

Effects of p27^{Kip1}- and p53- shRNAs on kanamycin damaged mouse cochlea

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Abstract

AIM: To study the effects of adeno-associated virus (AAV) delivered short hairpin RNAs (shRNAs) on adult CD-1 mouse cochlea damaged by aminoglycoside antibiotic kanamycin.

METHODS: Three different shRNAs were designed (p27^{Kip1}, p53 and p27^{Kip1}+p53) and tested in COS cells. A total of 20 adult CD-1 mice were used in the experiment. Mice were divided into five different groups (four

animals/group) depending on the AAV-shRNA construct they received and whether they received kanamycin or not. Saline and AAV-EGFP injected animals were used as controls. All constructs were injected through the round window membrane (RWM) into the cochlea. Cochleae were harvested after 1 mo. Apoptosis was detected with Tunel labeling from paraffin-embedded cochlear tissue sections.

RESULTS: AAV2/2-p27^{Kip1}-shRNA and AAV2/2-p53-shRNA were tested in COS cells. Western blotting analysis confirmed that both constructs silenced their target genes effectively in the cell culture. AAV2/2-shRNA constructs were injected into the cochlea of CD-1 mice through the intact RWM. Cotransduction of individual AAV2/2-shRNAs with AAV2/2-EGFP resulted in EGFP expression in the organ of Corti. Kanamycin treatment had no effect on the expression pattern of the EGFP. AAV2/2-shRNA treated mice (either with p53 or p27^{Kip1} and p53 together) showed fewer apoptotic hair cells in the cochlea than the control group ($P < 0.05$; AAV2/2-p53-shRNA vs saline $P = 0.00014$; AAV2/2-p27+p53-shRNA vs saline $P = 0.0011$). AAV2/2-p27-shRNA injected cochleae showed no significant difference in the number of apoptotic cells when compared to the saline injected cochleae.

CONCLUSION: Silencing of p53 protein in the kanamycin treated ears may decrease cell death in the organ of Corti.

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Key words: Inner ear; Adeno-associated virus; Short hairpin RNA; p27; p53; Kanamycin; Apoptosis

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INTRODUCTION

Specific genes can be efficiently silenced with RNA interference (RNAi). Fire *et al*^[1] showed in 1998 that double-stranded RNA (dsRNA) can be used in post-transcriptional gene silencing in animals. They also noticed that dsRNA was more effective at producing gene silencing when compared to single-stranded antisense or sense RNA. RNAi can be accomplished by introducing chemically synthesized small interfering RNAs (siRNAs) or vector-based short hairpin RNAs (shRNAs) into living cells^[2,3]. Adenovirus and adeno-associated virus (AAV) vector based shRNAs have been shown to induce RNAi in neural tissues *in vivo*^[4]. shRNAs can be delivered into the inner ear by cationic liposomes or different viral vectors (lentivirus, adenovirus or AAV). Transduction efficiency with cationic liposomes is known to be low and the effect is transient^[5,6], long-term transfection is possible with adeno-associated and lentiviral vectors^[7-9]. Long-term, stable expression of shRNAs can be achieved by using RNA polymerase III promoter U6^[10,11]. The unique sequence specificity of shRNAs makes them suitable to be applied to the inner ear diseases caused by gain of function mutations^[12].

Hair cells (HC) in the organ of Corti are terminally differentiated and maintenance of the post-mitotic state is essential for HC survival. Almost all cell divisions in the developing auditory epithelium of mammals and birds stop before birth. Damaged auditory epithelium of birds has an innate capacity to evoke proliferation of supporting cells (SC), which leads to regeneration of new HC^[13]. HCs in the utricular sensory epithelia of adult guinea pigs and adult humans can proliferate after aminoglycoside damage when maintained *in vitro*. Also, HCs of vestibular sensory epithelia of young guinea pigs are able to regenerate after *in vivo* administration of aminoglycoside antibiotics^[14]. Despite these observations, HC and SC of the mammalian organ of Corti are quiescent. The majority of mouse cochlear cells become post-mitotic by E14.5^[15] and the correct cell cycle withdrawal requires cyclin-dependent kinase inhibitor (CKI) p27^{Kip1} (also known as Cdkn1b). In newborn and adult mice, p27^{Kip1} is expressed in cochlear SCs but not in HCs. In SCs, p27^{Kip1} acts as a negative regulator of the G1-S transition in the cell cycle^[16,18]. The deletion of p27^{Kip1} affects the morphology of cochlear sensory cells and all p27^{Kip1} knockout mice have severe hearing impairment^[17]. How-

ever, p27^{Kip1} deficient mice have mitotically active cells in postnatal organ of Corti^[16,17]. p27^{Kip1} knockout mice also have supernumerary HC in both outer HC (OHC) and inner HC (IHC) rows^[16]. While p27^{Kip1} is required for the maintenance of the post-mitotic state of the SCs, other CKIs such as p19^{Ink4d}, p21^{Cip1} and the retinoblastoma tumor suppressor protein (pRb) operate in HCs^[19-21]. pRb is the primary protein involved in cell cycle regulation in HCs and its function is to repress transcription of genes required for G1-S transition. It is known that inactivation of pRb family members in neurons make the cells re-enter the S-phase. This results in abnormal DNA replication, which is followed by cell death. p53 is a tumor suppressor protein (encoded by the *TP53* gene) which has a major role in DNA damage-induced cell death. When a cell's DNA repair system fails, p53 is phosphorylated and the phosphorylated p53 upregulates Bax, a Bcl-2 family member which takes part in cell death regulation. p53 is also involved in initiating cell death in cochlear and vestibular HC^[22]. For now, the mechanisms behind the death of neurons and other post-mitotic cells after cell cycle re-entry are poorly known. A recent study by Sulg *et al*^[23] revealed that misexpression of human papillomavirus-16 E7 oncogene triggered a forced S-phase entry in HCs in explant cultures. They also discovered that DNA damage in OHC following the forced cell cycle entry activated p53 and led to rapid death of the cells. However, utricular cells showed long-term survival after p53 induction and some of the cells progressed into mitosis.

Damage in the mammalian HC is irreversible and leads inevitably to sensorineural hearing loss. Aminoglycoside antibiotics are known to damage primarily the OHCs of the organ of Corti. Aminoglycosides enter the HCs through mechano-electrical transducer channels. When aminoglycosides are inside HCs, they form aminoglycoside-iron complexes which results in the formation of reactive oxygen species (ROS)^[24]. ROS are believed to promote apoptotic cell death. Apoptosis of OHCs begins at the basal region of the cochlea and spreads towards the middle and apical zones. IHC and SC are usually less affected^[25]. It is known that deletion of the p53 offers significant protection against cisplatin-induced HC loss^[22]. Kaur *et al*^[26] found that siRNA against transient receptor potential vanilloidin 1 also offers protection against HC loss caused by cisplatin.

Here we have performed AAV2/2 vector mediated delivery of p27^{Kip1} and p53 shRNAs into adult mouse cochlea damaged by aminoglycoside antibiotic kanamycin. We investigated the effects of these constructs on the damaged organ of Corti.

MATERIALS AND METHODS

ShRNA constructs and cell culture testing

For gene-silencing experiments to produce shRNAs, four different pairs of p27^{Kip1} sequences were designed from Genbank BC014296 and NM_009875 sequences and synthesized (Oligomer, Finland): (1) p27^{Kip1} shRNA

1 lower strand: 5'-AATTAAAACTTACCGCCACAC-GAGTCTCGGAGCCGCGGATATCACTCGCG-GCTCCGAGACTCGCGCGGCGGCAAGGGCC-3' and upper strand: 5'-CTTGCCCGCCGCGGAGTCTC-GGAGCCGCGAGTGATATCCGCGGCTCCGAGA-CTCGTGTGGCGGTAAGTTTTT-3'; (2) p27^{Kip1} shRNA 2 lower strand: 5'-AATTAAAAACAACGACCACCC-GAGCCGAGTAGCGCGGATATCACTCGCGC-TACTGCGGCTCGGCGGTCGCTCGGGCC-3' and upper strand: CGAGCGACCGCCGAGCCG-CAGTAGCGGAGTGATATCCGCGTACTGCG-GCTCGGGTGGTCGTTTTGTTTTT-3'; (3) p27^{Kip1} shRNA 3 lower strand: 5'-AATTAAAAACCTCTGC-CATTCGTATCTGCCCTCCAGGGATATCACTCCT-GGAGGGCAGATACGAATGGCAGGAGGGGCC-3' and upper strand: CCTCTGCCATTTCGTATCT-GCCCTCCAGGAGTGATATCCCTGGAGGGCA-GATACGAATGGCAGGAGGTTTTT-3'; and (4) p27^{Kip1} shRNA 4 lower strand: 5'-AATTA-AAAAGTCGCAGAACTTCGAAGAGGATAT-CACTCTCTCGAAGTTCTGCGACGGCC-3' and upper strand: GTCGCAGAACTTCGAAGAGAGT-GATATCCCTCTCGAAGTTCTGCGACTTTTT-3'.

The pairs were annealed and the generated double-stranded DNA fragments were cloned into the pSilencer 1.0-U6 plasmid (Ambion, United States) between *ApaI* and *EcoRI* under the U6-promoter. The silencing effect of the different pairs was tested by transfecting 1 µg of shRNA-plasmid into COS-cells in culture with Lipofectamine or Lipofectamine 2000 (Invitrogen, United States) reagents according to the manufacturer's instructions. Since proliferating COS cells do not express p27^{Kip1}, we cotransfected 1 µg of p27^{Kip1} cDNA expressing vector together with the shRNA plasmids. Bluescript plasmid (Stratagene, United States) was used as a control. Expression levels of p27^{Kip1} in COS cells were analyzed from 10 µg of protein from cell lysates with 12% SDS-PAGE acrylamide gel-electrophoresis, Western blotting using Hybond-C Extra membrane (Pharmacia, United States), rabbit polyclonal anti-p27^{Kip1} antibody (1:1000; Santa Cruz, United States) and Anti-rabbit Ig-HRP visualized by ECL Western blotting substrate (Pierce, United States). The best silencing sequences (shRNAs 2 and 3) with their respective U6 promoters were digested out from the pSilencer 1.0-U6 with *BamHI* and cloned next to each other into the AAV-vector (serotype 2, AAV2/2) in front of the woodchuck hepatitis virus post-transcriptional element (WPRE)^[27] (Figure 1). The CMV promoter present in the AAV-CMV-WPRE construct^[27] was removed with *EcoRI* and *XhoI* and uncoding stuffer DNA was cloned instead to keep the viral DNA size optimal (Figure 1).

The following p53 shRNA sequences were designed from GenBank: AF051368 and synthesized (Oligomer): p53 lower strand: 5'-AATTAAAAAGGCCCAAGT-GAAGCCCTCCGAGTGTGATATCACTTAA-CACTCGAAGGGCTCCACTTAGACCGGCC-3' and upper strand: 5'-GGTCTAAGTGGAGCCCTTC-GAGTGTAAAGTGATATCTGACACTCG-GAGGGCTTCACTTGGGCCTTTTT-3'. As above, the annealed fragment was cloned in pSilencer 1.0-U6 and

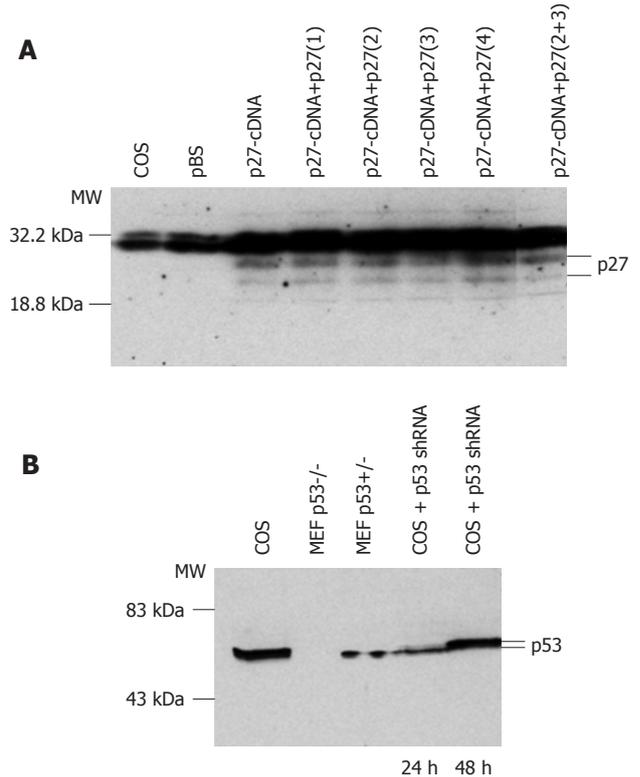


Figure 1 Short hairpin RNA testing in COS cells. A: Western blot analysis with anti-p27^{Kip1} antibody. Proteins were extracted from untransfected COS cells and COS cells transfected with Bluescript (pBS), p27^{Kip1} cDNA (p27-cDNA) or p27^{Kip1} cDNA together with 4 different p27^{Kip1} short hairpin RNA (shRNA) constructs, p27(1-4). p27^{Kip1} specific bands are marked on the right side. A decrease in p27^{Kip1} level was observed with p27(2), p27(3) and p27(2+3) after 24 h of transfection; B: Western blot analysis with anti-p53 antibody. Proteins were extracted from untransfected COS cells (COS), p53 null cells (p53^{-/-}), p53 heterozygous null cells (p53^{+/-}), COS cells transfected with p53 shRNA. Cells were collected 24 h or 48 h after transfection. p53 specific bands are marked on the right side. A decrease in p53 level was detected after 24 h of transfection. Kaleidoscope molecular weight (MW) standards are marked on the left side. MEF: Mouse embryonic fibroblasts.

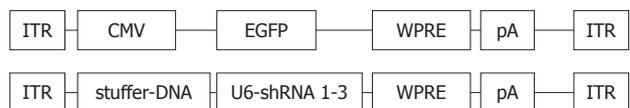


Figure 2 Schematic representation of used adeno-associated virus constructs. The constructs are not drawn in scale. ITR: Adeno-associated virus inverted terminal repeats; CMV: Human cytomegalovirus promoter; U6: U6-promoter; EGFP: Enhanced green fluorescent protein coding sequence; shRNA: Small hairpin RNA; 1: shp27Kip1(2)/shp27Kip1(3); 2: shp53; 3: shp53/shp27Kip1(2)/shp27Kip1(3); WPRE: Woodchuck hepatitis virus post-transcriptional regulatory element; pA: SV40 polyadenylation signal.

tested in COS cells. As controls for Western blotting, mouse embryonic fibroblasts (MEF) from p53 targeted embryos (p53^{-/-} and p53^{+/-}; Jackson laboratory, United States) were grown. Western blotting analysis was performed as above with anti-p53 antibody (1:300; sc-6243 Santa Cruz).

The U6-p53 fragment from pSilencer 1.0-U6 was cloned into the AAV-vector as above either alone or together with the p27^{Kip1} shRNAs 2 and 3 (Figure 1).

Table 1 Grouping of injected mice (*n* = 20)

Group	<i>n</i>	Construct	Kanamycin treatment
1	4	AAV-p27-shRNA	Yes
2	4	AAV-p53-shRNA	Yes
3	4	AAV-p27+p53-shRNA	Yes
4	4	Saline	Yes
5	4	AAV-EGFP	No

AAV: Adeno-associated virus; shRNA: Short hairpin RNA.

Virus production and transduction

The production of recombinant AAV vectors (Figure 2) was described earlier by Paterna *et al.*^[27] in 2000.

Animal surgery and transduction of mice *in vivo* with AAV-shRNA constructs

Twenty white, 11 wk to 13 wk old male CD-1 mice (weight approx. 30 g, Harlan, Netherlands) were used. The mice were divided into groups (Table 1) according to which AAV2/2-shRNA complex they received: p27^{Kip1} (AAV-p27-shRNA), p53 (AAV-p53-shRNA), p27^{Kip1}+p53 (AAV-p27+p53-shRNA). These groups were also transduced with the AAV2/2-EGFP. The fourth group received only saline injections. These four groups (16 mice) were pretreated with kanamycin injections 15 d prior to the surgery (Table 1). Each animal received 0.6 mL of kanamycin sulfate (0.02 mL/g daily, 2 × 0.3 mL/d intraperitoneally, i.p., 400 mg/kg, Sigma-Aldrich) daily. The fifth group received only AAV2/2-EGFP injections without kanamycin pretreatment. The animals were anesthetized with an ip injection of ketamine and midazolam (0.05 mg/10 g body weight). Exposure of the cranial base and middle ear bulla was performed through a ventral paramedian incision as described previously by Jero *et al.*^[28] in 2001. AAV-shRNA together with AAV-EGFP or saline was introduced into the inner ear of the mice by injecting directly through the round window membrane (RWM). Saline (1 μL) or AAV-shRNA and AAV-EGFP (0.5 + 0.5 μL) was injected into the inner ear with a glass capillary (inner diameter 0.5 mm, SM100F-15, Harvard apparatus, tip diameter 50 μm) with the help of a microinjector (Narishige, Japan) and a preparation microscope (Olympus SZX12). Virus titer was 1 × 10¹² pfu/mL. After the injection, the RWM was sealed with a small piece of thin fascia and the incision was closed in layers. The total operating time was approximately 15-20 min.

Ethics

Animal care and studies were approved by the Local Ethics Committee of Animal Experiments, University of Helsinki, and by the Provincial State Office of Southern Finland. Animal experiments were conducted in accordance with the European Convention guidelines.

Sample preparation, immunohistochemistry and statistical analysis

Animals were sacrificed 1 mo after the surgery with an ip overdose of sodium pentobarbital (0.3 mg/10 g, Me-

bunat, Orion, Finland) and perfused transcardially. Inner ears were harvested and postfixed o/n in 4% PFA and decalcified for 14 d in 0.5 mol/L EDTA. The cochleae were mounted in paraffin.

Paraffin-embedded cochlear tissue sections (5 μm) were dewaxed, mounted in fluorescent Vectashield Mounting Medium (Vector Laboratories, United States) under glass coverslips and analyzed for EGFP expression with Axioplan 2 fluorescence microscope (Zeiss).

Apoptosis in the organ of Corti was evaluated with Tunel staining (Roche Applied Science, Germany). Stained slides were mounted in Vectashield Mounting Medium, fluorescence (Vector Laboratories) under glass coverslips. All stained slides were analyzed with Axioplan 2 microscope (Zeiss).

Four animals per group and twenty sections per animal were studied. Results were evaluated statistically using Student's *t* test. Statistical significance was set at *P* < 0.05.

RESULTS

Testing of shRNA constructs in COS cells

The different pSilencer 1.0-U6 shRNA constructs were transfected into COS cells with Lipofectamine reagents. Cells were collected and proteins extracted after 24 h or 48 h of incubation. To test the p27^{Kip1} constructs, p27^{Kip1} cDNA was also cotransfected into COS cells because no p27^{Kip1} expression was detected in dividing COS cells (Figure 1A). Western blotting analysis with anti-p27^{Kip1} antibody showed that the best silencing was produced with two p27^{Kip1}-shRNAs 2 and 3. These two shRNAs were first cloned together into the pSilencer 1.0-U6 vector and tested in COS cells (Figure 1A). After successful testing in COS cells, the two p27^{Kip1}-shRNAs were cloned into the AAV2/2 vector^[18] to give rise to the AAV-p27-shRNA (Figure 1A).

The p53-shRNA was tested in transfections into COS cells which express p53 (Figure 1B). In Western blotting analysis, proteins were also extracted from MEFs isolated from gene targeted p53 null embryos (p53^{-/-}) and heterozygous embryos (p53^{+/-}). As expected, no p53 was detected in p53^{-/-} MEFs, whereas p53^{+/-} MEFs showed detectable levels of p53 expression (Figure 1B). The p53 expression in the COS cells was clearly decreased after 24 h of the transfection, whereas after 48 h the level appeared nearly normal (Figure 1B). The p53-shRNA fragment was cloned into the AAV2/2 vector either alone to give rise to the AAV-p53-shRNA or together with the two p27^{Kip1}-shRNAs to give rise to AAV-p27+p53-shRNA.

AAV-shRNAs in mouse cochlea

The AAV2/2-shRNA vectors were injected together with AAV2/2-EGFP into kanamycin pretreated mouse cochlea. After a month, EGFP expression was detected in the cochlea of all injected mice (Figure 3A and B). Expression was limited to the organ of Corti, histologically in both IHC and OHC and in SC, and was detected in all cochlear turns. The expression was, however, most

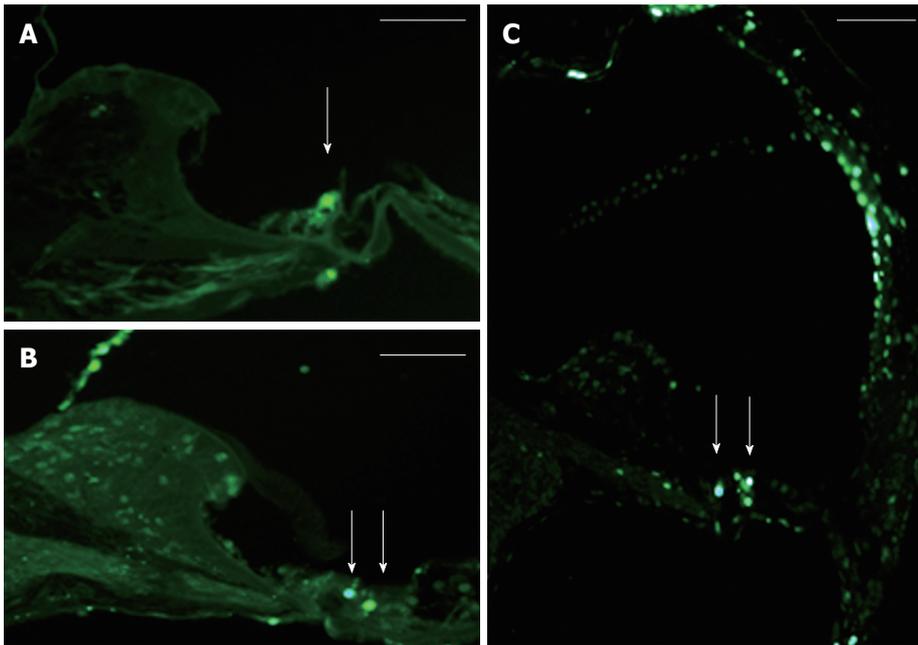


Figure 3 Adeno-associated virus-short hairpin RNAs in mouse cochlea. A: EGFP expression in mouse cochlea organ of Corti cotransduced with adeno-associated virus (AAV)-EGFP and AAV-short hairpin RNA (shRNA) (p27^{Kip1}+p53). Arrow shows EGFP-positive cell; FITC filter, Scale bar 200 μm; B: Uninjected control ear from the AAV-shRNA (p27^{Kip1}+p53) injected animal; Arrows point to apoptotic cells in organ of Corti, Scale bar 200 μm; C: NaCl injected animal's ear; Arrows point to apoptotic cells in organ of Corti. Scale bar 200 μm.

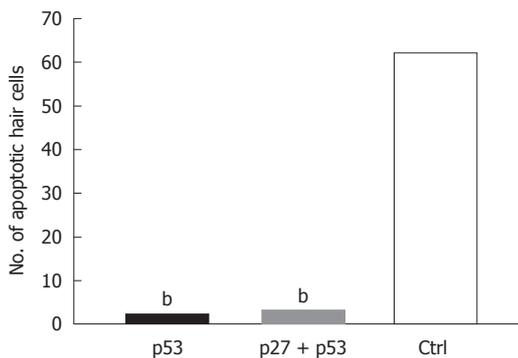


Figure 4 Number of apoptotic cells in the mouse cochlea after kanamycin treatment and injections with adeno-associated virus-p53-short hairpin RNA and adeno-associated virus-p27 + p53-short hairpin RNA constructs. Differences in the level of apoptosis between adeno-associated virus (AAV)-short hairpin RNA (shRNA) and saline injected cochleae were statistically significant, ^a $P < 0.001$. AAV-p53-shRNA vs saline $P = 0.00014$; AAV-p27 + p53-shRNA vs saline $P = 0.0011$.

intense at the site of injection. One group was injected with AAV2/2-EGFP but did not receive kanamycin injections prior the injections. The expression of EGFP was similar to the other injected mice, so kanamycin had no effect on the expression pattern of the EGFP (data not shown). Apoptosis in the inner ear due to aminoglycoside damage was demonstrated by Tunel staining. Tunel stained cells were detected in all cochlear turns (data not shown). AAV2/2-shRNA injected inner ears showed less apoptotic HC when compared to the contralateral control ear and to the inner ears of saline injected control animals (treated with kanamycin) (Figure 3A-C). AAV-p53-shRNA and AAV-p27+p53-shRNA injected co-

chleae showed significantly less apoptotic cells compared to saline injected cochleae, $P < 0.05$ (AAV-p53-shRNA *vs* saline $P = 0.00014$, AAV-p27+p53-shRNA *vs* saline $P = 0.0011$) (Figure 4, $P < 0.001$). AAV-p27-shRNA injected cochleae showed no significant difference compared to the saline injected cochleae.

DISCUSSION

In this study, we evaluated the effects of three AAV2/2-mediated shRNAs on mouse cochlea damaged by aminoglycoside antibiotic kanamycin. AAV2/2 was selected as the carrier for shRNAs based AAV vectors properties. AAV vectors have an ability to transduce non-dividing cells. AAV serotype 2/2 can transduce cells of the organ of Corti, especially both IHC and OHC but also the SC^[8]. AAV vectors are generally safe and cause only minimal damage in the inner ear structures^[8]. The only limiting factor is the small packaging size of the virus. Approximately only 4.5 kb of foreign DNA can be packaged into the virus capsid. We chose to use AAV2/2 in our study based on our own previous data^[8], which had shown that AAV2/2 is suitable for inner ear gene therapy applications. In contrast to adenovirus, AAV generates only minimal immune response and it is primarily a humoral response^[29,30]. **We chose to do the injections through the RWM in order to be sure that the vector constructs were delivered to scala tympani.** The surgery can cause damage to the inner ear and affect hearing, even although a previous study suggests that the direct microinjection through RWM causes only minimal damage and minor changes in the cochlear cytoarchitecture^[28].

Possible damage to hearing was not evaluated in this study.

Created shRNAs were first tested in a COS cell culture. Both shRNAs (p27^{Kip1} and p53) were successful in silencing their target genes in COS cells. **These results** suggest that the p27^{Kip1}-shRNAs 2 and 3 can be used in cells in order to silence p27^{Kip1}, and that the p53-shRNA can be used for silencing experiments in cells. Results from cell culture experiments led us to believe that p27^{Kip1}-and p53-shRNAs may also be used in silencing experiments in the cells of auditory epithelium. We hypothesized that silencing of p27^{Kip1} and p53 could improve cell survival in cochlea. The HC loss caused by kanamycin was evaluated with Tunel staining, which specifically stains apoptotic cells at various stages. Kanamycin injections led to apoptosis in cochlear auditory epithelium. Apoptotic HC were seen throughout the cochlea. Kanamycin has been used with CBA, C57Bl6 and BALB mouse strains^[51]. In this study, we used CD-1 mouse strain. With the CD-1 strain, kanamycin caused both IHC and OHC loss and was consistent with earlier studies with other mouse strains^[51]. **AAV-p53- and AAV-p27^{Kip1}+p53-shRNA treated ears showed less apoptotic HC when compared to the saline injected control ears. AAV-p27-shRNA alone had no significant effect to the number of apoptotic cells in the organ of Corti when compared to contralateral control ear or saline injected (kanamycin treated) ear. The combination of p27^{Kip1} and p53 shRNAs did not decrease the number of apoptotic cells compared to AAV-p53-shRNA alone. This suggests that silencing of p53 in the kanamycin treated ear may decrease cell death in cochlear auditory epithelium.**

The influence of shRNAs to the function of auditory epithelium was not studied. It is currently not known if these shRNAs have an effect on hearing *in vivo*. This needs to be studied in future.

In conclusion, shRNAs can be used in silencing genes in the cell of inner ear. Silencing of p53 in the kanamycin treated ears seems to decrease cell death in the organ of Corti.

ACKNOWLEDGMENTS

We are grateful to Raija Savolainen for the help with DNA-constructs, Western blottings and cell culture.

COMMENTS

Background

Hair cells (HC) in the organ of Corti are terminally differentiated and maintenance of the post-mitotic state is essential for HC survival. **Almost all cell divisions** in the developing auditory epithelium of mammals and birds stop before birth. The majority of mouse cochlear cells become post-mitotic by embryonic day 14.5 and the correct cell cycle withdrawal requires cyclin-dependent kinase inhibitor p27^{Kip1} (also known as Cdkn1b). p53 is a tumor suppressor protein which has a major role in DNA damage-induced cell death. p53 is also involved in initiating cell death in cochlear and vestibular HC. For now, the mechanisms behind the death of neurons and other post-mitotic cells after cell cycle re-entry are poorly known. Damage in the mammalian HC is irreversible and leads inevitably to sensorineural hearing loss. Aminoglycoside antibiotics are known

to damage primarily the outer HC of the organ of Corti. It is known that deletion of the p53 offers significant protection against cisplatin (chemotherapy drug) induced HC loss.

Research frontiers

HC and supporting cells of the mammalian organ of Corti are quiescent. Damage in these cells is irreversible and leads inevitably to sensorineural hearing loss. The main focus of the research is in finding a way to save the HC from trauma, make HC enter the cell cycle again, regenerate HC after cell death, and find a suitable carrier for inner ear gene transfer applications. Cochlear gene therapy studies are, at the moment, focused on the use of different types of vectors (viral and nonviral) as transgene carriers. Each viral vector possesses its own characteristics that limit their use in different applications. At the moment, there is no single vector that would be suitable for each cochlear gene therapy application.

Innovations and breakthroughs

In this study, we evaluated the effects of three adeno-associated virus (AAV)-short hairpin RNAs (shRNAs) on mouse cochlea damaged by aminoglycoside antibiotic kanamycin. Created shRNAs were first tested in a COS cell culture. Both shRNAs (p27^{Kip1} and p53) were successful in silencing their target genes in COS cells. **These results suggest that both shRNA can be used for silencing experiments in cells.** Results from cell culture experiments led us to believe that p27^{Kip1}-and p53-shRNAs may also be used to silence the expression of these genes in the cells of auditory epithelium. The study hypothesized that the silencing of p53 could improve cell survival in cochlea. It found that **AAV-p53- and AAV-p27^{Kip1}+p53-shRNA treated ears showed less apoptotic HC when compared to the saline injected control ears.** So, shRNAs can be used in silencing genes in the cell of inner ear. Silencing of p53 in the kanamycin treated ears seems to decrease cell death in the organ of Corti.

Applications

This study suggests that AAV-shRNAs are promising tools for promoting cell survival in damaged cochlea.

Terminology

shRNA is a short sequence of RNA that makes a tight hairpin turn that can be used to silence gene expression *via* RNA interference. AAV: AAV belongs to the genus *Dependovirus*, which in turn belongs to the family *Parvoviridae*. It is a small, replication-defective, non-enveloped virus. AAV can infect both dividing and non-dividing cells and may incorporate its genome into that of the host cell. These features make AAV a good candidate for creating viral vectors for gene therapy. p27^{Kip1} is an enzyme that in humans encodes a protein which belongs to the *Cip/Kip* family of cyclin dependent kinase (Cdk) inhibitor proteins. These proteins control the cell cycle re-entry. It is often called a cell cycle inhibitor protein because its major function is to stop or slow down the cell division cycle. p53 is a tumor suppressor protein. p53 regulates the cell cycle and functions as a tumor suppressor that is involved in preventing cancer. p53 has a role in conserving stability by preventing genome mutation. Kanamycin sulfate: is an aminoglycoside antibiotic, available in oral, intravenous and intramuscular forms. It is used to treat a wide variety of infections. Serious side effects include tinnitus or hearing loss, toxicity to kidneys and allergic reactions to the drug.

Peer review

The paper is well written and well designed. **This study describes a very interesting topic.** The effects of kanamycin on the cochlea have been well studied in the current literature, as well as the role of p53 and p27.

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