothy S, Borody TJ, Lin E, Finlayson S, Walsh AJ, Samuel D, van den Bogaerde J, Leong RW, 182



Connor S, Ng W, Mitchell HM, Kaakoush N, Kamm MA. Donor Recruitment for Fecal Microbiota Transplantation. Inflamm Bowel Dis 2015; 21: 1600-1606 [PMID: 26070003 DOI: 10.1097/MIB.000000000000405]

Kelly BJ, Tebas P. Clinical Practice and Infrastructure Review of Fecal Microbiota Transplantation for 183 Clostridium difficile Infection. Chest 2018; 153: 266-277 [PMID: 28923757 DOI: 10.1016/j.chest.2017.09.002]





Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

