World Journal of *Clinical Cases*

World J Clin Cases 2024 December 16; 12(35): 6754-6863





Published by Baishideng Publishing Group Inc

W J C C World Journal of Clinical Cases

Contents

Thrice Monthly Volume 12 Number 35 December 16, 2024

EDITORIAL

6754	Update on the aetiopathogenesis of obstructive sleep apnea: Role of inflammatory and immune mediated mechanisms
	Nag DS, Varghese K, Swain A, Patel R, Sahu S, Sam M
6760	Intensive care unit-acquired weakness: Unveiling significant risk factors and preemptive strategies through machine learning
	He XY, Zhao YH, Wan QW, Tang FS
6764	Advancing oral cancer care: Insights from Tongluo Jiedu prescription
	Cheng CY, Hao WR, Liu JC, Cheng TH
6770	Effects of atrial septal defects on the cardiac conduction system
	Kang JH, Wu HY, Long WJ
6775	Periodontitis and chronic kidney disease: A bidirectional relationship based on inflammation and oxidative stress
	Martínez Nieto M, De Leon Rodríguez ML, Anaya Macias RDC, Lomelí Martínez SM
6782	Cytokine release syndrome induced by anti-programmed death-1 treatment in a psoriasis patient: A dark side of immune checkpoint inhibitors
	Maldonado-García JL, Fragozo A, Pavón L
	REVIEW
6791	Acellular dermal matrices in reconstructive surgery; history, current implications and future perspectives for surgeons
	Dilek ÖF, Sevim KZ, Dilek ON
	ORIGINAL ARTICLE
	Retrospective Study
6808	Comprehensive epidemiological assessment of trauma incidents at a level I trauma center
	Su ZY, Wei H, Wang WN, Lin YF, He YL, Liu Y, Lin RB, Liu YT, Michael N
	SYSTEMATIC REVIEWS
6815	Gut microbiota changes associated with frailty in older adults: A systematic review of observational studies
	Wen NN, Sun LW, Geng Q, Zheng GH



World	Journal	of Clinical	Cases
,,	0000000000000	of connear	Cubeb

Contents

Thrice Monthly Volume 12 Number 35 December 16, 2024

CASE REPORT

6826 psk1 virulence gene-induced pulmonary and systemic tuberculosis in a young woman with normal immune function: A case report

Wu F, Yang B, Xiao Y, Ren LL, Chen HY, Hu XL, Pan YY, Chen YS, Li HR

6834 Rare primary gastric peripheral T-cell lymphoma not otherwise specified: A case report Jang HR, Lee K, Lim KH

6840 Cat scratch disease in children with nocturnal fever: A case report Yin QL, Liu YQ, Zhang HM, Zhang YL, Qi SM, Wen JQ, Zhang WH

LETTER TO THE EDITOR

- 6848 Understanding network meta-analysis Au SCL
- 6851 Effects of foot reflexology on disease He MY, Ud Din MJ, Xu HF, Wang SY, Ying GH, Qian H, Wu B, Qi HD, Wang X, Zhang G
- 6855 Clinical landscape and treatment of acute non-variceal upper gastrointestinal bleeding: Insights from a high-volume center in Shaanxi, China Improta L
- 6859 Role of high-dose amoxicillin dual therapy for Helicobacter pylori eradication in an Irish cohort: A prospective study

Palmirotta R, Cafiero C, Colella M



Contents

Thrice Monthly Volume 12 Number 35 December 16, 2024

ABOUT COVER

Peer Reviewer of World Journal of Clinical Cases, Muhamad Zakaria Brimo Alsaman, MD, Associate Chief Physician, Department of Vascular Surgery, Al-Razi Hospital, Aleppo, Syria; Faculty of Medicine, University of Aleppo, Aleppo 12212, Syria. dr.zakariabrimo@gmail.com

AIMS AND SCOPE

The primary aim of World Journal of Clinical Cases (WJCC, World J Clin Cases) is to provide scholars and readers from various fields of clinical medicine with a platform to publish high-quality clinical research articles and communicate their research findings online.

WJCC mainly publishes articles reporting research results and findings obtained in the field of clinical medicine and covering a wide range of topics, including case control studies, retrospective cohort studies, retrospective studies, clinical trials studies, observational studies, prospective studies, randomized controlled trials, randomized clinical trials, systematic reviews, meta-analysis, and case reports.

INDEXING/ABSTRACTING

The WJCC is now abstracted and indexed in PubMed, PubMed Central, Reference Citation Analysis, China Science and Technology Journal Database, and Superstar Journals Database. The 2024 Edition of Journal Citation Reports® cites the 2023 journal impact factor (JIF) for WJCC as 1.0; JIF without journal self cites: 0.9; 5-year JIF: 1.1; JIF Rank: 168/325 in medicine, general and internal; JIF Quartile: Q3; and 5-year JIF Quartile: Q3.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Lei Zhang, Production Department Director: Si Zhao, Cover Editor: Jin-Lei Wang.

NAME OF JOURNAL World Journal of Clinical Cases	INSTRUCTIONS TO AUTHORS https://www.wignet.com/bpg/gerinfo/204
ISSN ISSN 2307-8960 (online)	GUIDELINES FOR ETHICS DOCUMENTS
LAUNCH DATE April 16, 2013	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH https://www.wignet.com/bpg/gerinfo/240
FREQUENCY Thrice Monthly	PUBLICATION ETHICS
EDITORS-IN-CHIEF Bao-Gan Peng, Salim Surani, Jerzy Tadeusz Chudek, George Kontogeorgos,	PUBLICATION MISCONDUCT https://www.wignet.com/bpg/gerinfo/208
Maurizio Serati EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
https://www.wjgnet.com/2307-8960/editorialboard.htm PUBLICATION DATE	https://www.wjgnet.com/bpg/gerinfo/242 STEPS FOR SUBMITTING MANUSCRIPTS
December 16, 2024	https://www.wjgnet.com/bpg/GerInfo/239
© 2024 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2024 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: office@baishideng.com https://www.wjgnet.com



W J C C World Journal C Clinical Cases

World Journal of

Submit a Manuscript: https://www.f6publishing.com

World J Clin Cases 2024 December 16; 12(35): 6815-6825

DOI: 10.12998/wjcc.v12.i35.6815

ISSN 2307-8960 (online)

SYSTEMATIC REVIEWS

Gut microbiota changes associated with frailty in older adults: A systematic review of observational studies

Na-Na Wen, Li-Wei Sun, Qian Geng, Guo-Hua Zheng

Specialty type: Medicine, research and experimental

Provenance and peer review: Unsolicited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's classification

Scientific Quality: Grade C Novelty: Grade B Creativity or Innovation: Grade B Scientific Significance: Grade B

P-Reviewer: Duan SL

Received: August 15, 2024 Revised: September 4, 2024 Accepted: September 25, 2024 Published online: December 16, 2024

Processing time: 70 Days and 1.2 Hours



Na-Na Wen, Li-Wei Sun, Qian Geng, Guo-Hua Zheng, College of Nursing and Health Management, Shanghai University of Medicine and Health Sciences, Shanghai 201318, China

Na-Na Wen, Graduate School, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, China

Corresponding author: Guo-Hua Zheng, PhD, Professor, College of Nursing and Health Management, Shanghai University of Medicine and Health Sciences, No. 279 Zhouzhu Road, Pudong New District, Shanghai 201318, China. zhenggh@sumhs.edu.cn

Abstract

BACKGROUND

Frailty is a complex aging-related syndrome characterized by a cumulative loss of physiological reserve and increased vulnerability to adverse clinical outcomes, including falls, disability, incapacity and death. While an increasing number of studies suggest that the gut microbiota may play a key role in the pathophysiology of frailty, direct evaluation of the association between gut microbiome alterations and frailty in older adults remains limited.

AIM

To gain insight into gut dysbiosis in frail older adults.

METHODS

Seven electronic databases (China National Knowledge Infrastructure, VIP, SinoMed, Wanfang, PubMed, Web of Science and EMBASE) were searched for articles published before October 31, 2023 to identify observational studies that compared the microbiomes of older adults with and without frailty. The diversity and composition of the gut microbiota were the main outcomes used to analyze the associations of changes in the gut microbiota with frailty in older adults. The quality of the included studies was assessed via the Newcastle-Ottawa Scale and the Agency for Healthcare Research and Quality.

RESULTS

Eleven observational studies with 912 older adults were included in this review. Consistent results revealed a significant difference in the gut microbiota composition between frail and non-frail older adults, with a significant decrease in a diversity and a significant increase in β diversity in frail older adults. The pooled results revealed that at the phylum level, four microbiota (Actinobacteria, Proteobacteria, Verrucomicrobia and Synergistetes) were significantly enriched, and two



WJCC https://www.wjgnet.com

microbiota (*Firmicutes* and *Fusobacteria*) were significantly depleted in frail older adults. At the family level, the results consistently revealed that the abundances of 6 families, most of which belong to the *Actinobacteria* or *Proteobacteria* phylum, were greater in frail than in non-frail older adults. At the genus or species level, consistent results from more than two studies revealed that the abundances of the genera *Prevotella*, *Faecalibacterium*, and *Roseburia* were significantly lower in frail older adults; individual studies revealed that the abundances of some genera or species (*e.g.*, *Megamonas*, *Blautia*, and *Megasphaera*) were significantly lower, whereas those of other genera or species (*e.g.*, *Bifidobacterium*, *Oscillospira*, *Ruminococcus* and *Pyramidobacter*) were significantly greater in frail older adults.

CONCLUSION

This systematic review suggests that changes in the gut microbiota are associated with frailty in older adults, which is commonly reflected by a reduction in beneficial species and an increase in pathogenic species. However, further studies are needed to confirm these findings.

Key Words: Frailty; Gut microbiota; Observational study; Older adults; Systematic review

©The Author(s) 2024. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: A growing number of studies have reported changes in the composition and diversity of the gut microbiota between frail and healthy older adults, suggesting that alterations in the gut microbiota may play a key role in the pathophysiology of frailty; however, direct assessment of the associations between changes in the gut microbiome and frailty in older adults remains limited. This review revealed a significant decrease in α diversity and a significant increase in β diversity in frail older adults compared with non-frail older adults, which was commonly reflected by a reduction in beneficial species and an increase in pathogenic species. This study provides a comprehensive overview of the relationship between changes in the gut microbiota and frailty in older adults and suggests a possible role for the gut microbiota in the pathogenesis of frailty.

Citation: Wen NN, Sun LW, Geng Q, Zheng GH. Gut microbiota changes associated with frailty in older adults: A systematic review of observational studies. *World J Clin Cases* 2024; 12(35): 6815-6825 URL: https://www.wjgnet.com/2307-8960/full/v12/i35/6815.htm DOI: https://dx.doi.org/10.12998/wjcc.v12.i35.6815

INTRODUCTION

Frailty is a complex age-related geriatric syndrome characterized by decreased physiological reserves in the body with decreased anti-stress ability and vulnerability in the face of external stimuli, leading to increased risks of multiple adverse health outcomes, including falls, hospitalization, and even mortality[1,2]. Frailty is common among community-dwelling older adults, with a prevalence ranging from 4% to 59% among those aged 65 years and older and 25% among older adults over 85 years[3]. Current studies have shown that the risk factors associated with frailty in older adults are mainly related to increasing age, lower weight, female sex, living alone, low levels of physical activity, polypharmacy, unhealthy lifestyle, smoking, alcohol consumption, and poor diet. These factors interact and form a cycle to cause chronic malnutrition, inflammation, and disruption of hormone regulation[4-7]. Recently, intestinal dysbacteriosis has been newly identified as a risk factor for frailty in older adults[6,8]. For example, a study of 728 female twins revealed a negative association between gut microbiota α diversity and frailty, with increases in *Eubacterium dolichum* and *Eggerthella lenta* in the frail group[9].

The gut microbiota is a relatively stable community consisting of a large number of bacteria, fungi, and viruses that colonize the human gut. Gut dysbiosis has been shown to contribute to human diseases, including metabolic diseases, neurodegenerative disorders, and chronic inflammatory diseases[10-11]. The pathogenesis of frailty syndrome may involve chronic inflammation, immune activation, and the musculoskeletal system[12]. Many studies have shown that the diversity and composition of the gut microbiota are significantly altered in community-dwelling older adults with frailty, which in turn may play an important role in the development of frailty in community-dwelling older adults. For example, one study suggested that an imbalance in the gut microbiota triggers an inflammatory response, leading to an increase in intestinal permeability and the entry of pathogen-associated antibodies into the circulation[6,13]. However, most of the previous studies on frailty and the gut microbiota have only examined changes in the composition of bacterial species, but the characteristics of the gut microbiota of frail older people are still unclear, although interest in this topic is increasing. Therefore, there is a lack of adequate evidence on changes in the gut microbiota and frailty in older adults. This systematic literature review mainly summarizes the associations between changes in the gut microbiota and frailty in people over 60 years of age.

Zaishidena® WJCC | https://www.wjgnet.com

MATERIALS AND METHODS

Search strategy

Seven databases (PubMed, EMBASE, Web of Science, SinoMed, China National Knowledge Infrastructure, VIP, and Wanfang) were used to search for Chinese and English articles, respectively, using Medical Subject Headings terms or free words (e.g., "gastrointestinal microbiome" or "gut microbiota" or "intestinal microbiomes" or "gastric microbiome" or "enteric bacteria" or "gut microbiome") and ("frailty" or "frailty syndrome" or "frailelder"). The final search began in October 2023, with no publication date restrictions. A summary of the search strategy for the different databases is described in Supplementary Table 1.

Inclusion criteria

Eligible studies were identified according to the following inclusion criteria: (1) Participants were adults over 60 years of age; (2) The profile of the gut microbiota was compared between frail and non-frail older adults; and (3) The primary outcome was the abundance of bacterial phyla, families, genera and species of the human gut microbiota. Studies for which the required data could not be retrieved were excluded.

Study screening, data extraction, and assessment of risk bias

The retrieved records were imported into reference management software (Note Express 3.1) for repeated screening. Two reviewers independently identified the eligibility of the retrieved articles according to the inclusion criteria after the duplicate records were removed. Disagreements were discussed and resolved in consultation with a third reviewer. Data from the eligible studies were extracted by one reviewer via prepared data extraction tables and checked by another reviewer. The information extracted included study design, participants, methodological characteristics, sample size, outcomes, and measurement methods. The risk of bias for the eligible studies was assessed via the Newcastle-Ottawa Scale (NOS)[14] and the Agency for Healthcare Research and Quality (AHRQ)[15]. Disagreements were resolved by discussion with a third reviewer.

RESULTS

Literature search

A total of 1126 records were found by searching seven electronic databases, and 520 records were deleted because of duplication. A total of 583 studies were excluded based on the title and abstract. The remaining 23 studies were further assessed by reading the full texts. As a result, 12 studies were excluded for various reasons (10 did not match the inclusion criteria, and 2 did not have the full text available). Eleven studies were ultimately included in this review. A detailed flow chart of the literature screening process is shown in Figure 1.

Characteristics of the included studies

Table 1 summarizes the characteristics of the 11 studies included in the systematic review [16-26]. All included studies consisted of 7 cross-sectional studies[16-22], 3 cohort studies[23-25], and 1 case-control study[26], including 912 older adults ranging in age from 65 years to 100 years. Seven studies analyzed the α diversity of the gut microbiota [17-20,22-23, 26], whereas seven studies analyzed the β diversity of the gut microbiota[17,19,21-24,26]. For the outcome measures, two studies reported changes in the gut microbiota at the phylum level[17,22], two at the family level[19,22], eight at the genus level[16-22,25], and five at the species level[16,20,23-25]. Among these included studies, seven studies performed genetic analysis of the gut microbiota via the 16S rRNA method[17-22,26], three studies used the metagenomic sequencing method^[23-25], and one study used the fluorescence in situ hybridization method^[16]. The frailty measures used in the included studies varied widely, with the rockwood frailty index [22,25,26], FI[18], Clinical Frailty Scale [23,24], short physical performance battery[19], Groningen Frailty Indicator[16], Fried's definition[17], Fried's Frailty Phenotype[20] and Frailty Phenotype[21] being used. The non-frail controls were mainly healthy older adults.

Quality assessment

Table 2, 3 and 4 summarizes the study quality of the included studies, as assessed by the AHRQ for 7 cross-sectional studies and by the NOS tool for 3 cohort studies and one case-control study [16-21,23-25]. Of the seven cross-sectional studies, six were of moderate quality [16-21], and one was of high quality [22]. All three cohort studies were of moderate quality^[23-25], and one case-control study was of high quality^[26].

Outcome assessment

Changes in the diversity of the gut microbiota: A total of seven studies analyzed the difference in α diversity of the gut microbiota measured by the Chao index, Simpson index, and Shannon index between frail older adults and non-frail controls[17-20,22,23,26], and two studies reported a significant decrease in frail older adults[18,23]. Seven studies[17,19, 21,23,24,26,27] compared the β diversity measured by principal coordinate analysis in frail older adults with that in controls, and two studies reported significantly greater β diversity in frail than non-frail older adults[21,22].

Changes in the gut microbiota composition: Figure 2 summarizes the changes in the gut microbiota composition at each level in frail older adults compared with non-frail older adults. Two studies reported results at the phylum level [17,22],



Table 1 Main	Table 1 Main characteristics of the included studies in this review											
Ref.	Picca <i>et al</i> [<mark>19</mark>], 2020	Van Tongeren <i>et al</i> [<mark>16</mark>], 2005	Xu e <i>t al</i> [<mark>17</mark>], 2021	Lim e <i>t a</i> 2021	d [18] ,	Zhang e <i>t al</i> [<mark>22</mark>], 2020	Margiotta e <i>t al</i> [<mark>20]</mark> , 2020	Ticinesi <i>et</i> <i>al</i> [26], 2017	Haran <i>et al</i> [<mark>24</mark>], 2018	Haran e <i>t al<mark>[23]</mark>,</i> 2021	Larson e <i>t al</i> [<mark>25</mark>], 2020	Zhang e <i>t al</i> [<mark>21</mark>], 2022
Study design	CSS	CSS	CSS	CSS		CSS	CSS	CCS	CHS	CHS	CHS	CSS
Participants	Samples	35	23	94	176	27	64	76	23	166	47	181
	Age (years)	> 70	70-100	80.7 ± 5.7	74.7	81.63 ± 7.90	≥ 65	83.3 ± 7.5	≥65	86.2 ± 9.1	> 65	≥ 65
	Male/female	20/15	4/19	44/50	54/122	17/10	43/21	39/37	23	30/136	47	72/109
Genetic analysis	16S rRNA V3-V4	Fluorescence in situ hybri	16sRNA V3-V4	165 rRN.	A	16S rRNA	16sRNA V3-V4	16S rRNA	metagenomic sequencing	metagenomic sequencing	metagenomic sequencing	16sRNA V3-V4
		dization										
Frailty diagnosis	Short physical performance battery	Groningen Frailty Indicator	Fried's definition	Frailty ir	ndex	Rockwood Frailty Index	Fried's Frailty Phenotype	Rockwood Index	CFS	CFS	Rockwood Index	Frailty Phenotype
Outcome measurement	α-diversity, β- diversity, family level, genus level	Genus level, species level	α-diversity, β- diversity, phylum level, genus level		ty, genus ecies level	α-diversity, β- diversity, phylum level, family level, genus level	α-diversity, phylum level, genus level	α-diversity, β- diversity	β-diversity, species level	α-diversity, β- diversity, species level	Genus level, species level	β-diversity, phylum level, genus level

CSS: Cross-sectional study: CCS: Case-control study; CHS: Cohort study; CFS: Clinical Frailty Scale.

Table 2 The Agency for Healthcare Research and Quality assessment for the cross-sectional study												
Ref.	Item 1	Item 2	Item 3	Item 4	Item 5	Item 6	Item 7	Item 8	ltem 9	Item 10	Item 11	Scores
Picca <i>et al</i> [19], 2020	Yes	Yes		Yes	Yes			Yes		Yes		6
Van Tongeren <i>et al</i> [<mark>16</mark>], 2005	Yes	Yes		Yes	Yes		Yes			Yes		6
Xu et al[17], 2021	Yes			Yes	Yes		Yes	Yes		Yes		6
Lim <i>et al</i> [18], 2021	Yes	Yes			Yes		Yes	Yes		Yes		6
Zhang <i>et al</i> [22], 2020	Yes	Yes		Yes	Yes	Yes	Yes	Yes		Yes		8
Margiotta et al[<mark>20]</mark> , 2020	Yes	Yes		Yes	Yes		Yes	Yes		Yes		7
Zhang <i>et al</i> [21], 2022	Yes	Yes	Yes		Yes	Yes	Yes	Yes				7

Yes is one point, the total number of stars represents a number of points. Item 1: Define the source of information (survey, record review)? Item 2: Inclusion and exclusion criteria for exposed and unexposed subjects (cases and controls) or refer to previous publications? Item 3: Indicate time period used for identifying patients? Item 4: Indicate whether or not subjects were consecutive if not population-based? Item 5: Indicate if evaluators of subjective components of study were masked to other aspects of the status of the participant? Item 6: Describe any assessments undertaken for quality assurance purposes (*e.g.*, testretest of primary out come measuroments)? Item 7: Explain any patient exclusions from analysis? Item 8: Describe how confounding was assessed and/or controlled? Item 9: If applicable, explain how missing data were handled in the analysis? Item 10: Summarize patient response rates and completeness of data collection? Item 11: Clarify what follow-up, if any, was expected and the percentage of patients for which incomplete data or follow-up was obtained?

Table 3 The Newcastle-Ottawa Scale assessment for the cohort studies

	Selection				Comparability		Exposure			
Ref.	Representativeness of selection of the non- exposed cohort	Selection of the non- exposed cohort	Ascertainment of exposure	Demonstration that outcome of interest was not present at start of study	Comparability of cohort on the basis of the design or analysis	Ascertainment of outcome	Ascertainment of outcome	Was follow-up long enough for outcomes to occur	Adequacy of follow-up of cohorts	Scores
Haran <i>et</i> <i>al</i> [24], 2018	Yes	Yes			Yes	Yes, Yes				5
Haran <i>et</i> <i>al</i> [23], 2021	Yes	Yes				Yes, Yes		Yes	Yes	6
Larson <i>et</i> <i>al</i> [25], 2020	Yes	Yes			Yes	Yes, Yes				5

Yes is one point, the total number of stars represents a number of points.

and all of them reported that frail older adults had a significantly greater relative abundance of the *Actinobacteria* phylum [17,22]. An individual study reported that frail older adults had significantly greater relative abundances of the *Proteobacteria*, *Verrucomicrobia* and *Synergistetes* phyla[17] and significantly lower abundances of the *Firmicutes*[17] and *Fusobacteria* phyla[22].

Two studies reported differences in the gut microbial composition at the family level between frail and non-frail older adults[19,22], with significantly greater abundances of the *Peptostreptococcaceae*[19], *Mogibacteriaceae*, *Bifidobacteriaceae*[19], *Coriobacteriaceae*, *Enterobacteriaceae*, and *Moraxellaceae* families[22] in frail older adults.

Eight studies reported changes in gut microbiota composition at the genus level. Three studies reported a significantly lower abundance of the genus *Prevotella* in frail older adults[16-18]; two studies reported that frail older adults presented significantly lower relative abundances of the genera *Faecalibacterium* and *Roseburia*[17,21]. Several individual studies reported that the relative abundances of *Megamonas*, *Blautia*, *Megasphaera*, *Haemophilus*[17], *Adlercreutzia*, *Clostridium*, *Coprococcus*, *Phascolarctobacterium*, *Turicibacter*[21], *Eubacterium*[19], *Gemella*, *Lachnoanaerobaculum*, *[Eubacterium]_ruminantium_group*, *Tyzzerella*, *Azospira*, *Cloacibacterium* and *EU455341_g*[22] genera, most of which belong to the

	Selection				Comparability	Comparability Exposure						
Ref.	Adequate definition of case	Representativeness of the case	Selection of controls	Definition of controls	Control for important factor	Ascertainment of exposure	Same method of ascertain for cases and controls	Non- responsese rate	Scores			
Ticinesi <i>et al</i> [<mark>26]</mark> , 2017	Yes	Yes	Yes	Yes	Yes	Yes, Yes		Yes	7			

Yes is one point, the total number of stars represents a number of points.

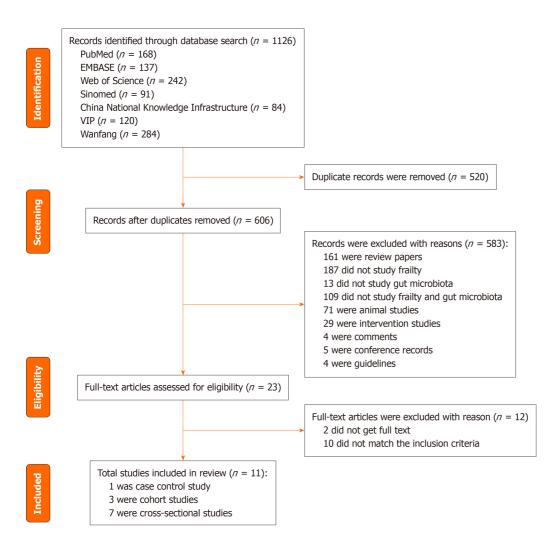


Figure 1 Preferred Items for Systematic Reviews and Meta-Analysis flow diagram of literature search.

Firmicutes phylum, were significantly lower in frail older adults. Moreover, individual studies reported that frail older adults presented increased relative abundances of *Bifidobacterium*[17], *Eggerthella*, *Olsenella*[16], *Atopobium*[22], *Parabacteroides*[17], *Alistipes*[17], *Bacteroides*[18], *Oscillospira*, *Ruminococcus*, *Pyramidobacter* and *Dialister*[19], *Akkermansia* and *Klebsiella*[17], *KF843164_g*, *Pseudoxanthomonas*, *EF434341_g* and *Prevotella_9*[22], and *Oscillospira* and *Coprobacillus*[20]. In addition, the genera of *Lactobacillus* in the included studies were heterogeneous, and three studies[17,20-21] reported a significantly higher relative abundance, but one study[16] reported a significantly lower relative abundance in frail older adults.

At the species level, two studies[16,23] reported that the relative abundance of *Faecalibacterium prausnitzii* was significantly lower in frail than in non-frail older adults. Furthermore, individual studies reported that the abundances of

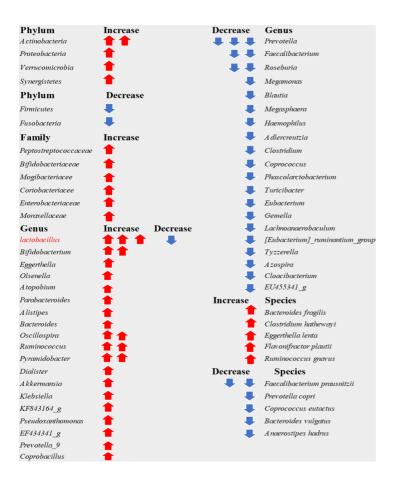


Figure 2 Changes in the gut microbiota composition in older adults with frailty compared to the controls. Red arrows indicate increasing relative abundance; blue arrows indicate decreasing relative abundance in older adults with frailty compared to the controls; One arrow represents one analyzed study.

the Prevotella copri[18], Coprococcus eutactus[18], Bacteroides vulgatus[23] and Anaerostipes hadrus species[23] were significantly lower in frail older adults. However, the abundances of *Bacteroides fragilis*[18], *Clostridium hathewayi*[18], *Egger*thella lenta[20], Flavonifractor plautii[23] and Ruminococcus gnavus[24] were significantly increased in frail older adults.

DISCUSSION

This qualitative systematic review, which included 11 eligible studies with 912 older adults over 65 years of age, investigated the relationships between changes in gut microbiota diversity and composition and frailty in older adults. For gut microbiota diversity, the results, which are based on consistent findings reported by more than two eligible studies, revealed a significant decrease in α diversity and a significant increase in β diversity in frail older adults. In terms of the gut microbiota composition, although there was wide variation in the gut microbiota composition reported in the included studies, the consistent results revealed significant differences in the relative abundance of some gut microbiota compositions at different levels, including phylum, family, genus and species, between frail and non-frail older adults. These findings suggest that changes in the gut microbiota may be associated with frailty in older adults.

An increasing number of studies have reported that altered gut microbiota play an important role as a risk factor in the development of many chronic diseases [28-30]. The gut microbiota in healthy individuals maintains a symbiotic relationship with the host but also triggers some pathological processes and causes the evolution of some diseases if potentially pathogenic bacteria overgrow and alter the diversity and abundance of the gut microbiota[31]. The mechanism is related to a deficiency or excess of metabolites resulting from an imbalance in the gut microbiota, which fundamentally affects the physiological status of the host cells and has direct or indirect toxic effects on hormones and the host organism[32]. The gut microbiota is a highly complex and diverse ecosystem of microorganisms living in the digestive tract, and the balance of beneficial and pathogenic bacteria in the gut microbiome is helpful for maintaining host health and homeostasis[33]. However, both environmental factors and host genetics can affect the homeostatic balance of the gut microbiota and lead to a dysbiotic microbiome configuration by altering the diversity and richness of the gut microbiota[34]. Seven studies included in this review compared the differences in gut microbiota diversity between frail and non-frail older adults. Two of the seven studies reported significantly greater β diversity and significantly lower α diversity in frail than non-frail older adults. These findings suggest a possible separation in gut microbiota diversity in older adults with frailty.

Zaishidena® WJCC | https://www.wjgnet.com

The composition of the gut microbiota in older adults can be altered by the constant influence of external environmental factors, such as diet, medication, physical activity, and social environment. Altered gut microbiota composition has also been shown to play an important role in the development of age-related chronic diseases[35]. The gut microbiome can influence host physical function by regulating nutrient absorption, inflammation, oxidative stress, immune function, and anabolic balance and is associated with the progression of aged-physical frailty [35,36]. Among the relative abundances at the phylum level between frail and non-frail older adults, the current review revealed a significant increase in the Actinobacteria, Proteobacteria, Verrucomicrobia and Synergistetes phyla and a significant decrease in the Firmicutes and Fusobacteria phyla in frail older adults. These phyla are dominant in healthy humans and are pivotal in the maintenance of gut homeostasis[37,38]. There is evidence of positive associations between increased Actinobacteria, Proteobacteria, Verrucomicrobia, and Synergistetes phyla and inflammation-related diseases[39,40]. Conversely, the abundance of the *Firmicutes* phylum was negatively associated with inflammatory responses[41].

With respect to the relative abundance of families between frail and non-frail older adults, the current review revealed that the abundances of Peptostreptococcaceae, Bifidobacteriaceae, Mogibacteriaceae, and Coriobacteriaceae as well as Enterobacteriaceae and Moraxellaceae families were greater in frail than in non-frail older adults. Most of them (Bifidobacteriaceae, Mogibacteriaceae, Coriobacteriaceae, Enterobacteriaceae X and Moraxellaceae) belong to the Actinobacteria or Proteobacteria phylum and have previously been implicated in accelerating the aging process through telomere attrition, cellular senescence, inflammasome activation and impaired mitochondrial function, which have been described as correlates of biological aging or are abundant in elderly individuals[42,43]; furthermore, some of them also seem to be positively correlated with various nutritional and physical features[44,45].

With respect to the genera and species levels, more than one study in the current review reported that the abundances of genera (Roseburia, Faecalibacterium, and Prevotella) and species (Faecalibacterium prausnitzii and Prevotella copri) were significantly lower in frail older adults. The Roseburia and Faecalibacterium genera have anti-inflammatory properties, which are likely mediated by the short-chain fatty acid butyrate[46,47]. Roseburia is a anaerobic bacteria that produces butyrate that metabolizes indigestible carbohydrates to produce short-chain fatty acids (particularly high levels of butyric acid), which maintain intestinal function, immune function, and anti-inflammatory properties[47,48]. Furthermore, a lower Roseburia level was also found to be associated with inflammation-related diseases such as diabetes, obesity, atherosclerosis, and nonalcoholic liver steatohepatitis[47]. Faecalibacterium prausnitzii, which is the only species of the Faecalibacterium genus, is a genus of bacteria that produces butyrate and has anti-inflammatory effects [46]. Moreover, the abundance of Faecalibacterium prausnitzii is obviously lower in patients with gastrointestinal inflammation and ulcerative colitis (UC)[49]. Hedin et al[50] also reported that the abundances of Faecalibacterium prausnitzii and Roseburia are decreased in patients with the inflammatory Crohn's disease. Prevotella and Prevotella copri, which belong to the Prevotellaceae in this review, were reported to be decreased in frail older adults. Prevotella species significantly colonize the human intestine, especially Prevotella copri, which is prevalent in populations fed high-fiber diets and is associated with beneficial outcomes, including reduced visceral fat and improved glucose tolerance[51,52]. Studies have shown that Prevotella copri transplantation may attenuate oxidative stress and blood-brain barrier damage and alleviate motor and cognitive deficits [53]. In addition, some single studies included in this review reported that the abundances of some genera or species, such as Eubacterium, Gemella, Lachnoanaerobaculum, Bacteroides vulgatus, Megasphaera, Haemophilus, Adlercreutzia, Clostridium, Coprococcus, and Blautia, were significantly lower in frail than in non-frail older adults. Most of these microbiomes have been found to be beneficial. For example, several members of the genus Eubacterium can produce butyrate, which plays important roles in the immunomodulation and inhibition of inflammation in the gut microbiome[54]. Eubacterium, Gemella, Lachnoanaerobaculum and Tyzzerella belong to Firmicutes, and an increase in Firmicutes is associated with a reduction in inflammatory responses[41]. Moreover, the current review revealed that some genera or species, including Oscillospira, Ruminococcus, Alistipes, Bacteroides, Bacteroides fragilis, Pyramidobacter, Eggerthella, Olsenella, Atopobium, Parabacteroides, etc., were more enriched in the frail than the non-frail older adults in the individual included studies; some of these genera or species may be related to the pathological mechanisms of frailty. It has been reported that Oscillospira abundance is positively correlated with inflammation in type II diabetes mellitus patients [55] and is associated with a lower body mass index [56]. Ruminococcus gnavus, which is a type of Ruminococcus, is enriched in inflammatory diseases, such as inflammatory bowel disease[57]. Treatment of UC patients with fecal microbiome transplants revealed that disease progression was more likely to recrudesce in those who received high concentrations of *Ruminococcus* donors[58]. Ruminococcus also aggravated amyotrophic lateral sclerosis in mice, leading to further frailty [59]. Alistipes, Bacteroides and Bacteroides vulgatus are commonly associated with chronic intestinal inflammation [41,60]. In this review, the results of Lactobacillus in the included studies were not consistent, and its relative abundance was greater in the frail older adults in three studies but was lower in the frail older adults in one study. The reason may be related to the different diets of the participants in those studies.

This systematic review provides a comprehensive summary and overview of the current research on the gut microbiota and frailty in adults over 60 years of age. By focusing on older adults over 60 years of age, the frailty states assessed by comprehensive tools, and the non-frail control group consisting mainly of community-dwelling healthy older adults or those with comorbidities, the findings of this review may therefore provide informative guidance for the prevention or rehabilitation of frailty in community-dwelling older adults. However, the following limitations should be acknowledged, as they may affect the interpretation of these findings. First, most of the included studies were crosssectional in design, which limits the interpretation of the results regarding causality between changes in the gut microbiota and frailty in older adults. Second, the small sample size and insufficient number of studies may hinder the generalization of the findings of this review. The human gut microbiota is complex and may be influenced by internal and external factors. The small number of included studies may limit the ability to observe the influence of confounding factors such as diet, physical activity, comorbid conditions, and medications. In addition, the variation in the measurement of frailty and the gut microbiota across the included studies also makes accurate assessment or analysis of the



WJCC | https://www.wjgnet.com

associations between the gut microbiota outcomes and frailty difficult.

CONCLUSION

The current review revealed significant changes in the α -diversity and β -diversity and composition of the gut microbiota at the phylum, family, genus, or species level in frail and non-frail older adults aged over 60 years. These changes are commonly reflected by a decrease in the beneficial microbiota (e.g., Faecalibacterium prausnitzii at the species level; Roseburia, Eubacterium, and Faecalibacterium at the genus level) and an increase in the pathogenic microbiota (e.g., Oscillospira, Ruminococcus, Alistipes, Eggerthella, and Bacteroides at the genus level). Future research with large samples and a prospective design is needed to further investigate the impact of specific gut microbiota on frailty in adults over 60 years of age.

FOOTNOTES

Author contributions: Wen NN was responsible for conceptualization, literature screening, data curation, extraction, methodology and writing original draft; Sun LW and Geng Q were responsible for literature screening, data curation, extraction and methodology; Zheng GH was responsible for conceptualization, methodology, project administration, supervision, writing original draft, writing review, and editing; all of the authors read and approved the final version of the manuscript to be published.

Conflict-of-interest statement: All the authors report no relevant conflicts of interest for this article.

PRISMA 2009 Checklist statement: The authors have read the PRISMA 2009 Checklist, and the manuscript was prepared and revised according to the PRISMA 2009 Checklist.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

Country of origin: China

ORCID number: Guo-Hua Zheng 0000-0002-4229-7346.

S-Editor: Luo ML L-Editor: A P-Editor: Xu ZH

REFERENCES

- Walston J, Hadley EC, Ferrucci L, Guralnik JM, Newman AB, Studenski SA, Ershler WB, Harris T, Fried LP. Research agenda for frailty in 1 older adults: toward a better understanding of physiology and etiology: summary from the American Geriatrics Society/National Institute on Aging Research Conference on Frailty in Older Adults. J Am Geriatr Soc 2006; 54: 991-1001 [PMID: 16776798 DOI: 10.1111/j.1532-5415.2006.00745.x]
- 2 Hoogendijk EO, Afilalo J, Ensrud KE, Kowal P, Onder G, Fried LP. Frailty: implications for clinical practice and public health. Lancet 2019; **394**: 1365-1375 [PMID: 31609228 DOI: 10.1016/S0140-6736(19)31786-6]
- Collard RM, Boter H, Schoevers RA, Oude Voshaar RC. Prevalence of frailty in community-dwelling older persons: a systematic review. J 3 Am Geriatr Soc 2012; 60: 1487-1492 [PMID: 22881367 DOI: 10.1111/j.1532-5415.2012.04054.x]
- Rensa R, Setiati S, Laksmi PW, Rinaldi I. Factors Associated with Physical Frailty in Elderly Women with Low Socioeconomic Status in 4 Urban Communities: A Cross-Sectional Study. Acta Med Indones 2019; 51: 220-229 [PMID: 31699945]
- Xie B, Ma C, Chen Y, Wang J. Prevalence and risk factors of the co-occurrence of physical frailty and cognitive impairment in Chinese 5 community-dwelling older adults. Health Soc Care Community 2021; 29: 294-303 [PMID: 32657490 DOI: 10.1111/hsc.13092]
- Wang X, Wu M. Research progress of gut microbiota and frailty syndrome. Open Med (Wars) 2021; 16: 1525-1536 [PMID: 34712824 DOI: 6 10.1515/med-2021-0364]
- 7 Rizka A, Indrarespati A, Dwimartutie N, Muhadi M. Frailty among Older Adults Living in Nursing Homes in Indonesia: Prevalence and Associated Factors. Ann Geriatr Med Res 2021; 25: 93-97 [PMID: 33975423 DOI: 10.4235/agmr.21.0033]
- Chen SY, Wang TY, Zhao C, Wang HJ. Oxidative stress bridges the gut microbiota and the occurrence of frailty syndrome. World J Gastroenterol 2022; 28: 5547-5556 [PMID: 36304085 DOI: 10.3748/wjg.v28.i38.5547]
- Jackson MA, Jeffery IB, Beaumont M, Bell JT, Clark AG, Ley RE, O'Toole PW, Spector TD, Steves CJ. Erratum to: signatures of early frailty 9 in the gut microbiota. Genome Med 2016; 8: 21 [PMID: 26888550 DOI: 10.1186/s13073-016-0275-2]
- Meng X, Zhang G, Cao H, Yu D, Fang X, de Vos WM, Wu H. Gut dysbacteriosis and intestinal disease: mechanism and treatment. J Appl 10 Microbiol 2020; 129: 787-805 [PMID: 32277534 DOI: 10.1111/jam.14661]
- Schirmer M, Garner A, Vlamakis H, Xavier RJ. Microbial genes and pathways in inflammatory bowel disease. Nat Rev Microbiol 2019; 17: 11 497-511 [PMID: 31249397 DOI: 10.1038/s41579-019-0213-6]



- Hogan DB, Maxwell CJ, Afilalo J, Arora RC, Bagshaw SM, Basran J, Bergman H, Bronskill SE, Carter CA, Dixon E, Hemmelgarn B, 12 Madden K, Mitnitski A, Rolfson D, Stelfox HT, Tam-Tham H, Wunsch H. A Scoping Review of Frailty and Acute Care in Middle-Aged and Older Individuals with Recommendations for Future Research. Can Geriatr J 2017; 20: 22-37 [PMID: 28396706 DOI: 10.5770/cgj.20.240]
- 13 Ticinesi A, Nouvenne A, Cerundolo N, Catania P, Prati B, Tana C, Meschi T. Gut Microbiota, Muscle Mass and Function in Aging: A Focus on Physical Frailty and Sarcopenia. Nutrients 2019; 11: 1633 [PMID: 31319564 DOI: 10.3390/nu11071633]
- Stang A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. Eur J 14 *Epidemiol* 2010; **25**: 603-605 [PMID: 20652370 DOI: 10.1007/s10654-010-9491-z]
- Chou R, Baker WL, Bañez LL, Iyer S, Myers ER, Newberry S, Pincock L, Robinson KA, Sardenga L, Sathe N, Springs S, Wilt TJ. Agency for 15 Healthcare Research and Quality Evidence-based Practice Center methods provide guidance on prioritization and selection of harms in systematic reviews. J Clin Epidemiol 2018; 98: 98-104 [PMID: 29409913 DOI: 10.1016/j.jclinepi.2018.01.007]
- van Tongeren SP, Slaets JP, Harmsen HJ, Welling GW. Fecal microbiota composition and frailty. Appl Environ Microbiol 2005; 71: 6438-16 6442 [PMID: 16204576 DOI: 10.1128/AEM.71.10.6438-6442.2005]
- 17 Xu Y, Wang Y, Li H, Dai Y, Chen D, Wang M, Jiang X, Huang Z, Yu H, Huang J, Xiong Z. Altered Fecal Microbiota Composition in Older Adults With Frailty. Front Cell Infect Microbiol 2021; 11: 696186 [PMID: 34485176 DOI: 10.3389/fcimb.2021.696186]
- 18 Lim MY, Hong S, Kim JH, Nam YD. Association Between Gut Microbiome and Frailty in the Older Adult Population in Korea. J Gerontol A Biol Sci Med Sci 2021; 76: 1362-1368 [PMID: 33437992 DOI: 10.1093/gerona/glaa319]
- Picca A, Ponziani FR, Calvani R, Marini F, Biancolillo A, Coelho-Junior HJ, Gervasoni J, Primiano A, Putignani L, Del Chierico F, Reddel S, 19 Gasbarrini A, Landi F, Bernabei R, Marzetti E. Gut Microbial, Inflammatory and Metabolic Signatures in Older People with Physical Frailty and Sarcopenia: Results from the BIOSPHERE Study. Nutrients 2019; 12: 65 [PMID: 31887978 DOI: 10.3390/nu12010065]
- 20 Margiotta E, Miragoli F, Callegari ML, Vettoretti S, Caldiroli L, Meneghini M, Zanoni F, Messa P. Gut microbiota composition and frailty in elderly patients with Chronic Kidney Disease. PLoS One 2020; 15: e0228530 [PMID: 32236095 DOI: 10.1371/journal.pone.0228530]
- 21 Zhang B, Yu J, Yang Y, Wang Z, Lu S, Chen R, Lu W, Yu Z, Hong K. Relationship between frailty degree and intestinal microbiome in elderly patients with hypertension complicated with type 2 diabetes mellitus in community. Shiyong Laonian Yixue 2022; 36: 663-669 [DOI: 10.3969/j.issn.1003-9198.2022.07.005]
- Zhang L, Liao J, Chen Q, Chen M, Kuang Y, Chen L, He W. Characterization of the gut microbiota in frail elderly patients. Aging Clin Exp 22 Res 2020; 32: 2001-2011 [PMID: 31656031 DOI: 10.1007/s40520-019-01385-2]
- Haran JP, Zeamer A, Ward DV, Dutta P, Bucci V, McCormick BA. The Nursing Home Older Adult Gut Microbiome Composition Shows 23 Time-dependent Dysbiosis and Is Influenced by Medication Exposures, Age, Environment, and Frailty. J Gerontol A Biol Sci Med Sci 2021; 76: 1930-1938 [PMID: 34125200 DOI: 10.1093/gerona/glab167]
- 24 Haran JP, Bucci V, Dutta P, Ward D, McCormick B. The nursing home elder microbiome stability and associations with age, frailty, nutrition and physical location. J Med Microbiol 2018; 67: 40-51 [PMID: 29134939 DOI: 10.1099/jmm.0.000640]
- Larson PJ, Oh J, Robison J, Grady J, Kuchel G. 1206. Association of Aging, Frailty and Place of Residence with Skin, Oral and Gut 25 Microbiome Characteristics and Pathogenicity Reservoirs. Open Forum Infectious Diseases 2020; 7: S625-S625 [DOI: 10.1093/ofid/ofaa439.13911
- Ticinesi A, Milani C, Lauretani F, Nouvenne A, Mancabelli L, Lugli GA, Turroni F, Duranti S, Mangifesta M, Viappiani A, Ferrario C, 26 Maggio M, Ventura M, Meschi T. Gut microbiota composition is associated with polypharmacy in elderly hospitalized patients. Sci Rep 2017; 7: 11102 [PMID: 28894183 DOI: 10.1038/s41598-017-10734-y]
- Kang L, Li P, Wang D, Wang T, Hao D, Qu X. Alterations in intestinal microbiota diversity, composition, and function in patients with 27 sarcopenia. Sci Rep 2021; 11: 4628 [PMID: 33633246 DOI: 10.1038/s41598-021-84031-0]
- Wei W, Wang S, Xu C, Zhou X, Lian X, He L, Li K. Gut microbiota, pathogenic proteins and neurodegenerative diseases. Front Microbiol 28 2022; 13: 959856 [PMID: 36466655 DOI: 10.3389/fmicb.2022.959856]
- Tiwari P, Dwivedi R, Bansal M, Tripathi M, Dada R. Role of Gut Microbiota in Neurological Disorders and Its Therapeutic Significance. J 29 Clin Med 2023; 12: 1650 [PMID: 36836185 DOI: 10.3390/jcm12041650]
- Katsimichas T, Theofilis P, Tsioufis K, Tousoulis D. Gut Microbiota and Coronary Artery Disease: Current Therapeutic Perspectives. 30 Metabolites 2023; 13: 256 [PMID: 36837875 DOI: 10.3390/metabo13020256]
- 31 Olvera-Rosales LB, Cruz-Guerrero AE, Ramírez-Moreno E, Quintero-Lira A, Contreras-López E, Jaimez-Ordaz J, Castañeda-Ovando A, Añ orve-Morga J, Calderón-Ramos ZG, Arias-Rico J, González-Olivares LG. Impact of the Gut Microbiota Balance on the Health-Disease Relationship: The Importance of Consuming Probiotics and Prebiotics. Foods 2021; 10: 1261 [PMID: 34199351 DOI: 10.3390/foods10061261]
- Liu J, Tan Y, Cheng H, Zhang D, Feng W, Peng C. Functions of Gut Microbiota Metabolites, Current Status and Future Perspectives. Aging 32 Dis 2022; 13: 1106-1126 [PMID: 35855347 DOI: 10.14336/AD.2022.0104]
- 33 DAS B, Nair GB. Homeostasis and dysbiosis of the gut microbiome in health and disease. J Biosci 2019; 44: 117 [PMID: 31719226]
- Sommer F, Anderson JM, Bharti R, Raes J, Rosenstiel P. The resilience of the intestinal microbiota influences health and disease. Nat Rev 34 Microbiol 2017; 15: 630-638 [PMID: 28626231 DOI: 10.1038/nrmicro.2017.58]
- 35 Coman V, Vodnar DC. Gut microbiota and old age: Modulating factors and interventions for healthy longevity. Exp Gerontol 2020; 141: 111095 [PMID: 32979504 DOI: 10.1016/j.exger.2020.111095]
- Ni Lochlainn M, Bowyer RCE, Steves CJ. Dietary Protein and Muscle in Aging People: The Potential Role of the Gut Microbiome. Nutrients 36 2018; **10**: 929 [PMID: 30036990 DOI: 10.3390/nu10070929]
- Binda C, Lopetuso LR, Rizzatti G, Gibiino G, Cennamo V, Gasbarrini A. Actinobacteria: A relevant minority for the maintenance of gut 37 homeostasis. Dig Liver Dis 2018; 50: 421-428 [PMID: 29567414 DOI: 10.1016/j.dld.2018.02.012]
- Arumugam M, Raes J, Pelletier E, Le Paslier D, Yamada T, Mende DR, Fernandes GR, Tap J, Bruls T, Batto JM, Bertalan M, Borruel N, 38 Casellas F, Fernandez L, Gautier L, Hansen T, Hattori M, Hayashi T, Kleerebezem M, Kurokawa K, Leclerc M, Levenez F, Manichanh C, Nielsen HB, Nielsen T, Pons N, Poulain J, Qin J, Sicheritz-Ponten T, Tims S, Torrents D, Ugarte E, Zoetendal EG, Wang J, Guarner F, Pedersen O, de Vos WM, Brunak S, Doré J; MetaHIT Consortium, Antolín M, Artiguenave F, Blottiere HM, Almeida M, Brechot C, Cara C, Chervaux C, Cultrone A, Delorme C, Denariaz G, Dervyn R, Foerstner KU, Friss C, van de Guchte M, Guedon E, Haimet F, Huber W, van Hylckama-Vlieg J, Jamet A, Juste C, Kaci G, Knol J, Lakhdari O, Layee S, Le Roux K, Maguin E, Mérieux A, Melo Minardi R, M'rini C, Muller J, Oozeer R, Parkhill J, Renault P, Rescigno M, Sanchez N, Sunagawa S, Torrejon A, Turner K, Vandemeulebrouck G, Varela E, Winogradsky Y, Zeller G, Weissenbach J, Ehrlich SD, Bork P. Enterotypes of the human gut microbiome. Nature 2011; 473: 174-180 [PMID:



WJCC | https://www.wjgnet.com

21508958 DOI: 10.1038/nature09944]

- Li Z, Lu G, Li Z, Wu B, Luo E, Qiu X, Guo J, Xia Z, Zheng C, Su Q, Zeng Y, Chan WY, Su X, Cai Q, Xu Y, Chen Y, Wang M, Poon WS, 39 Luo X. Altered Actinobacteria and Firmicutes Phylum Associated Epitopes in Patients With Parkinson's Disease. Front Immunol 2021; 12: 632482 [PMID: 34276644 DOI: 10.3389/fimmu.2021.632482]
- 40 Rodriguez-Palacios A, Cominelli F. Myeloperoxidases and Proteobacteria: Reliable Interspecies Biomarkers to Identify and Monitor Proinflammatory Diets in Humans. Inflamm Bowel Dis 2019; 25: e1-e2 [PMID: 29846576 DOI: 10.1093/ibd/izy197]
- Samaddar A, van Nispen J, Armstrong A, Song E, Voigt M, Murali V, Krebs J, Manithody C, Denton C, Ericsson AC, Jain AK. Lower 41 systemic inflammation is associated with gut firmicutes dominance and reduced liver injury in a novel ambulatory model of parenteral nutrition. Ann Med 2022; 54: 1701-1713 [PMID: 35706376 DOI: 10.1080/07853890.2022.2081871]
- Boopathi S, Kumar RMS, Priya PS, Haridevamuthu B, Nayak SPRR, Chulenbayeva L, Almagul K, Arockiaraj J. Gut Enterobacteriaceae and 42 uraemic toxins - Perpetrators for ageing. Exp Gerontol 2023; 173: 112088 [PMID: 36646294 DOI: 10.1016/j.exger.2023.112088]
- Clavel T, Desmarchelier C, Haller D, Gérard P, Rohn S, Lepage P, Daniel H. Intestinal microbiota in metabolic diseases: from bacterial 43 community structure and functions to species of pathophysiological relevance. Gut Microbes 2014; 5: 544-551 [PMID: 25003516 DOI: 10.4161/gmic.29331]
- Aleman FDD, Valenzano DR. Microbiome evolution during host aging. PLoS Pathog 2019; 15: e1007727 [PMID: 31344129 DOI: 44 10.1371/journal.ppat.1007727]
- 45 Rautio M, Eerola E, Väisänen-Tunkelrott ML, Molitoris D, Lawson P, Collins MD, Jousimies-Somer H. Reclassification of Bacteroides putredinis (Weinberg et al., 1937) in a new genus Alistipes gen. nov., as Alistipes putredinis comb. nov., and description of Alistipes finegoldii sp. nov., from human sources. Syst Appl Microbiol 2003; 26: 182-188 [PMID: 12866844 DOI: 10.1078/072320203322346029]
- Ferreira-Halder CV, Faria AVS, Andrade SS. Action and function of Faecalibacterium prausnitzii in health and disease. Best Pract Res Clin 46 Gastroenterol 2017; 31: 643-648 [PMID: 29566907 DOI: 10.1016/j.bpg.2017.09.011]
- Tamanai-Shacoori Z, Smida I, Bousarghin L, Loreal O, Meuric V, Fong SB, Bonnaure-Mallet M, Jolivet-Gougeon A. Roseburia spp.: a 47 marker of health? Future Microbiol 2017; 12: 157-170 [PMID: 28139139 DOI: 10.2217/fmb-2016-0130]
- Kasahara K, Krautkramer KA, Org E, Romano KA, Kerby RL, Vivas EI, Mehrabian M, Denu JM, Bäckhed F, Lusis AJ, Rey FE. Interactions 48 between Roseburia intestinalis and diet modulate atherogenesis in a murine model. Nat Microbiol 2018; 3: 1461-1471 [PMID: 30397344 DOI: 10.1038/s41564-018-0272-x
- Laursen MF, Laursen RP, Larnkjær A, Mølgaard C, Michaelsen KF, Frøkiær H, Bahl MI, Licht TR. Faecalibacterium Gut Colonization Is 49 Accelerated by Presence of Older Siblings. *mSphere* 2017; 2: e00448-17 [PMID: 29202044 DOI: 10.1128/mSphere.00448-17]
- Hedin CR, McCarthy NE, Louis P, Farquharson FM, McCartney S, Taylor K, Prescott NJ, Murrells T, Stagg AJ, Whelan K, Lindsay JO. 50 Altered intestinal microbiota and blood T cell phenotype are shared by patients with Crohn's disease and their unaffected siblings. Gut 2014; 63: 1578-1586 [PMID: 24398881 DOI: 10.1136/gutjnl-2013-306226]
- Lev RE. Gut microbiota in 2015: Prevotella in the gut: choose carefully. Nat Rev Gastroenterol Hepatol 2016; 13: 69-70 [PMID: 26828918 51 DOI: 10.1038/nrgastro.2016.4]
- 52 Ahmed HS. The Impact of Prevotella on Neurobiology in Aging: Deciphering Dendritic Cell Activity and Inflammatory Dynamics. Mol Neurobiol 2024; 61: 9240-9251 [PMID: 38613648 DOI: 10.1007/s12035-024-04156-x]
- Gu N, Yan J, Tang W, Zhang Z, Wang L, Li Z, Wang Y, Zhu Y, Tang S, Zhong J, Cheng C, Sun X, Huang Z. Prevotella copri transplantation 53 promotes neurorehabilitation in a mouse model of traumatic brain injury. J Neuroinflammation 2024; 21: 147 [PMID: 38835057 DOI: 10.1186/s12974-024-03116-5
- Mukherjee A, Lordan C, Ross RP, Cotter PD. Gut microbes from the phylogenetically diverse genus Eubacterium and their various 54 contributions to gut health. Gut Microbes 2020; 12: 1802866 [PMID: 32835590 DOI: 10.1080/19490976.2020.1802866]
- 55 Zhu Y, Dong L, Huang L, Shi Z, Dong J, Yao Y, Shen R. Effects of oat β-glucan, oat resistant starch, and the whole oat flour on insulin resistance, inflammation, and gut microbiota in high-fat-diet-induced type 2 diabetic rats. JFF 2020; 69: 103939 [DOI: 10.1016/j.jff.2020.103939
- Tims S, Derom C, Jonkers DM, Vlietinck R, Saris WH, Kleerebezem M, de Vos WM, Zoetendal EG. Microbiota conservation and BMI 56 signatures in adult monozygotic twins. ISME J 2013; 7: 707-717 [PMID: 23190729 DOI: 10.1038/ismej.2012.146]
- Hall AB, Yassour M, Sauk J, Garner A, Jiang X, Arthur T, Lagoudas GK, Vatanen T, Fornelos N, Wilson R, Bertha M, Cohen M, Garber J, 57 Khalili H, Gevers D, Ananthakrishnan AN, Kugathasan S, Lander ES, Blainey P, Vlamakis H, Xavier RJ, Huttenhower C. A novel Ruminococcus gnavus clade enriched in inflammatory bowel disease patients. Genome Med 2017; 9: 103 [PMID: 29183332 DOI: 10.1186/s13073-017-0490-5]
- Fuentes S, Rossen NG, van der Spek MJ, Hartman JH, Huuskonen L, Korpela K, Salojärvi J, Aalvink S, de Vos WM, D'Haens GR, Zoetendal 58 EG, Ponsioen CY. Microbial shifts and signatures of long-term remission in ulcerative colitis after faecal microbiota transplantation. ISME J 2017; 11: 1877-1889 [PMID: 28398347 DOI: 10.1038/ismej.2017.44]
- Blacher E, Bashiardes S, Shapiro H, Rothschild D, Mor U, Dori-Bachash M, Kleimeyer C, Moresi C, Harnik Y, Zur M, Zabari M, Brik RB, 59 Kviatcovsky D, Zmora N, Cohen Y, Bar N, Levi I, Amar N, Mehlman T, Brandis A, Biton I, Kuperman Y, Tsoory M, Alfahel L, Harmelin A, Schwartz M, Israelson A, Arike L, Johansson MEV, Hansson GC, Gotkine M, Segal E, Elinav E. Potential roles of gut microbiome and metabolites in modulating ALS in mice. Nature 2019; 572: 474-480 [PMID: 31330533 DOI: 10.1038/s41586-019-1443-5]
- Parker BJ, Wearsch PA, Veloo ACM, Rodriguez-Palacios A. The Genus Alistipes: Gut Bacteria With Emerging Implications to 60 Inflammation, Cancer, and Mental Health. Front Immunol 2020; 11: 906 [PMID: 32582143 DOI: 10.3389/fimmu.2020.00906]



WJCC | https://www.wjgnet.com



Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: office@baishideng.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

